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UTILITY PATENT APPLICATION **TRANSMITTAL**

(Only for new nonprovisional applications under 37 CFR 1.53(b))

Attomey Docket No.	P05562US00					
First Inventor	FARID, Abdol Hossain, et al.					
Title	INSULIN-LIKE GROWTH FACTOR-1					
Express Mail Label No.	EV 330573076 US					

APPLICATION ELEMENTS See MPEP chapter 600 concerning utility patent application contents.	ADDRESS TO: Mall Stop Patent Application Commissioner for Patents P.O. Box 1450 Alexendria VA 22313-1450								
1. Fee Transmittal Form (e.g., PTO/SB/17) (Submit an original and a duplicate for fee processing) Applicant claims small entity status. See 37 CFR 1.27. 3. Specification [Total Pages	7. CD-ROM or CD-R in duplicate, large table or Computer Program (Appendix) 8. Nucleotide and/or Amino Acid Sequence Submission (if applicable, all necessary) a. Computer Reader Form (CRF) b. Specification Sequence Listing on: i. CD-ROM or CD-R (2 copies); or ii. Paper c. Statements verifying identity of above copies ACCOMPANYING APPLICATION PARTS 9. Assignment Papers (cover sheet & document(s)) 10. 37 CFR 3.73(b) Statement Power of (when there is an assignee) Attorney 11. English Translation Document (if applicable) 12. Information Disclosure Copies of IDS Statement (IDS)/PTO-1449 Citations 13. Preliminary Amendment 14. Return Receipt Postcard (MPEP 503) (Should be specifically itemized) 15. Certified Copy of Priority Document(s) (if foreign priority is claimed) Nonpublication Request under 35 U.S.C. 122 (b)(2)(B)(i). Applicant must attach form PTO/SB/35 or its equivalent. Other:								
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Effective 01/01/2003. Patent fees are subject to annual revision.		First Named Inventor			FARID, Abdol Hossain, et al.			
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TITLE: INSULIN-LIKE GROWTH FACTOR-1 RECEPTOR (IGF-1R) POLYMORPHIC ALLELES AND USE OF THE SAME TO IDENTIFY DNA MARKERS FOR REPRODUCTIVE LONGEVITY

5 BACKGROUND OF THE INVENTION

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Genetic mutations are the basis of evolution and genetic diversity. Genetic markers represent specific loci in the genome of a species, population or closely related species, and sampling of different genotypes at these marker loci reveals genetic variation. The genetic variation at marker loci can then be described and applied to genetic studies, commercial breeding, diagnostics, and cladistics. Genetic markers have the greatest utility when they are codominant, highly heritable, multi-allelic, and numerous. Most genetic markers are heritable because their alleles are determined by the nucleotide sequence of DNA which is highly conserved from one generation to the next, and the detection of their alleles is unaffected by the natural environment. Markers have multiple alleles because, in the evolutionary process, rare, genetically-stable mutations in DNA sequences defining marker loci arose and were disseminated through the generations along with other existing alleles. The highly conserved nature of DNA combined with rare occurrences of stable mutations allows genetic markers to be both predictable and discerning of different genotypes. The repertoire of genetic-marker technologies today allows multiple technologies to be used simultaneously in the same project. The invention of each new genetic-marker technology and each new DNA polymorphism adds additional utility to genetic markers. Many genetic-marker technologies exist. Some examples are restriction-fragment-length polymorphism (RFLP) Bostein et al (1980) Am J Hum Genet 32:314-331; single-strand conformation polymorphism (SSCP) Fischer et al. (1983) Proc Natl Acad Sci USA 80:1579-1583, Orita et al. (1989) Genomics 5:874-879; amplified fragment-length polymorphism (AFLP) Vos et al. (1995) Nucleic Acids Res 23:4407-4414; microsatellite or single-sequence repeat (SSR) Weber J L and May P E (1989) Am J Hum Genet 44:388-396; random-amplified polymorphic DNA (RAPD) Williams et al (1990) Nucleic Acids Res 18:6531-6535; sequence tagged site (STS) Olson et al. (1989) Science 245:1434-1435; genetic-bit analysis (GBA) Nikiforov et al (1994) Nucleic Acids Res 22:4167-4175; allelespecific polymerase chain reaction (ASPCR) Gibbs et al. (1989) Nucleic Acids Res 17:2437-2448, Newton et al. (1989) Nucleic Acids Res 17:2503-2516; nick-translation

PCR (e.g., TAQMAN^{TM.}) Lee et al. (1993) Nucleic Acids Res 21:3761-3766; and allelespecific hybridization (ASH) Wallace et al. (1979) Nucleic Acids Res 6:3543-3557, (Sheldon et al. (1993) Clinical Chemistry 39(4):718-719) among others. Each technology has its own particular basis for detecting polymorphisms in DNA sequence.

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The ability to follow a specific favorable genetic allele involves a novel and lengthy process of the identification of a DNA molecular marker for a major effect gene. The marker may be linked to a single gene with a major effect or linked to a number of genes with additive effects. DNA markers have several advantages; segregation is easy to measure and is unambiguous, and DNA markers are co-dominant, i.e., heterozygous and homozygous animals can be distinctively identified. Once a marker system is established selection decisions could be made very easily, since DNA markers can be assayed any time after a tissue or blood sample can be collected from the individual infant animal, or even an embryo.

Poor reproductive performance is one of the major causes for culling in dairy (Beaudeau et al. 1995; Durr et al. 1997; Kulak et al. 1997; Bascom and Young 1998) and beef cattle (Tanida et al. 1988), and leads to a decrease in profitability (Tanida et al. 1988; Beaudeau et al. 1995; Kulak et al. 1997; Bascom and Young 1998). The highest level of profitability in a dairy herd is achieved when high yielding cows are maintained in the herd for several lactations (Gill and Allaire 1976; Allaire and Gibson 1992; Kulak et al. 1997). An increase in length of production from 3 to 4 lactations increases milk yield per lactation 20 and profit per year by 11 and 13% respectively (Strandberg 1996). Reproductive longevity is even more important in beef cattle, sheep, swine and fur bearing animals, where replacement cost is, after nutrition, the second highest source of expenditure. Clearly, improving reproductive longevity offers one of the greatest opportunities for increasing productive efficiency and economic return in the multi-billion dollar livestock industry in 25 the world. This is illustrated by the fact that reproductive longevity is included in the national dairy genetic evaluation systems in Canada (herd life) and the U.S. (production life).

Moderate variation exists for reproductive longevity within and among different breeds of cattle (Silva et al. 1986; Smith and Quass 1984; Bailey 1991; Arthur et al. 1993), suggesting the possibility for genetic improvement in this trait. However, despite its

obvious economic importance, it is difficult to improve reproductive longevity through conventional breeding methods because of the low heritability of this trait (Smith and Quass 1984; Tanida et al. 1988; Boldman et al. 1992; VanRaden and Klaaskate 1993) and the long time necessary to obtain information on reproductive longevity in livestock. Attempts to improve reproductive longevity of dairy cattle through indirect selection, such as the use of 'type traits' that are measured early in life, has been ineffective (Smith and Quass 1984; Boldman et al. 1992; VanRaden and Klasskate 1993).

The above limitations make reproductive longevity an ideal candidate trait for the use of DNA markers (Lande and Thompson 1990), which would provide a means of identification of animals with superior breeding value at an early age on the basis of a simple laboratory test. Developing DNA markers for reproductive longevity is, however, a difficult and time-consuming task in long-lived livestock resources. A logical strategy would involve identification of candidate genes in a mammalian model with a short generation interval and later validating them in livestock (Copeland et al. 1993). This is especially true in the case of genes that control reproductive longevity and life span (Rose and Nusbaum 1994), since direct selection for prolonged reproductive age in large mammals is very time consuming and prohibitively expensive. The genes identified in animals will be putative candidates for the development of DNA markers for reproductive longevity in other species.

Although there are several reports on the quantitative genetics aspects of reproductive longevity in livestock (VanRaden and Klaaskate 1993; Smith and Quass 1984; Kulak et al. 1997; Bascom and Young 1998), little information is available on the genetic control of this trait in any mammalian species. Most of the available information on the genetic control of reproductive longevity and life span has been obtained on simple organisms, such as Drosophila and Caenorhabditis elegans (C. elegans). In C. elegans, for example, the daf genes (daf-2, -12, -16, -18 -23), which are components of the IGF-1R signaling cascade, have been shown to control the regulation of metabolism, development, reproduction and life span (Lakowski and Hekimi 1996; Apfeld and Kenyon 1998; Hekimi et al. 1998). Also, there is a positive relationship between life span and reproduction in C. elegans (Hsin and Kenyon 1999) and among mammals (Packer et al. 1998; Tissenbaum and Ruvkun 1998). Although information on lower organisms is useful, their usefulness in

mammals should be assessed in an appropriate mammalian model that exhibits widely contrasting reproductive longevity phenotypes.

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The use of DNA markers will facilitate the identification of animals that are genetically prone to a) reproduce longer than the average and, separately b) those that have a higher likelihood, compared with the average, of conceiving during lactation (sustained lactation and pregnancy stress). The marker may be directly involved in prolonging reproductive life, or may be linked to a single gene with a major effect, or may be linked to a number of genes with additive effects on animals' phenotype. Their segregation is easy to measure and is unambiguous, and DNA markers are co-dominant, i.e., heterozygous and homozygous animals can be distinctively identified. Once a marker system is established, selection decisions can be made easily, since DNA markers can be assayed any time after a tissue or blood sample can be collected from the individual infant animal, or even an embryo.

For the foregoing reasons, there is a need for a method of selecting animals with improved reproductive longevity and/or ability to better sustain stress factors. More particularly, a need for identifying markers which may be used to improve economically beneficial characteristics in animals by identifying and selecting animals with these favorable characteristics at the genetic level.

Therefore, an object of the present invention is to provide a method of identifying polymorphisms in the IGF-1R gene which are indicative of reproductive longevity in mammals and their ability to sustain performance in combination with stress factors such as lactation, pregnancy, and health status.

Another object of the invention is to provide assays for determining the presence of these genetic markers.

A further object of the invention is to provide methods for screening animals to determine those more likely to exhibit favorable traits associated with reproductive longevity and the ability to sustain performance under stress, which increases the accuracy of selection and breeding methods.

Yet another object of the invention is to provide PCR amplification and detection tests which will greatly expedite the determination of presence of the markers.

A still further object of the invention is to provide a method for determining the haplotype of the IGF-1R gene indicative of reproductive longevity and the ability to sustain performance under stress.

Additional objects and advantages of the invention will be set forth in part in the description that follows, and in part will be obvious from the description, or may be learned by the practice of the invention. The objects and advantages of the invention will be attained by means of the instrumentalities and combinations particularly pointed out in the appended claims.

BRIEF SUMMARY OF THE INVENTION

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This invention relates to the discovery of alternate forms of the insulin-like growth factor-1 receptor (IGF-1R) gene which are useful as a genetic markers associated with reproductive longevity and the ability to better sustain stress factors in animals such as lactation and pregnancy in animals.

According to an embodiment of the present invention there are provided methods for identifying a polymorphism in an animal. One embodiment includes a method for genetically identifying an animal comprising obtaining a sample of genetic material from an animal and assaying for the presence of a polymorphism in the insulin-like growth factor 1 receptor gene (IGF-1R), wherein said polymorphism is associated with reproductive longevity and/or ability to better sustain stress factors such as lactation and pregnancy stress.

A further embodiment includes a method for screening animals to determine those more likely to exhibit favorable traits associated with reproductive longevity and ability to sustain stress factors such as lactation and pregnancy stress. These methods include obtaining a genetic sample from the animal. The methods can further include assaying for the presence or absence of a polymorphism in the IGF-1R gene associated with reproductive longevity and/or the ability to sustain stress factors in animals such as lactation and pregnancy.

Further embodiments of the invention can include amplifying the gene or a region of the gene, which contains at least one polymorphism. Since one of the polymorphisms may involve changes in the amino acid composition of the IGF-1R protein, assay methods

may even involve ascertaining the amino acid composition of these proteins. Methods for this type or purification and analysis typically involve isolation of the protein through means including fluorescence tagging with antibodies, separation and purification of the protein (i.e., through reverse phase HPLC system), and use of an automated protein sequencer to identify the amino acid sequence present. Protocols for this assay are standard and known in the art and are disclosed in Ausubel et al. (eds.), Short Protocols in Molecular Biology 4th ed. (John Wiley and Sons 1999).

Another embodiment includes a method for determining the haplotype of the IGF-IR gene of an animal wherein the haplotype is indicative of reproductive longevity and/or ability to sustain stress factors.

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In a preferred embodiment, a sample of genetic material is obtained from an animal and the sample is analyzed to determine the presence or absence of a polymorphism in the IGF-1R gene, which is correlated with reproductive longevity and/or ability to sustain stress factors such as lactation and pregnancy stress.

As is well known to those of skill in this art, a variety of techniques may be utilized when comparing nucleic acid molecules for sequence differences. These include by way of example, restriction fragment length polymorphism analysis, heteroduplex analysis, single-strand conformation polymorphism analysis, denaturing gradient electrophoresis and temperature gradient electrophoresis.

In a preferred embodiment the polymorphism is a 12-bp deletion and two restriction fragment length polymorphism and the assay comprises identifying the animal's IGF-1R gene from isolated genetic material; exposing the gene to a restriction enzyme that yields restriction fragments of the gene of varying length; separating the restriction fragments to form a restriction pattern, such as by electrophoresis or HPLC separation; and comparing the resulting restriction fragment pattern from a IGF-1R gene that is either known to have or not to have the desired marker.

In a most preferred embodiment the gene is isolated by the use of primers and DNA polymerase to amplify a specific region of the gene which contains the polymorphism. Next the amplified region is digested with a restriction enzyme and fragments are again separated. Visualization of the RFLP pattern is by simple staining of the fragments, or by labeling the primers or the nucleoside triphosphates used in amplification.

It expected that with no more than routine testing as described herein this marker can be applied to different animal species to select for reproductive longevity and/or sustained performance in a situation with stress caused by lactation, pregnancy, or health status based on the teachings herein. Female animals of the same breed or breed cross or similar genetic lineage are bred, and the reproductive longevity and/or sustained lactation and pregnancy stress shown by each animal is determined and correlated. For other species in which sequences are available a BLAST comparison of the IGF-1R may be used to ascertain whether the particular allele disclosed herein is present.

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The term "analogous polymorphism" shall be a polymorphism which is the same as any of those disclosed herein as determined by BLAST comparisons.

The following terms are used to describe the sequence relationships between two or more nucleic acids or polynucleotides: (a) "reference sequence", (b) "comparison window", (c) "sequence identity", (d) "percentage of sequence identity", and (e) "substantial identity".

- (a) As used herein, "reference sequence" is a defined sequence used as a basis for sequence comparison. In this case the Reference is the IGF-1R sequence. A reference sequence may be a subset or the entirety of a specified sequence; for example, as a segment of a full-length cDNA or gene sequence, or the complete cDNA or gene sequence.
- (b) As used herein, "comparison window" includes reference to a contiguous and specified segment of a polynucleotide sequence, wherein the polynucleotide sequence may be compared to a reference sequence and wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Generally, the comparison window is at least 20 contiguous nucleotides in length, and optionally can be 30, 40, 50, 100, or longer. Those of skill in the art understand that to avoid a high similarity to a reference sequence due to inclusion of gaps in the polynucleotide sequence, a gap penalty is typically introduced and is subtracted from the number of matches.

Methods of alignment of sequences for comparison are well-known in the art.

Optimal alignment of sequences for comparison may be conducted by the local homology algorithm of Smith and Waterman, Adv. Appl. Math. 2:482 (1981); by the homology alignment algorithm of Needleman and Wunsch, J. Mol. Biol. 48:443 (1970); by the search

for similarity method of Pearson and Lipman, Proc. Natl. Acad. Sci. 85:2444 (1988); by computerized implementations of these algorithms, including, but not limited to: CLUSTAL in the PC/Gene program by Intelligenetics, Mountain View, California; GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group (GCG), 575 Science Dr., Madison, Wisconsin, USA; the 5 CLUSTAL program is well described by Higgins and Sharp, Gene 73:237-244 (1988); Higgins and Sharp, CABIOS 5:151-153 (1989); Corpet, et al., Nucleic Acids Research 16:10881-90 (1988); Huang, et al., Computer Applications in the Biosciences 8:155-65 (1992), and Pearson, et al., Methods in Molecular Biology 24:307-331 (1994). The BLAST family of programs which can be used for database similarity searches includes: 10 BLASTN for nucleotide query sequences against nucleotide database sequences; BLASTX for nucleotide query sequences against protein database sequences; BLASTP for proteinquery sequences against protein database sequences; TBLASTN for protein query sequences against nucleotide database sequences; and TBLASTX for nucleotide query sequences against nucleotide database sequences. See, Current Protocols in Molecular 15 Biology, Chapter 19, Ausubel, et al., Eds., Greene Publishing and Wiley-Interscience, New York (1995).

Unless otherwise stated, sequence identity/similarity values provided herein refer to the value obtained using the BLAST 2.0 suite of programs using default parameters.

Altschul et al., *Nucleic Acids Res.* 25:3389-3402 (1997). Software for performing BLAST analyses is publicly available, e.g., through the National Center for Biotechnology-Information (http://www.ncbi.nlm.nih.gov/).

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This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold (Altschul et al., supra). These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are then extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always > 0) and N (penalty score for mismatching residues;

always < 0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T, and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation (E) of 10, a cutoff of 100, M=5, N=-4, and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults a wordlength (W) of 3, an expectation (E) of 10, and the BLOSUM62 scoring matrix (see Henikoff & Henikoff (1989) *Proc. Natl. Acad. Sci. USA* 89:10915).

In addition to calculating percent sequence identity, the BLAST algorithm also performs a statistical analysis of the similarity between two sequences (see, e.g., Karlin & Altschul, *Proc. Natl. Acad. Sci. USA* 90:5873-5787 (1993)). One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance.

BLAST searches assume that proteins can be modeled as random sequences. However, many real proteins comprise regions of nonrandom sequences which may be homopolymeric tracts, short-period repeats, or regions enriched in one or more amino acids. Such low-complexity regions may be aligned between unrelated proteins even though other regions of the protein are entirely dissimilar. A number of low-complexity filter programs can be employed to reduce such low-complexity alignments. For example, the SEG (Wooten and Federhen, *Comput. Chem.*, 17:149-163 (1993)) and XNU (Claverie and States, *Comput. Chem.*, 17:191-201 (1993)) low-complexity filters can be employed alone or in combination.

(c) As used herein, "sequence identity" or "identity" in the context of two nucleic acid or polypeptide sequences includes reference to the residues in the two sequences which are the same when aligned for maximum correspondence over a specified comparison window. When percentage of sequence identity is used in reference to proteins it is recognized that residue positions which are not identical often differ by conservative

amino acid substitutions, where amino acid residues are substituted for other amino acid residues with similar chemical properties (e.g. charge or hydrophobicity) and therefore do not change the functional properties of the molecule. Where sequences differ in conservative substitutions, the percent sequence identity may be adjusted upwards to correct for the conservative nature of the substitution. Sequences which differ by such conservative substitutions are said to have "sequence similarity" or "similarity". Means for making this adjustment are well-known to those of skill in the art. Typically this involves scoring a conservative substitution as a partial rather than a full mismatch, thereby increasing the percentage sequence identity. Thus, for example, where an identical amino acid is given a score of 1 and a non-conservative substitution is given a score of zero, a conservative substitution is given a score between zero and 1. The scoring of conservative substitutions is calculated, e.g., according to the algorithm of Meyers and Miller, Computer Applic. Biol. Sci., 4:11-17 (1988) e.g., as implemented in the program PC/GENE (Intelligenetics, Mountain View, California, USA).

- (d) As used herein, "percentage of sequence identity" means the value determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison and multiplying the result by 100 to yield the percentage of sequence identity.
- (e) The term "substantial identity" of polynucleotide sequences means that a polynucleotide comprises a sequence that has at least 70% sequence identity, preferably at least 80%, more preferably at least 90% and most preferably at least 95%, compared to a reference sequence using one of the alignment programs described using standard parameters. One of skill will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning and

the like. Substantial identity of amino acid sequences for these purposes normally means sequence identity of at least 60%, or preferably at least 70%, 80%, 90%, and most preferably at least 95%.

These programs and algorithms can ascertain the analogy of a particular polymorphism in a target gene to those disclosed herein. It is expected that this polymorphism will exist in other animals and use of the same in other animals than disclosed herein involved no more than routine optimization of parameters using the teachings herein.

It is also possible to establish linkage between specific alleles of alternative DNA markers and alleles of DNA markers known to be associated with a particular gene (e.g. the IGF-1R gene discussed herein), which have previously been shown to be associated with a particular trait. Thus, in the present situation, taking the IGF-1R gene, it would be possible, at least in the short term, to select for animals likely to produce one or more of the traits of reproductive longevity and/or the ability to better sustain stress caused by lactation and pregnancy, or alternatively against animals less likely to exhibit the traits of reproductive longevity and/or the ability to better sustain stress caused by lactation and pregnancy, indirectly, by selecting for certain alleles of a IGF-1R associated marker through the selection of specific alleles of alternative chromosome markers. As used herein the term "genetic marker" shall include not only the polymorphism disclosed by any means of assaying for the protein changes associated with the polymorphism, be they linked markers, use of microsatellites, or even other means of assaying for the causative protein changes indicated by the marker and the use of the same to influence the traits of reproductive longevity and/or the ability to sustain stress in an animal.

As used herein, often the designation of a particular polymorphism is made by the name of a particular restriction enzyme. This is not intended to imply that the only way that the site can be identified is by the use of that restriction enzyme. There are numerous databases and resources available to those of skill in the art to identify other restriction enzymes which can be used to identify a particular polymorphism. Two examples are: http://www.geneseo.edu/~bio/ and http://www.firstmarket.com/cutter/cut2.html. In fact, as disclosed in the teachings herein there are numerous ways of identifying a particular

polymorphism or allele with alternate methods which may not even include a restriction enzyme, but which assay for the same genetic or proteomic alternative form.

The invention is intended to include these sequences as well as all conservatively modified variants thereof as well as those sequences which will hybridize under conditions of high stringency to the sequences disclosed. The term IGF-1R is used herein shall be interpreted to include these conservatively modified variants as well as those hybridized sequences.

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The term "conservatively modified variants" applies to both amino acid and nucleic acid sequences. With respect to particular nucleic acid sequences, conservatively modified variants refer to those nucleic acids which encode identical or conservatively modified variants of the amino acid sequences. Because of the degeneracy of the genetic code, a large number of functionally identical nucleic acids encode any given protein. For instance, the codons GCA, GCC, GCG and GCU all encode the amino acid alanine. Thus, at every position where an alanine is specified by a codon, the codon can be altered to any of the corresponding codons described without altering the encoded polypeptide. Such nucleic acid variations are "silent variations" and represent one species of conservatively modified variation. Every nucleic acid sequence herein that encodes a polypeptide also, by reference to the genetic code, describes every possible silent variation of the nucleic acid. One of ordinary skill will recognize that each codon in a nucleic acid (except AUG, which is ordinarily the only codon for methionine; and UGG, which is ordinarily the only codon for tryptophan) can be modified to yield a functionally identical molecule. Accordingly, each silent variation of a nucleic acid which encodes a polypeptide of the present invention is implicit in each described polypeptide sequence and is within the scope of the present invention.

As to amino acid sequences, one of skill will recognize that individual substitutions, deletions or additions to a nucleic acid, peptide, polypeptide, or protein sequence which alters, adds or deletes a single amino acid or a small percentage of amino acids in the encoded sequence is a "conservatively modified variant" where the alteration results in the substitution of an amino acid with a chemically similar amino acid. Thus, any number of amino acid residues selected from the group of integers consisting of from 1 to 15 can be so altered. Thus, for example, 1, 2, 3, 4, 5, 7, or 10 alterations can be made.

Conservatively modified variants typically provide similar biological activity as the unmodified polypeptide sequence from which they are derived. For example, substrate specificity, enzyme activity, or ligand/receptor binding is generally at least 30%, 40%, 50%, 60%, 70%, 80%, or 90% of the native protein for its native substrate. Conservative substitution tables providing functionally similar amino acids are well known in the art.

The following six groups each contain amino acids that are conservative substitutions for one another:

- 1) Alanine (A), Serine (S), Threonine (T);
- Aspartic acid (D), Glutamic acid (E);
- 3) Asparagine (N), Glutamine (Q);

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- 4) Arginine (R), Lysine (K);
- 5) Isoleucine (I), Leucine (L), Methionine (M), Valine (V); and
- 6) Phenylalanine (F), Tyrosine (Y), Tryptophan (W).

See also, Creighton, Proteins, W.H. Freeman and Company (1984).

By "encoding" or "encoded", with respect to a specified nucleic acid, is meant comprising the information for translation into the specified protein. A nucleic acid encoding a protein may comprise non-translated sequences (e.g., introns) within translated regions of the nucleic acid, or may lack such intervening non-translated sequences (e.g., as in cDNA). The information by which a protein is encoded is specified by the use of codons. Typically, the amino acid sequence is encoded by the nucleic acid using the "universal" genetic code. However, variants of the universal code, such as are present in some plant, animal, and fungal mitochondria, the bacterium Mycoplasma capricolum, or the ciliate Macronucleus, may be used when the nucleic acid is expressed therein.

The term "stringent conditions" or "stringent hybridization conditions" includes reference to conditions under which a probe will hybridize to its target sequence, to a detectably greater degree than to other sequences (e.g., at least 2-fold over background). Stringent conditions are sequence-dependent and be different in different circumstances. By controlling the stringency of the hybridization and/or washing conditions, target sequences can be identified which are 100% complementary to the probe (homologous probing). Alternatively, stringency conditions can be adjusted to allow some mismatching in sequences so that lower degrees of similarity are detected (heterologous probing).

Generally, a probe is less than about 1000 nucleotides in length, optionally less than 500 nucleotides in length.

Typically, stringent conditions will be those in which the salt concentration is less than about 1.5 M Na ion, typically about 0.01 to 1.0 M Na ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30°C for short probes (e.g., 10 to 50 nucleotides) and at least about 60°C for long probes (e.g., greater than 50 nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide. One of ordinary skill is apprised in knowing that the time of the hybridization is dependent on the concentration of the probe. Exemplary low stringency conditions include hybridization with a buffer solution of 30 to 35% formamide, 1 M NaCl, 1% SDS (sodium dodecyl sulphate) at 37°C, and a wash in 1X to 2X SSC (20X SSC = 3.0 M NaCl/0.3 M trisodium citrate) at 50 to 55°C. Exemplary moderate stringency conditions include hybridization in 40 to 45% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0.5X to 1X SSC at 55 to 50°C. Exemplary high stringency conditions include hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0.1X SSC at 60 to 65°C for at least 15 minutes.

Specificity is typically the function of post-hybridization washes, the critical factors being the ionic strength and temperature of the final wash solution. For DNA-DNA hybrids, the T_m can be approximated from the equation of Meinkoth and Wahl, Anal. Biochem., 138:267-284 (1984): T_m =81.5°C + 16.6 (log M) + 0.41 (%GC) -0.61 (% form) - 500/L; where M is the molarity of monovalent cations, %GC is the percentage of guanosine and cytosine nucleotides in the DNA, % form is the percentage of formamide in the hybridization solution, and L is the length of the hybrid in base pairs. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the complementary target sequence hybridizes to a perfectly matched probe. T_m is reduced by about 1°C for each 1% of mismatching; thus, T_m , hybridization and/or wash conditions can be adjusted to hybridize to sequences of the desired identity. For example, if sequences with \geq 90% identity are sought, the T_m can be decreased 10°C. Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence and its complement at a defined ionic strength and pH. However, severely stringent conditions can utilize a hybridization and/or wash at 1, 2, 3, or 4°C lower than the

thermal melting point (T_m); moderately stringent conditions can utilize a hybridization and/or wash at 6, 7, 8, 9, or 10°C lower than the thermal melting point (T_m); low stringency conditions can utilize a hybridization and/or wash at 11, 12, 13, 14, 15, or 20°C lower than the thermal melting point (T_m). Using the equation, hybridization and wash compositions, and desired T_m, those of ordinary skill will understand that variations in the stringency of hybridization and/or wash solutions are inherently described. If the desired degree of mismatching results in a T_m of less than 45°C (aqueous solution) or 32°C (formamide solution) it is preferred to increase the SSC concentration so that a higher temperature can be used. An extensive guide to the hybridization of nucleic acids is found in Tijssen, Laboratory Techniques in Biochemistry and Molecular Biology—Hybridization with Nucleic Acids Probes, Part I, Chapter 2, Ausubel, et al., Eds., Greene Publishing and Wiley-Interscience, New York (1995).

These and other features, aspects, and advantages of the present invention will become better understood with regard to the following description, appended claims, and accompanying drawings.

BRIEF DESCRIPTION OF THE DRAWINGS

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Figure 1 depicts the nucleotide sequence of the insulin-like growth factor-1 receptor in mice (SEQ ID NO:1)(GenBank accession number AF056187).

Figure 2 depicts the amino acid sequence of the insulin-like growth factor-1 receptor in mice (SEQ ID NO:2)(GenBank protein id AAC12782.1).

Figure 3 depicts the mRNA sequence of insulin-like growth factor I receptor in mice (SEQ ID NO:3) (Genbank accession number XM_133508).

Figure 4 depicts the alignment of exon 21 of the mouse IGF1-R sequences from Genbank accession number AF056187 (SEQ ID NO: 1) and Genbank accession number XM_133508 (SEQ ID NO:3), and the amino acid sequence of this region (SEQ ID NO:4). The A to G substitution at position 3876 of the Genbank accession number AF056187 (Hpall site, locus B) is bolded and underlined. The 12 bp insertion/deletion is bolded and underlined. The junction of exon 20 and exon 21 is shown by "0".

Figure 5 depicts intron 16 (SEQ ID NO:5) of the mouse IGF1-R gene and the surrounding exons amplified by primers PSEQ16F (SEQ ID NO:12) and PSEQ16R (SEQ

ID NO:13), and its alignment with the mouse IGF1-R gene (Genbank accession number AC101879; SEQ ID NO:6). This sequence contains 102 bp of exon 16 (nucleotides 1 to 102), 283 bp of intron 16 (nucleotides 103 to 385) and 101 bp of exon 17 (nucleotides 386 to 486) of the mouse IFG1-R gene. Exon-intron junctions are shown by 0. The 'G' insertion is at position 176 of SEQ ID NO:5 after nucleotide 56456 of SEQ ID NO:6 (Genbank accession number AC101879). This insertion is bolded and underlined. Note that SEQ ID NO:6 (Genbank accession number AC101879) is the reverse complement of other sequences of the IGF1-R in Genbank. The 'G' to 'A' substitution (*DpnII* site, locus A) is at position 331 of SEQ ID NO:5, corresponding to nucleotide 556303 of SEQ ID NO:6 (Genbank accession number AC101879). This nucleotide is bolded and underlined. The forward (PSEQ16F) and reverse (PSEQ16R) primers are underlined.

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Figure 6 depicts mouse clone RP23-378H21, complete sequence (SEQ ID NO:6) (Genbank accession number AC101879).

Figure 7 depicts the nucleotide sequence of the insulin-like growth factor-1 receptor in pig (SEQ ID NO:7). cDNA sequence in lower case letters comes from Accession No. AB003362. Intron 9 sequence in lower case letters comes from Accession No. AJ491314. Intron sequence in upper case letters was derived from Applicants sequencing efforts.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENT

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Unless mentioned otherwise, the techniques employed or contemplated herein are standard methodologies well known to one of ordinary skill in the art.

As used herein, "reproductive longevity" means a biologically significant increase in the number of pregnancies and/or the duration of time an animal is capable of reproduction, relative to the mean of a given population, group or species.

As used herein, "the ability to sustain performance under stress" means a biologically significant increase in performance, in situations with stress, i.e., increase in the number of pregnancies and/or the duration of time while the animal is lactating and raising progeny, i.e., carrying a fetus while lactating at the same time, relative to the mean of a given population.

The insulin-like growth factor-1 receptor (IGF-1R) gene is a plasma membrane-bound disulfide-bonded heterotetrameric glycoprotein composed of two extracellular α -subunits containing a ligand binding domain and two transmembrane β -subunits that include a cytoplasmic tyrosine kinase domain (Richards et al., 1998). The IGF-1R gene plays a vital role in growth and development in several different ways, such as mediating mitogenic and metabolic responses, maintaining transformed cell phenotype, protecting cells from apoptotic injuries, and inducing differentiation in certain cell types especially myoblasts, adipocytes, osteoblasts and cells of the central nervous system (Valentinis et al., 1999; Jin et al., 2000).

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Binding of the ligand to IGF-1R leads to autophosphorylation of the α-subunit and activation of the β-subunit tyrosine kinase domains resulting in phosphorylation of several intracellular proteins including insulin receptor substrates (IRS) and Shc with the subsequent trigger of multiple signaling cascades, for instance those of the Ras-Raf-MAP kinase network and phosphatidylinositol 3-kinase. The various effects may depend on specific domains of the receptor and the availability of different substrates (Peruzzi et al., 1999; Swantek et al., 1999; Valentinis et al., 1999; Xu et al., 1999; Soni et al.; 2000).

The IGF-1R gene also plays a role in certain functions of other growth factors and hormones. There is evidence that a signal generated by a functional IGF-1R is required for the mitogenic effects of other growth factors, such as epidermal growth factor (EGF) and platelet-derived growth factor (PDGF) (Swantek and Baserga, 1999). Furthermore, the estradiol-induced mitogenic effects in the mouse uterus and differentiation of rat adipocytes are dependent on the IGF-1R (Richards et al., 1998; Dieudonne et al., 2000).

According to an embodiment of the present invention variants or polymorphic sites in the IGF-1R gene have been located, and these genetic polymorphisms are associated with reproductive longevity and/or the ability the sustain stress factors such as lactation and pregnancy in mice. These four variants include an 'A' to 'G' substitution in intron 16, a 'G' nucleotide insertion in intron 16, an 'A' to 'G' substitution in exon 21, and a 12 bp-deletion in exon 21 which resulted in four fewer amino acids in the IGF-1R protein.

In another embodiment, assays are provided for detection of these different variants. The assays preferably involve amplifying the genomic DNA purified from blood, tissue,

semen, or other convenient source of genetic material by the use of primers and standard techniques, such as polymerase chain reaction (PCR).

A 12 bp deletion, PCR product was identified in mice. The PCR product can be sized in a variety of ways, such as by agarose or polyacrylamide gel electrophoresis, use of an automated DNA sequencer, or mass spectrometry.

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An 'A' to 'G' substitution, at position 3876 of SEQ ID NO:1 (Genbank accession number AF056187) was identified in mice. The PCR product was digested with a restriction enzyme (e.g., *HpaII*) so as to yield gene fragments of varying lengths, as separating at least some of the fragments from others using agarose or polyacrylamide gel electrophoresis. Since the 'A' to 'G' substitution is 20 base pairs upstream from the 12-base pair deletion, both polymorphisms may be detected by the digestion of PCR product with the enzyme *HpaII*.

A 'G' to 'A' substitution (GGTC to GATC) was detected in intron 16 of the gene in mice. The 486 bp PCR product, spanning exons 16 and 17 and intron 16, was cut into 454 and 32 bp fragments (A₁ allele) by the enzyme *DpnII* (↑GATC). This nucleotide substitution resulted in the creation of a new recognition site for this enzyme, which cleaved the 454 bp fragment into 328 and 125 bp fragments (A₂ allele). In addition, sequence information revealed a 'G' nucleotide insertion in intron 16, 153 bp 5' to the above point mutation, but no restriction enzyme was found for discriminatory typing of this deletion.

In porcine, the following single nucleotide polymorphisms were found:

A 'G' to 'A' substitution, designated SNP16i27, at position 27 from the end of intron 16 was detected with an AvaII restriction site.

A 'G' to 'C' substitution, designated SNP16i73, was detected at position 73 from the end of intron 16. This nucleotide substitution resulted in a Mn11 restriction site.

A 'G' to 'A' substitution, designated SNP1772, was detected in exon 8. This nucleotide substitution resulted in a *TaqI* restriction site.

The polymorphisms in animals may also be identified using a variety of methods such as direct sequencing, and hybridizing with nucleotide probes labeled with radioactive or chemiluminescence. The probes may be sequences containing all or a portion of the IGF-1R gene containing the polymorphisms, which will be hybridized to the separated

digestion PCR products or digested genomic DNA. The polymorphism may also be detected by restriction fragment length polymorphism (RFLP) analysis, the single-stranded conformation polymorphism of the PCR product (SSCP-PCR), PCR amplification of specific alleles, the amplification of DNA target by PCR followed by single base extension which will be detected by fluorescent or radioactive substances or mass spectrometry, allelic discrimination during PCR, Genetic Bit Analysis, Pyrosequencing, oligonucleotide ligation assay, analysis of melting curves or other methods which detect differences in the length of a DNA fragment at this region or detect a single nucleotide substitution.

Another embodiment of the invention includes novel PCR primers comprising 4 to 30 contiguous bases on either side of the polymorphism to provide an amplification system allowing for detection of the polymorphism by PCR and identification of the fragments by standard methods. Any primers amplifying the region of the polymorphism may be used as taught herein and are also publically available.

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The preferred primers for revealing the 12 bp deletion are PSEQDF: 5'-GGA GAT CAT CGG CAG CAT CAA G-3' (SEQ ID NO:8), wherein the 5' end is at position 3786 of the mouse IGF-1R gene and PSEQDR: 5'-GCC ATT CTC AGC CTT GTG TCC-3' (SEQ ID NO:9), wherein the 5' end is at the position 4002 of the mouse IGF-1R gene.

The preferred primers for revealing the A to G substitution in exon 21 of the IGF-1R gene are PSECAF: 5'-GCA TGT GCT GGC AGT ATA ACC-3' (SEQ ID NO:10), wherein the 5' end is at position 3743 of the IGF-1R gene and PSECAR: 5'CAG AGG CCC ATG TCA GTT AAG (SEQ ID NO:11), wherein the 5' end is at position 4376 of the IGF-1R gene.

The preferred primers for revealing the G to A substitution in intron 16 of the IGF-1R gene are PSEQ16F: 5' AGA GTG GCC ATC AAG ACG GTA 3' (SEQ ID NO:12) and PSEQ16R: 5' GGC CTC AGA GAC CGG AGA T 3' (SEQ ID NO:13).

In porcine, the preferred primers for revealing SNP16i27 identified with an AvaII restriction site are Primer 16: 5' - CCT CCG TGA TGA AGG AGT TC - 3' (SEQ ID NO:14) and Primer 17: 5' - TCA GTT CCA TGA TGA CCA GC - 3' (SEQ ID NO:15).

The preferred primers for revealing SNP16i73 identified with a *Mn11* restriction site are Primer 16: 5' – CCT CCG TGA TGA AGG AGT TC – 3' (SEQ ID NO:16) and Primer 17: 5' – TCA GTT CCA TGA TGA CCA GC – 3' (SEQ ID NO:17).

The preferred primers for revealing SNP1772 identified with a *TaqI* restriction site are designated as Primer 9: 5' – GGA GTA TGA TGG GCA GGA T – 3' (SEQ ID NO:18) and Primer 8: 5' – GAA GCA TTG GTG CGA ATG TA – 3' (SEQ ID NO:19).

Computer programs available on the world wide web allows one of ordinary skill in the art to design other primers capable of amplifying polymorphic segments of the IGF-1R gene such as those shown above and depicted in Table 1. See Steve Rozen and Helen J. Skaletsky (2000) Primer3 on the WWW for general users and for biologist programmers. In: Krawetz S, Misener S (eds) Bioinformatics Methods and Protocols: Methods in Molecular Biology. Humana Press, Totowa, NJ, pp 365-386.

A further embodiment comprises a breeding method whereby assays of the above types are conducted on a plurality of gene sequences from different animals or animal embryos of various species to be selected from and, based on the results, certain animals are either selected or dropped out of the breeding program.

The following is a general overview of techniques which can be used to assay for the polymorphisms of the invention.

In the present invention, a sample of genetic material is obtained from an animal. Samples can be obtained from blood, tissue, semen, etc. Generally, peripheral blood cells are used as the source, and the genetic material is DNA. A sufficient amount of cells are obtained to provide a sufficient amount of DNA for analysis. This amount will be known or readily determinable by those skilled in the art. The DNA is isolated from the blood cells by techniques known to those skilled in the art.

Isolation and Amplification of Nucleic Acid

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Samples of genomic DNA are isolated from any convenient source including saliva, buccal cells, hair roots, blood, cord blood, amniotic fluid, interstitial fluid, peritoneal fluid, chorionic villus, and any other suitable cell or tissue sample with intact nuclei. The cells can also be obtained from solid tissue as from a fresh or preserved organ or from a tissue sample or biopsy. The sample can contain compounds which are not naturally intermixed with the biological material such as preservatives, anticoagulants, buffers, fixatives, nutrients, antibiotics, or the like.'

Methods for isolation of genomic DNA from these various sources are described in, for example, Kirby, DNA Fingerprinting, An Introduction, W.H. Freeman & Co. New York (1992). Genomic DNA can also be isolated from cultured primary or secondary cell cultures or from transformed cell lines derived from any of the aforementioned tissue samples.

Samples of animal RNA can also be used. RNA can be isolated from tissues expressing the IGF-1R gene as described in Sambrook et al., supra. RNA can be total cellular RNA, mRNA, poly A+ RNA, or any combination thereof. For best results, the RNA is purified, but can also be unpurified cytoplasmic RNA. RNA can be reverse transcribed to form DNA which is then used as the amplification template, such that the PCR indirectly amplifies a specific population of RNA transcripts. See, e.g., Sambrook, supra, Kawasaki et al., Chapter 8 in *PCR Technology*, (1992) *supra*, and Berg et al., Hum. Genet. 85:655-658 (1990).

15 PCR Amplification

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The most common means for amplification is polymerase chain reaction (PCR), as described in U.S. Pat. Nos. 4,683,195, 4,683,202, 4,965,188 each of which is hereby incorporated by reference. If PCR is used to amplify the target regions in blood cells, heparinized whole blood should be drawn in a sealed vacuum tube kept separated from other samples and handled with clean gloves. For best results, blood should be processed immediately after collection; if this is impossible, it should be kept in a sealed container at 4°C until use. Cells in other physiological fluids may also be assayed. When using any of these fluids, the cells in the fluid should be separated from the fluid component by centrifugation.

Tissues should be roughly minced using a sterile, disposable scalpel and a sterile needle (or two scalpels) in a 5 mm Petri dish. Procedures for removing paraffin from tissue sections are described in a variety of specialized handbooks well known to those skilled in the art.

To amplify a target nucleic acid sequence in a sample by PCR, the sequence must be accessible to the components of the amplification system. One method of isolating target DNA is crude extraction which is useful for relatively large samples. Briefly,

mononuclear cells from samples of blood, amniocytes from amniotic fluid, cultured chorionic villus cells, or the like are isolated by layering on sterile Ficoll-Hypaque gradient by standard procedures. Interphase cells are collected and washed three times in sterile phosphate buffered saline before DNA extraction. If testing DNA from peripheral blood lymphocytes, an osmotic shock (treatment of the pellet for 10 sec with distilled water) is suggested, followed by two additional washings if residual red blood cells are visible following the initial washes. This will prevent the inhibitory effect of the heme group carried by hemoglobin on the PCR reaction. If PCR testing is not performed immediately after sample collection, aliquots of 10^6 cells can be pelleted in sterile Eppendorf tubes and the dry pellet frozen at -20°C until use.

The cells are resuspended (10^6 nucleated cells per $100~\mu$ l) in a buffer of 50 mM Tris-HC1 (pH 8.3), 50 mM KC1 1.5 mM MgC1₂, 0.5% Tween 20, 0.5% NP40 supplemented with $100~\mu$ g/ml of proteinase K. After incubating at 56° C for 2 hr. the cells are heated to 95° C for 10 min to inactivate the proteinase K and immediately moved to wet ice (snap-cool). If gross aggregates are present, another cycle of digestion in the same buffer should be undertaken. Ten μ l of this extract is used for amplification.

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When extracting DNA from tissues, e.g., chorionic villus cells or confluent cultured cells, the amount of the above mentioned buffer with proteinase K may vary according to the size of the tissue sample. The extract is incubated for 4-10 hrs at 50°-60°C and then at 95°C for 10 minutes to inactivate the proteinase. During longer incubations, fresh proteinase K should be added after about 4 hr at the original concentration.

When the sample contains a small number of cells, extraction may be accomplished by methods as described in Higuchi, "Simple and Rapid Preparation of Samples for PCR", in *PCR Technology*, Ehrlich, H.A. (ed.), Stockton Press, New York, which is incorporated herein by reference. PCR can be employed to amplify target regions in very small numbers of cells (1000-5000) derived from individual colonies from bone marrow and peripheral blood cultures. The cells in the sample are suspended in 20 μl of PCR lysis buffer (10 mM Tris-HC1 (pH 8.3), 50 mM KC1, 2.5 mM MgC1₂, 0.1 mg/ml gelatin, 0.45% NP40, 0.45% Tween 20) and frozen until use. When PCR is to be performed, 0.6 μl of proteinase K (2 mg/ml) is added to the cells in the PCR lysis buffer. The sample is then heated to about

60°C and incubated for 1 hr. Digestion is stopped through inactivation of the proteinase K by heating the samples to 95°C for 10 min and then cooling on ice.

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A relatively easy procedure for extracting DNA for PCR is a salting out procedure adapted from the method described by Miller et al., Nucleic Acids Res. 16:1215 (1988), which is incorporated herein by reference. Mononuclear cells are separated on a Ficoll-Hypaque gradient. The cells are resuspended in 3 ml of lysis buffer (10 mM Tris-HC1, 400 mM NaC1, 2 mM Na₂ EDTA, pH 8.2). Fifty µl of a 20 mg/ml solution of proteinase K and 150 µl of a 20% SDS solution are added to the cells and then incubated at 37°C overnight. Rocking the tubes during incubation will improve the digestion of the sample. If the proteinase K digestion is incomplete after overnight incubation (fragments are still visible), an additional 50 µl of the 20 mg/ml proteinase K solution is mixed in the solution and incubated for another night at 37°C on a gently rocking or rotating platform. Following adequate digestion, one ml of a 6M NaCl solution is added to the sample and vigorously mixed. The resulting solution is centrifuged for 15 minutes at 3000 rpm. The pellet contains the precipitated cellular proteins, while the supernatant contains the DNA. The supernatant is removed to a 15 ml tube that contains 4 ml of isopropanol. The contents of the tube are mixed gently until the water and the alcohol phases have mixed and a white DNA precipitate has formed. The DNA precipitate is removed and dipped in a solution of 70% ethanol and gently mixed. The DNA precipitate is removed from the ethanol and airdried. The precipitate is placed in distilled water and dissolved.

Kits for the extraction of high-molecular weight DNA for PCR include a Genomic Isolation Kit A.S.A.P. (Boehringer Mannheim, Indianapolis, Ind.), Genomic DNA Isolation System (GIBCO BRL, Gaithersburg, Md.), Elu-Quik DNA Purification Kit (Schleicher & Schuell, Keene, N.H.), DNA Extraction Kit (Stratagene, LaJolla, Calif.), TurboGen Isolation Kit (Invitrogen, San Diego, Calif.), and the like. Use of these kits according to the manufacturer's instructions is generally acceptable for purification of DNA prior to practicing the methods of the present invention.

The concentration and purity of the extracted DNA can be determined by spectrophotometric analysis of the absorbance of a diluted aliquot at 260 nm and 280 nm. After extraction of the DNA, PCR amplification may proceed. The first step of each cycle of the PCR involves the separation of the nucleic acid duplex formed by the primer

extension. Once the strands are separated, the next step in PCR involves hybridizing the separated strands with primers that flank the target sequence. The primers are then extended to form complementary copies of the target strands. For successful PCR amplification, the primers are designed so that the position at which each primer hybridizes along a duplex sequence is such that an extension product synthesized from one primer, when separated from the template (complement), serves as a template for the extension of the other primer. The cycle of denaturation, hybridization, and extension is repeated as many times as necessary to obtain the desired amount of amplified nucleic acid.

In a particularly useful embodiment of PCR amplification, strand separation is achieved by heating the reaction to a sufficiently high temperature for a sufficient time to cause the denaturation of the duplex but not to cause an irreversible denaturation of the polymerase (see U.S. Pat. No. 4,965,188, incorporated herein by reference). Typical heat denaturation involves temperatures ranging from about 80°C to 105°C for times ranging from seconds to minutes. Strand separation, however, can be accomplished by any suitable denaturing method including physical, chemical, or enzymatic means. Strand separation may be induced by a helicase, for example, or an enzyme capable of exhibiting helicase activity. For example, the enzyme RecA has helicase activity in the presence of ATP. The reaction conditions suitable for strand separation by helicases are known in the art (see Kuhn Hoffman-Berling, 1978, CSH-Quantitative Biology, 43:63-67; and Radding, 1982, Ann. Rev. Genetics 16:405-436, each of which is incorporated herein by reference).

Template-dependent extension of primers in PCR is catalyzed by a polymerizing agent in the presence of adequate amounts of four deoxyribonucleotide triphosphates (typically dATP, dGTP, dCTP, and dTTP) in a reaction medium comprised of the appropriate salts, metal cations, and pH buffering systems. Suitable polymerizing agents are enzymes known to catalyze template-dependent DNA synthesis. In some cases, the target regions may encode at least a portion of a protein expressed by the cell. In this instance, mRNA may be used for amplification of the target region. Alternatively, PCR can be used to generate a cDNA library from RNA for further amplification, the initial template for primer extension is RNA. Polymerizing agents suitable for synthesizing a complementary, copy-DNA (cDNA) sequence from the RNA template are reverse transcriptase (RT), such as avian myeloblastosis virus RT, Moloney murine leukemia virus

RT, or Thermus thermophilus (Tth) DNA polymerase, a thermostable DNA polymerase with reverse transcriptase activity marketed by Perkin Elmer Cetus, Inc. Typically, the genomic RNA template is heat degraded during the first denaturation step after the initial reverse transcription step leaving only DNA template. Suitable polymerases for use with a DNA template include, for example, *E. coli* DNA polymerase I or its Klenow fragment, T4 DNA polymerase, Tth polymerase, and *Taq* polymerase, a heat-stable DNA polymerase isolated from *Thermus aquaticus* and commercially available from Perkin Elmer Cetus, Inc. The latter enzyme is widely used in the amplification and sequencing of nucleic acids. The reaction conditions for using *Taq* polymerase are known in the art and are described in Gelfand, 1989, PCR Technology, *supra*.

Allele Specific PCR

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Allele-specific PCR differentiates between target regions differing in the presence of a polymorphism. PCR amplification primers are chosen which bind only to certain alleles of the target sequence. This method is described by Gibbs, *Nucleic Acid Res*. 17:12427-2448 (1989).

Allele Specific Oligonucleotide Screening Methods

Further diagnostic screening methods employ the allele-specific oligonucleotide (ASO) screening methods, as described by Saiki et al., *Nature* 324:163-166 (1986). Oligonucleotides with one or more base pair mismatches are generated for any particular allele. ASO screening methods detect mismatches between variant target genomic or PCR amplified DNA and non-mutant oligonucleotides, showing decreased binding of the oligonucleotide relative to a mutant oligonucleotide. Oligonucleotide probes can be designed that under low stringency will bind to both polymorphic forms of the allele, but which at high stringency, bind to the allele to which they correspond. Alternatively, stringency conditions can be devised in which an essentially binary response is obtained, i.e., an ASO corresponding to a variant form of the target gene will hybridize to that allele, and not to the wild-type allele.

Ligase Mediated Allele Detection Method

Target regions of the DNA of a test subject can be compared with target regions in unaffected and affected family members by ligase-mediated allele detection. See Landegren et al., Science 241:107-1080 (1988). Ligase may also be used to detect point mutations in the ligation amplification reaction described in Wu et al., Genomics 4:560-569 (1989). The ligation amplification reaction (LAR) utilizes amplification of specific DNA sequence using sequential rounds of template dependent ligation as described in Wu, supra, and Barany, Proc. Nat. Acad. Sci. 88:189-193 (1990).

10 Denaturing Gradient Gel Electrophoresis

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Amplification products generated using the polymerase chain reaction can be analyzed by the use of denaturing gradient gel electrophoresis. Different alleles can be identified based on the different sequence-dependent melting properties and electrophoretic migration of DNA in solution. DNA molecules melt in segments, termed melting domains, under conditions of increased temperature or denaturation. Each melting domain melts cooperatively at a distinct, base-specific melting temperature (T_m). Melting domains are at least 20 base pairs in length, and may be up to several hundred base pairs in length.

Differentiation between alleles based on sequence specific melting domain differences can be assessed using polyacrylamide gel electrophoresis, as described in Chapter 7 of Erlich, ed., *PCR Technology*, Principles and Applications for DNA Amplification, W.H. Freeman and Co., New York (1992), the contents of which are hereby incorporated by reference.

Generally, a target region to be analyzed by denaturing gradient gel electrophoresis is amplified using PCR primers flanking the target region. The amplified PCR product is applied to a polyacrylamide gel with a linear denaturing gradient as described in Myers et al., Meth. Enzymol. 155:501-527 (1986), and Myers et al., in Genomic Analysis, A Practical Approach, K. Davies Ed. IRL Press Limited, Oxford, pp. 95-139 (1988), the contents of which are hereby incorporated by reference. The electrophoresis system is maintained at a temperature slightly below the Tm of the melting domains of the target sequences.

In an alternative method of denaturing gradient gel electrophoresis, the target sequences may be initially attached to a stretch of GC nucleotides, termed a GC clamp, as described in Chapter 7 of Erlich, supra. Preferably, at least 80% of the nucleotides in the GC clamp are either guanine or cytosine. Preferably, the GC clamp is at least 30 bases long. This method is particularly suited to target sequences with high T_m 's.

Generally, the target region is amplified by the polymerase chain reaction as described above. One of the oligonucleotide PCR primers carries at its 5' end, the GC clamp region, at least 30 bases of the GC rich sequence, which is incorporated into the 5' end of the target region during amplification. The resulting amplified target region is run on an electrophoresis gel under denaturing gradient conditions as described above. DNA fragments differing by a single base change will migrate through the gel to different positions, which may be visualized by ethidium bromide staining.

Temperature Gradient Gel Electrophoresis

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Temperature gradient gel electrophoresis (TGGE) is based on the same underlying principles as denaturing gradient gel electrophoresis, except the denaturing gradient is produced by differences in temperature instead of differences in the concentration of a chemical denaturant. Standard TGGE utilizes an electrophoresis apparatus with a temperature gradient running along the electrophoresis path. As samples migrate through a gel with a uniform concentration of a chemical denaturant, they encounter increasing temperatures. An alternative method of TGGE, temporal temperature gradient gel electrophoresis (TTGE or tTGGE) uses a steadily increasing temperature of the entire electrophoresis gel to achieve the same result. As the samples migrate through the gel the temperature of the entire gel increases, leading the samples to encounter increasing temperature as they migrate through the gel. Preparation of samples, including PCR amplification with incorporation of a GC clamp, and visualization of products are the same as for denaturing gradient gel electrophoresis.

Single-Strand Conformation Polymorphism Analysis

Target sequences or alleles at the IGF-1R locus can be differentiated using single-strand conformation polymorphism analysis, which identifies base differences by alteration in electrophoretic migration of single stranded PCR products, as described in Orita et al., *Proc. Nat. Acad. Sci.* 85:2766-2770 (1989). Amplified PCR products can be generated as described above, and heated or otherwise denatured, to form single stranded amplification products. Single-stranded nucleic acids may refold or form secondary structures which are partially dependent on the base sequence. Thus, electrophoretic mobility of single-stranded amplification products can detect base-sequence difference between alleles or target sequences.

Chemical or Enzymatic Cleavage of Mismatches

Differences between target sequences can also be detected by differential chemical cleavage of mismatched base pairs, as described in Grompe et al., Am. J. Hum. Genet. 48:212-222 (1991). In another method, differences between target sequences can be detected by enzymatic cleavage of mismatched base pairs, as described in Nelson et al., Nature Genetics 4:11-18 (1993). Briefly, genetic material from an animal and an affected family member may be used to generate mismatch free heterohybrid DNA duplexes. As used herein, "heterohybrid" means a DNA duplex strand comprising one strand of DNA from one animal, and a second DNA strand from another animal, usually an animal differing in the phenotype for the trait of interest. Positive selection for heterohybrids free of mismatches allows determination of small insertions, deletions or other polymorphisms that may be associated with IGF-1R polymorphisms.

25 Non-gel Systems

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Other possible techniques include non-gel systems such as TAQMANTM (Perkin Elmer). In this system oligonucleotide PCR primers are designed that flank the mutation in question and allow PCR amplification of the region. A third oligonucleotide probe is then designed to hybridize to the region containing the base subject to change between different alleles of the gene. This probe is labeled with fluorescent dyes at both the 5' and 3' ends. These dyes are chosen such that while in this proximity to each other the fluorescence of

one of them is quenched by the other and cannot be detected. Extension by Taq DNA polymerase from the PCR primer positioned 5' on the template relative to the probe leads to the cleavage of the dye attached to the 5' end of the annealed probe through the 5' nuclease activity of the Taq DNA polymerase. This removes the quenching effect allowing detection of the fluorescence from the dye at the 3' end of the probe. The discrimination between different DNA sequences arises through the fact that if the hybridization of the probe to the template molecule is not complete, i.e., there is a mismatch of some form, the cleavage of the dye does not take place. Thus only if the nucleotide sequence of the oligonucleotide probe is completely complimentary to the template molecule to which it is bound will quenching be removed. A reaction mix can contain two different probe sequences each designed against different alleles that might be present thus allowing the detection of both alleles in one reaction.

Yet another technique includes an Invader Assay which includes isothermic amplification that relies on a catalytic release of fluorescence.

Non-PCR Based DNA Diagnostics

The identification of a DNA sequence linked to IGF-1R can be made without an amplification step, based on polymorphisms including restriction fragment length polymorphisms in an animal and a family member. Hybridization probes are generally oligonucleotides which bind through complementary base pairing to all or part of a target nucleic acid. Probes typically bind target sequences lacking complete complementarity with the probe sequence depending on the stringency of the hybridization conditions. The probes are preferably labeled directly or indirectly, such that by assaying for the presence or absence of the probe, one can detect the presence or absence of the target sequence. Direct labeling methods include radioisotope labeling, such as with ³²P or ³⁵S. Indirect labeling methods include fluorescent tags, biotin complexes which may be bound to avidin or streptavidin, or peptide or protein tags. Visual detection methods include photoluminescents, Texas red, rhodamine and its derivatives, red leuco dye and 3,3',5,5'-tetramethylbenzidine (TMB), fluorescein, and its derivatives, dansyl, umbelliferone and the like or with horse radish peroxidase, alkaline phosphatase and the like.

Hybridization probes include any nucleotide sequence capable of hybridizing to the mouse chromosome where IGF-1R resides, and thus defining a genetic marker linked to IGF-1R, including a restriction fragment length polymorphism, a hypervariable region, repetitive element, or a variable number tandem repeat. Hybridization probes can be any gene or a suitable analog. Further suitable hybridization probes include exon fragments or portions of cDNAs or genes known to map to the relevant region of the chromosome.

Preferred tandem repeat hybridization probes for use according to the present invention are those that recognize a small number of fragments at a specific locus at high stringency hybridization conditions, or that recognize a larger number of fragments at that locus when the stringency conditions are lowered.

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One or more additional restriction enzymes and/or probes and/or primers can be used. Additional enzymes, constructed probes, and primers can be determined by routine experimentation by those of ordinary skill in the art and are intended to be within the scope of the invention.

Although the methods described herein may be in terms of the use of a single restriction enzyme and a single set of primers, the methods are not so limited. One or more additional restriction enzymes and/or probes and/or primers can be used, if desired. Indeed in some situations it may be preferable to use combinations of markers giving specific haplotypes. Additional enzymes, constructed probes and primers can be determined through routine experimentation, combined with the teachings provided and incorporated herein. Stand alone software as well as web-based software are available that allows the user to identify other restriction mapping sites in the DNA sequence, e.g., http://www.restrictionmapper.org/.

According to the invention, polymorphisms in the IGF-1R gene have been identified which have been associated with reproductive longevity and/or sustained performance under stress. The presence or absence of the markers, in one embodiment may be assayed by PCR-RFLP analysis using the restriction endonucleases and amplification primers may be designed using analogous human, mouse, or other IGF-1R sequences due to high homology in the region surrounding the polymorphisms, or may be designed using known IGF-1R gene sequence data as exemplified in Genbank or even designed from sequences obtained from linkage data from closely surrounding genes based

upon the teachings and references herein. The sequences surrounding the polymorphism will facilitate the development of alternate PCR tests in which a primer of about 4-30 contiguous bases taken from the sequence immediately adjacent to the polymorphism is used in connection with a polymerase chain reaction to greatly amplify the region before treatment with the desired restriction enzyme. The primers need not be the exact complement; substantially equivalent sequences are acceptable. The design of primers for amplification by PCR is known to those of skill in the art and is discussed in detail in Ausubel (ed.), "Short Protocols in Molecular Biology, Fourth Edition" John Wiley and Sons 1999. The following is a brief description of primer design.

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Primer Design Strategy

Increased use of polymerase chain reaction (PCR) methods has stimulated the development of many programs to aid in the design or selection of oligonucleotides used as primers for PCR. Four examples of such programs that are freely available via the Internet are: PRIMER by Mark Daly and Steve Lincoln of the Whitehead Institute (UNIX, VMS, DOS, and Macintosh), Oligonucleotide Selection Program (OSP) by Phil Green and LaDeana Hiller of Washington University in St. Louis (UNIX, VMS, DOS, and Macintosh), PGEN by Yoshi (DOS only), and Amplify by Bill Engels of the University of Wisconsin (Macintosh only). Generally these programs help in the design of PCR primers by searching for bits of known repeated-sequence elements and then optimizing the T_m by analyzing the length and GC content of a putative primer. Commercial software is also available and primer selection procedures are rapidly being included in most general sequence analysis packages.

25 Sequencing and PCR Primers

Designing oligonucleotides for use as either sequencing or PCR primers requires selection of an appropriate sequence that specifically recognizes the target, and then testing the sequence to eliminate the possibility that the oligonucleotide will have a stable secondary structure. Inverted repeats in the sequence can be identified using a repeat-identification or RNA-folding program such as those described above (see prediction of Nucleic Acid Structure). If a possible stem structure is observed, the sequence of the

primer can be shifted a few nucleotides in either direction to minimize the predicted secondary structure. The sequence of the oligonucleotide should also be compared with the sequences of both strands of the appropriate vector and insert DNA. Obviously, a sequencing primer should only have a single match to the target DNA. It is also advisable to exclude primers that have only a single mismatch with an undesired target DNA sequence. For PCR primers used to amplify genomic DNA, the primer sequence should be compared to the sequences in the GenBank database to determine if any significant matches occur. If the oligonucleotide sequence is present in any known DNA sequence or, more importantly, in any known repetitive elements, the primer sequence should be changed. Depending on the desired test conditions, the sequences of the primers should be designed to provide for both efficient and faithful replication of the target nucleic acid. Methods of PCR primer design are common and well known in the art. (Rychlik, W. (1993) In White, B. A. (ed.), Methods in Molecular Biology, Vol. 15, pages 31-39, PCR Protocols: Current Methods and Applications. Humania Press, Inc., Totowa, N.J.).

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The methods and materials of the invention may be used as the basis to search for polymorphisms in the IGF-1R gene of species that are associated with reproductive longevity and sustained performance under stress. This would allow uses to genetically type individual animals by detecting genetic differences in those animals. For instance, a sample of mouse genomic DNA may be evaluated by reference to one or more controls to determine if a polymorphism in the IGF-1R gene is present. Preferably, RFLP analysis is performed with respect to the mouse IGF-1R gene, and the results are compared with a control. The control is the result of a RFLP analysis of the mouse IGF-1R gene of a different mouse where the polymorphism of the mouse IGF-1R gene is known. Similarly, the IGF-1R genotype of a mouse may be determined by obtaining a sample of its genomic DNA, conducting RFLP analysis of the IGF-1R gene in the DNA, and comparing the results with a control. Again, the control is the result of RFLP analysis of the IGP-1R gene of a different mouse. The results genetically type the mouse by specifying the polymorphism(s) in its IGF-1R genes. Finally, genetic differences among mice can be detected by obtaining samples of the genomic DNA from at least two mice, identifying the presence a polymorphism in the IGF-1R gene, and comparing the results.

Such assays are useful for identifying the genetic markers relating reproductive longevity and the ability to sustained stress factors such as lactation and pregnancy, as discussed above and for the general scientific analysis of mouse genotypes' and phenotypes'.

The examples and methods herein disclose certain genes which have been identified to have a polymorphism which is associated either positively or negatively with a beneficial trait that will have an effect on performance under stress in animals, such as cattle, birds, and aquatic species, such as shrimp carrying this polymorphism. The identification of the existence of a polymorphism within a gene is often made by a single base alternative that results in a restriction site in certain allelic forms. A certain allele, however, as demonstrated and discussed herein, may have a number of base changes associated with it that could be assayed for which are indicative of the same polymorphism (allele). Further, other genetic markers or genes may be linked to the polymorphisms disclosed herein so that assays may involve identification of other genes or gene fragments, but which ultimately rely upon genetic characterization of animals for the same polymorphism. Any assay which sorts and identifies animals based upon the allelic differences disclosed herein are intended to be included within the scope of this invention.

Linkage Analysis

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Diagnostic screening may be performed for polymorphisms that are genetically linked to a phenotypic variant in IGF-1R activity or expression, particularly through the use of microsatellite markers or single nucleotide polymorphisms (SNP). The microsatellite or SNP polymorphism itself may not be phenotypically expressed, but is linked to sequences that result in altered activity or expression. Two polymorphic variants may be in linkage disequilibrium, i.e., where alleles show non-random associations between genes even though individual loci are in Hardy-Weinberg equilibrium.

Linkage analysis may be performed alone, or in combination with direct detection of phenotypically evident polymorphisms. The use of microsatellite markers for genotyping is well documented. For examples, see Mansfield et al. (1994) *Genomics* 24:225-233; and Ziegle et al. (1992) *Genomics* 14:1026-1031. The use of SNPs for genotyping is illustrated in Underhill et al. (1996) *Proc. Natl. Acad. Sci. USA* 93:196-200.

Genetic linkage maps show the relative locations of specific DNA markers along a chromosome. Any inherited physical or molecular characteristic that differs among animals and is easily detectable in the laboratory is a potential genetic marker. DNA sequence polymorphisms are useful markers because they are plentiful and easy to characterize precisely. Many such polymorphisms are located in non-coding regions and do not affect the phenotype of the organism, yet they are detectable at the DNA level and can be used as markers. Examples include restriction fragment length polymorphisms (RFLPs), which reflect sequence variations in DNA sites or differences in the length of the product, which can be cleaved by DNA restriction enzymes, microsatellite markers, which are short repeated sequences that vary in the number of repeated units, single nucleotide polymorphisms (SNPs), and the like.

The "linkage" aspect of the map is a measure of how frequently two markers are inherited together. The closer the markers are to each other physically, the less likely a recombination event will fall between and separate them. Recombination frequency thus provides an estimate of the distance between two markers. The value of the genetic map is that an inherited trait can be located on the map by following the inheritance of a DNA marker present in affected animals, but absent in unaffected animals, even though the molecular basis for the trait may not yet be understood.

SNPs are generally biallelic systems, that is, there are two alleles that a population may have for any particular marker. This means that the information content per SNP marker is relatively low when compared to microsatellite markers, which may have upwards of 10 alleles. SNPs also tend to be population-specific; a marker that is polymorphic in one population may not be very polymorphic in another. SNP markers offer a number of benefits that will make them an increasingly valuable tool. SNPs, found approximately every kilobase (see Wang et al. (1998) *Science* 280:1077-1082), offer the potential for generating high density genetic maps, which will be extremely useful for developing haplotyping systems for genes or regions of interest, and because of the nature of SNPs, they may in fact be the polymorphisms associated with the traits under study. The low mutation rate of SNPs also makes them excellent markers for studying complex genetic traits.

One of skill in the art, once a polymorphism has been identified and a correlation to a particular trait established, will understand that there are many ways to genotype animals for this polymorphism. The design of such alternative tests merely represents optimization of parameters known to those of skill in the art and is intended to be within the scope of this invention as fully described herein.

The following examples serves to better illustrate the invention described herein and are not intended to limit the invention in any way. Those skilled in the art will recognize that there are several different parameters which may be altered using routine experimentation and which are intended to be within the scope of this invention.

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Example 1 Identify polymorphisms at the daf-2 (Insulin-like growth factor-1 receptor) gene in lines of mice selected for reproductive longevity and evaluating this gene as putative candidates for DNA markers for reproductive longevity in livestock.

15 Materials and Methods

The mouse population: The original mouse population, which was established by Agriculture and Agri-Food Canada in Ottawa in 1965, was a cross between two strains of mice (P and Q). The P strain was a cross between three inbred lines (C3H/HeJ, C57BL/6J, CBA/J, SWR/J) and the Q was Falconer's strain, which had a substantial heterogeneous background (Garnett and Falconer 1975). Ancestry of the Q strain goes back to 1948, with a large contribution from the 'J' stain (Falconer 1973). The 'J' strain was a heterogeneous population of mixed origin, which was made from crosses between Bateman's highlactation line, Goodale's and MacArthur's large body weight selected lines, and four mutant stocks with the C57-BL inbred line as part of their ancestry (Brown and Falconer 1960). This population 'was about as close as one could get with laboratory mice to a natural random-bred population' (Brown and Falconer 1960). Several strains were derived from the J stock, including Falconer's control line (JC), an inbred line (JU), and a high litter size selected line (JH). The JC and JU lines constituted half of the ancestry of the Q strain. The other half was from crosses between Goodale's and MacArthur's large body weight selected lines (that had contributed to the J stock), MacArthur's small body weight selected line, JH, and a line that derived from the J stock and had been selected for high growth rate on a restricted diet (Falconer 1960). The four inbred lines and two of the lines that contributed to the Q strain (MacArthur's small body weight selected line (SM/J) and

Goodale's large body weight selected line (LG/J)), are currently maintained at the Jackson Laboratories, Bar Harbor, Maine. The contribution of so many strains to this colony, which is the only non-inbred mouse model in the world selected for reproductive longevity, was important for ensuring that the base population was heterozygous at many loci.

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Prior to the implementation of the selection program for reproductive longevity, both the P and Q stocks were maintained by random mating for 23 generations (80 breeding pairs in P and 45 males and 90 females in Q) to achieve linkage equilibrium. Two lines from each of the P and Q strains were then established, each with 92 pairs of breeders. One line derived from each of the P and Q stocks was selected for nursing ability of the mother, and the other for body weight of progeny at 42 days of age. After 21 generations of selection, these four lines were crossed, and the synthetic stock was maintained by random mating for 12 generations to allow it to approach linkage equilibrium. One control (C1) and two selected lines, with (SA1) and without (SU1) standardizing litter size to 8, were established in 1982 and have been continuously selected for reproductive longevity since then (Nagai et al. 1995). Replications from each of the control and selected lines were established (C2, SA2, SU2) in 1993 using the existing lines (generation 18 of the SA1 and SU1 and generation 44 of the C1). Also, the high performing animals from the different selected lines were mated to generate a new line with a more diverse genetic background, and a sample from the control lines was used to generate a new control line. In each of the selected lines, one male and one female were caged at about eight weeks of age, and each pair was maintained in the same cage continuously until the next generation was established, using progeny from the latest parities. In the control lines, progenies from the first parity were used as breeders. The control and selected lines were maintained with 42 and 30 breeding pairs, respectively, avoiding full-sib mating (Nagai et al. 1995). Performance of the three original lines (SA1, SA2, C1) at generations 12 and 16 is reported by Nagai et al. (1995), and at generation 24 by Farid et al. (2002). The average number of days from mating to the last parturition in generation 12 was 236, 265 and 159 for lines SA1, SU1 and C1, respectively, showing that reproductive longevity was improved by 48% in the SA1 and 67% in the SU1. The corresponding values at generation 16 were 79% and 80%, and at generation 24 were 86% and 61% for the SA1 and SU1 lines, respectively. The number of parturitions during lifetime has not changed in the control line (5.34, 4.90,

5.30 at generations 12, 16 and 24, respectively), while the SA1 line showed a steady improvement: 8.63, 8.84 and 10.6 (61.6%, 80.4% and 100%). The corresponding values for the SU1 line were 79.9%, 93.0% and 83.0%.

5 Source of DNA: DNA was extracted from blood or tissue of 261 breeder males and females from the lines C1 (generation 69), C2 (generation 70), SA1 and SU1 (generation 24), and from one progeny from each of 153 families from lines C1, C2, SA1, SA2, SU1 and SU2. DNA samples from the four inbred lines that have contributed to the base population (C3H/HeJ, C57BL/6J, CBA/J, SWR/J) were obtained from the Jackson Laboratories, Bar Harbor, Maine.

Laboratory procedures: There are two sequences of the mouse insulin-like growth factor-1 receptor cDNA in Genbank (accession numbers AF056187 (SEQ ID NO:1) and XM_133508 (SEQ ID NO:3)), and sequences of most of this gene's exons and introns included in the clone RP23-378H21 (Genbank accession number AC101879) (SEQ ID NO:6). Several overlapping PCR primers were designed to cover the entire coding region of the IGF-1R gene and its 3' UTR using the Oligo 6.0 primer analysis software (Molecular Biology Insight, cascade, CO, USA). Information on a few of these primers which amplified polymorphic regions is shown in Table 1.

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Table 1. Information on the primers used to amplify polymorphic segments of the IGF-1R

ene in mice. SEQ Primer ID NO name		Sequence (5'-3')	Location	MgCl2 (mM)	Anneal. temp.	Size, bp
8	PSEQDF	GGAGATCATCGGCAGCATCAAG	Exon 21	2.5	(°C) 58.0	216or 204
9	PSEQDR	GCCATTCTCAGCCTTGTGTCC	Exon 21	7		•
10	PSECAF	GCATGTGCTGGCAGTATAACC	Exon 21	1.5	58.5	634
11	PSECAR	CAGAGGCCCATGTCAGTTAAG	3' UTR	7		·
12	PSEQ16F	AGAGTGGCCATCAAGACGGTA	Exon 16	2.0	58.5	486
13	PSEQ16R	GGCCTCAGAGACCGGAGAT	Exon 17	7		

PCR amplifications were performed in 50 µL volumes containing (final concentration) 0.1% Tween 20, 1 x PCR buffer, 1.5-2.0 mM MgCl₂, 0.2 mM each dNTP,

400 nM each primer, 2 units of *Taq* polymerase (Roche) and 100 ng template DNA. The thermal cycler was set at 95°C for 2 min followed by 34 cycles at 94°C for 1 min, 55-67°C (depending on the primer) for 1 min, 72°C for 1 min and a final 9 min extension at 72°C. Long fragments were amplified using PCR cocktails similar to those explained above, except using 0.35 mM of each dNTP and 2.5 units of Long-Range *Taq* polymerase (Roche). Thermal cycler was set at an initial 2 min denaturation at 95°C, followed by 10 cycles of 94°C for 10 sec, 55-67°C for 30 sec and 68°C for 10 min. The next 20 cycles consisted of 94°C for 10 sec, annealing at 55-67°C for 30 sec, elongation at 68°C for 10 min plus an additional 20 sec for each new cycle and a final 9 min extension at 68°C.

Genotyping for the 12 bp deletion in exon 21 was performed using the GenScan option of an ABI 377 automated DNA sequencer. Two primers flanking the deletion were designed. The Hex Amidite label was placed on the forward primer. Since the deletion was from 3896 to 3907, the PCR product was 216 bp in the wild type (4002-3786) or 204 bp for the deletion. The PCR cocktail contained 1.25 μL of a 10X buffer, 1.25 μL of a 25mM MgCl₂, 1.0 μL of a 1.25 mM dNTPs, 5 pmol of each primer, 0.2 μL of a 5 U/μL Amplitaq gold polymerase, 25 ng of DNA and water to 12.5 μL total volume. Thermal cycler conditions were 95°C for 8 minutes initial denaturation, followed by 30 cycles of 95°C for 30 sec, 58°C for 30 sec, 72°C for 60 sec, and a final extension of 72°C for 30 minutes. PCR products were maintained at 6°C until processed. One μL of PCR products were loaded into the sequencer.

Data analysis:

Conformation of genotype frequencies to Hardy-Weinberg equilibrium was tested using the GENEPOP computer package (http://wbiomed.curtin.edu.au/genepop) using the default options (1000 dememorisation, 100 batches and 1000 iterations). The program uses the Markov chain method to estimate the exact Hardy-Weinberg probability without bias (Guo and Thompson, 1992). The probability of rejecting H₀, i.e., genotype frequencies are in Hardy-Weinberg equilibrium and the standard error of this estimate were computed. When standard errors were larger than 0.01, the data were re-analysed using a larger number of batches. This program does not perform any test when a locus is monomorphic or quasi monomorphic (two alleles, but one is represented only once).

Pairwise tests for homogeneity of allele and genotype frequency distributions were also performed using the GENEPOP computer package which follows the Raymond and Rousset (1995) method. The hypotheses tested were that allele and genotype distributions were independent of lines (no difference between lines). An unbiased estimate of the Fisher's exact test on contingency tables is performed using the Markov chain method (1000 dememorisation, 100 batches and 1000 iterations). The program computes the probability of being wrong when H_o is rejected. Rare alleles (frequency of less than 5%) were not pooled together prior to the above tests. F_{IS} statistics, as the measures of inbreeding within each line (Wright, 1943, 1978), were computed for each polymorphic site in every line using the GENEPOP computer program.

Results

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<u>Polymorphism:</u> A total of 4434 bp of the IGF-1R gene, consisting of exons 2, 3, 9, 10, 12, 13, 14, 15, 16, 17 and 21 (2344 bp) and introns 10, 12, 13, 14 and 16 (2090 bp) in five to seven individuals from each of the three main lines (C1, SA1, SU1) were sequenced. No polymorphism was detected in exons 2, 3, 9, 10, 12, 13, 14, 15, 16, 17 or in introns 10, 12, 13 and 14. The following polymorphic sites have been detected:

Site A: A 'G' to 'A' substitution (GGTC to GATC) was detected in intron 16 of the gene. The 486 bp PCR product, spanning exons 16 and 17 and intron 16, was cut into 454 and 32 bp fragments (A₁ allele) by the enzyme *DpnII* (†GATC). This nucleotide substitution resulted in the creation of a new recognition site for this enzyme, which cleaved the 454 bp fragment into 328 and 125 bp fragments (A₂ allele). In addition, sequence information revealed a 'G' nucleotide insertion in intron 16, 153 bp 5' to the above point mutation, but no restriction enzyme was found for discriminatory typing of this insertion.

Site B: An *Hpall* (CTCGG) polymorphism was detected as a result of an 'A' to 'G' substitution at position 3876 in exon 21 (CCAG to CCGG). The enzyme had one recognition site in the PCR product (373 and 261 bp fragments, B₁ allele) and the nucleotide substitution resulted in an additional recognition site for the enzyme (373, 134 and 127 bp fragments, B₂ allele). This is a silent mutation, as both CCA and CCG code for the amino acid proline. The marker for coping with pregnancy and lactation stress in mice

is the sequence containing the 'A' nucleotide at position 3876 of the mouse IGF-1R gene, identified by the 373/261 bp fragments (B₁ allele). Since the substitution is 20 base pairs upstream from the 12 base pair deletion, the 261 bp and 127 bp bands will shift by 12 base pairs when animals are homozygous or heterozygous for the deletion allele (D₂). As is known in the art, however, restriction patterns are not exact determinants of the sizes of fragments and are only approximate.

Site D: Site D: A 12 bp deletion was detected 20 bp 3' to the site B in exon 21 (positions 3896-3907 of the IGF-1R gene cDNA, Genbank accession number AF056187, SEQ ID NO:1). This 12 bp fragment (tggagatggage) (SEQ ID NO:20) appears twice in tandem (D₁ allele) in or only once (D₂ allele) in this region, resulting in the deletion of four amino acids (leucine, glutamic acid, methionine, and glutamic acid) from the IGF-1R protein. One IGF-1R sequence (Genbank accession number AF056187, SEQ ID NO:1) has two copies of this fragment while two others (Genbank accession numbers XM_133508 (SEQ ID NO:3) and AC101879 (, SEQ ID NO:6) have one copy.

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Allele and genotype frequency distributions: Although sites A and B are approximately 22 kb apart, all 153 juveniles and 261 breeders had exactly the same genotypes at these two sites, constituting only two alleles (A_1 and A_2). Replicate lines of juvenile mice were not different from the main lines for allele or genotype frequencies at site A. The frequency of A_1 allele in breeders from the SU1 line (0.84) was significantly greater than those in the other three lines (0.48, 0.62, 0.63, Tables 2 and 3). A similar pattern was observed in the juveniles, where frequencies of the A_1 allele in the SU1 and SU2 lines (0.83 and 0.89) were significantly greater than those in SA1 (0.55), SA2 (0.46), C1 (0.48) and C2 (0.61) lines (Tables 4 and 5). Frequencies of A_1 allele in the C1 line were similar in breeders and juveniles (0.48), and were smaller than those in the C2 line in breeders (0.67, P<0.01) and juveniles (0.61, NS). Frequency of A_1 allele in selected and control lines in which litter size was not standardized (SU1, SU2, C2) was greater than that in the lines in which litter size was standardized (SA1, SA2, C1) in both breeders and juveniles, suggesting that the A_1 allele was possibly selected for under high levels of maternal stress.

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The frequency of the A_1A_1 genotype in breeders from SU1 line (0.71) was significantly greater than in other lines, which ranged between 0.23 (C1) and 0.47 (C2)

(Tables 2 and 3). Juveniles from the SU1 and SU2 lines had greater frequencies of the A₁A₁ genotype (0.75 and 0.77) and lower frequencies of the A₂A₂ genotype (0.10 and 0.0) than the other four lines, in which the frequencies of A₁A₁ ranged from 0.17 to 0.44 and frequencies of A₂A₂ ranged from 0.22 to 0.26 (Tables 4 and 5). Differences in genotype frequencies between SU1 and SU2 and the other lines were all significant, except for SU1 and C2 that approached significance (P=0.079). Genotype frequency distributions conformed to Hardy-Weinberg proportions in all the lines. All four inbred lines (C3H/HeJ, C57BL/6J, CBA/J, SWR/J) had the A₁A₁ genotype at site A and the B₁B₁ genotype at site B, indicating that the A₂ and B₂ alleles must have been introduced into the base population by the O-strain.

No D_2 allele was detected in any of the control mice. The frequency of the D_2 allele (deletion) ranged from 0.10 to 0.19 in the selected lines in the juveniles and breeders. The selected lines within breeder and juvenile groups had comparable allele and genotype frequencies at site D. All selected lines had significantly different allele and genotype frequency distributions compared with the control lines in which the D_1 allele was fixed (Tables 6, 7, 8, 9). Replicate lines of juvenile mice were not different from the main lines for allele or genotype frequencies at site D. Genotype frequency distributions conformed to Hardy-Weinberg proportions in all the lines, except in juveniles from the SA1 line, which was deficient in heterozygotes (F_{IS} =+0.449, Table 4). High proportions of the D_2 allele appeared in the heterozygous state (0.179 to 0.385), and low proportions (0.0 to 0.107) were in homozygous form in all the selected lines, which is expected from a population in Hardy-Weinberg equilibrium in which one allele has a low frequency. The C57BL/6J had the D_2D_2 genotype, but the other three inbred lines had the D_1D_1 genotype.

Only six of the 10 possible genotypes were present in the population when the joint distribution of A and D sites was considered (Tables 10, 12), indicating the presence of three of the four possible haplotypes (A_1D_1, A_1D_2, A_2D_1) . Haplotype and genotype frequency distributions were significantly different among all the lines within breeder and juvenile groups, except those between replicate lines (Tables 10, 11, 12, 13). Haplotype frequency differences between selected and control lines were largely due to the absence of the A_1D_2 haplotype in the latter. Differences among non-replicate selected lines for haplotype frequency distributions were mainly the result of higher frequencies of A_1D_1

(0.69 to 0.74) and lower frequencies of A_2D_1 (0.12 to 0.18) in selected non-standardized lines compared with those in standardized selected lines, which had lower frequencies of A_1D_1 (0.28 to 0.46) and higher frequencies of A_2D_1 (0.38 to 0.52). Genotype frequencies conformed to Hardy-Weinberg proportions in all the lines in both breeders and juveniles, except in the SA1 line in juveniles, which was deficient in heterozygotes (F_{IS} =+0.341, Table 12). There was no difference between male and female breeders for allele or genotype frequencies at any of the sites (data not shown).

Discussion

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Similarities between the replicate lines for allele (haplotype) and genotype frequencies at all sites may indicate that the observed differences among non-replicate lines had happened before divergence of replicate lines from the main lines. These findings also imply that the size of the lines was great enough to make genetic drift a negligible force in changing the genetic profile of the lines in the last 8 generations of the selected lines (generations 18 to 24) and 26 generations of the controls (generations 44 to 69). The observed differences among the lines for allele frequency distributions can thus be largely attributed to the selection pressure applied to each line.

The finding that the A₁ allele had a significantly greater frequency in breeder animals in which litter size was not standardized to 8 (selected and control lines) may suggest that although this gene has not been under selection pressure for reproductive longevity, the A₁ allele may be linked to a QTL that has a favorable effect on maternal stress. Most female mice conceive while still nursing, which imposes a great pressure on them, and the effect will be more pronounced when litter size is large. It seems that the A₁ allele is associated with animals that may be able to better cope with such a stress. This finding has some ramifications in the livestock industry, such as swine and dairy cattle, where lactation and pregnancy often coincide. This is the first evidence showing that such a characteristic is genetically controlled.

The results from site D provide a different picture than of site A. The absence of the D_2 allele (deletion) in the control lines, and the similarity between all the selected lines for the allele and genotype frequencies within breeders and juveniles may suggest that the D_2 allele (or an allele which is linked to D_2) had a negative effect on early reproduction, and

has therefore been eliminated from the control lines. This conclusion is based on three notions. First, the frequency of the D₂ allele in the original population was expected to be at least 0.125, because C57BL/6J with the D2D2 genotype provided 1/8 of the genes to the original population, and this line had also contributed to the Q-strain. The effects of 21 generations of selection for nursing ability of the mother and body weight of progeny that was applied to the original population before the establishment of the base population for this experiment is not known. Assuming, however, that the frequency of the D2 allele was not drastically changed, it is unlikely that the D2 allele with such a frequency had not been included in the first generation of the control line merely by chance. Second, absence of the D₂ allele in the control lines was not because of the small number of mice that were genotyped. The probability (a) that an allele with the frequency of Y or less in a population falls into a sample of size n (i.e., 2n alleles) is $\log (1-\alpha)=2n \log (1-Y)$. Setting n=25, which was the smallest sample size of the control lines in juveniles, and Y=0.10 (the smallest estimate of the D_2 frequency in any line) will result in α =0.994, i.e., there is at least 99% probability that the D₂ allele with a frequency of 0.10 would be included in a sample of size 25. Combining the two control lines of juveniles will increase this probability to 99.99%. The total number of control mice tested (juveniles and breeders) was 217, suggesting that the D₂ allele certainly does not exist in the control lines. Third, in the control lines, the male is removed from the cage 14 to 17 days following pairing. Replacement mice in the control lines are thus selected from females that conceived within the first 14 to 17 days after exposure to a male. The control lines, therefore, have been under mild selection for early reproduction. Although more studies are needed, it seems logical to believe that deletion of four amino acids from the IGF-1R would have some negative effect on the function of this polypeptide. The only explanation for the D2 allele to have a frequency of 0.10 to 0.20 in the selected lines is that this allele, or one which is linked to it, had a positive effect on reproduction at a later age. In addition, the D2 allele was largely in the heterozygous state, which will mask any negative effect of the allele.

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The D and A sites are only 20 bp apart, and thus the likelihood of a crossing-over between them is very slim. Differences between lines for allele frequencies at sites A and D should be sought in the origin of the haplotypes. Since the only source of the A_2 allele was the Q strain, and there was no A_2D_2 haplotype in the population, it seems logical to assume

that the A₂D₁ haplotype originated from the Q-strain and the A₁D₂ haplotype originated from C57BL/6J (the only inbred line carrying the A₁D₂ haplotype). Line C57BL/6J had a minor contribution to the Q strain, indicating that the Q strain might have carried the A₁D₂ haplotype as well. The A₁D₁ haplotype originated from the other three inbred lines as well as from the Q-strain. It seems reasonable to conclude that the A₁D₂ haplotype, which originated from the C57BL/6J line and has been eliminated from the control lines, is a QTL with a negative effect on early reproduction and a positive effect on reproductive longevity. The A₂D₁ haplotype that originated from the Q strain and had high frequencies in non-standardized lines (SU1, SU2, C2) may be a QTL that has been selected for under maternal pressure (large litter size, high milk production, pregnancy).

F_{IS} is a measure of the inbreeding coefficient of individuals in a subdivided population due to nonrandom mating, or inbreeding of an individual relative to the subpopulation to which it belongs (Wright, 1943, 1978; Nei, 1973; Hartl and Clark, 1989). When mating is at random in a sub-population, F_{IS} is equal to zero. Positive F_{IS} values indicate within sub-populations inbreeding (more homozygosity than expected) due to mating between relatives. Negative F_{IS} values show less homozygosity than expected from a population at Hardy-Weinberg equilibrium. Conformation of genotype frequency distributions to Hardy-Weinberg values and small F_{IS} estimates indicate that mating between animals with respect to sites A and D and their joint distribution has been at random in all the lines except SA1 in juveniles. This is expected in view of the fact that the effect of individual alleles on phenotype (reproductive longevity) has not been visible.

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Many lines of mice contributed to the base population, making it a heterogeneous stock with many segregating loci upon which selection pressure has been applied for 24 generations. The fact that allele frequencies at sites A and D in the entire sample were 0.63 and 0.89 in juveniles and 0.63 and 0.94 in breeders, respectively, point to the heterogeneity of the population at the present time. The observed genetic variability makes this colony unique.

Table 2. Distribution of allele and genotype frequencies at site A^1 at the IGF-1R locus in breeder mice, test for Hardy-Weinberg equilibrium and F_{IS} estimates by line and sex.

breeder mice, te	st for Har	dy-Wein	berg equi	llibrium	and Pis	esumates	by line a	ilu sex.	
Line	ne Sex Allele frequency			Genotype frequency			No. of mice	H-W prob	F _{IS}
		. A _l	A ₂	A_1A_1	A ₂ A ₂	A ₁ A ₂			
SAI	F	0.648	0.352	0.407	0.111	0.481	27		'
Selected (standardized)	M	0.603	0.397	0.379	0.172	0.448	29		
•	Total	0.625	0.375	0.393	0.143	0.464	56	1.00	0.019
SU1	F	0.881	0.119	0.809	0.048	0.143	21		
Selected (Non- standardized)	М	0.810	0.190	0.619	0.000	0.381	21		
	Total	0.845	0.155	0.714	0.024	0.262	42	1.00	0.011
Cl	F	-0.488	0.512	0.190	0.214	0.595	42		
Control (standardized)	M	0.476	0.523	0.262	0.310	0.429	42		
	Total	0.482	0.518	0.226	0.262	0.512	84	1.00	-0.019
C2	F	0.700	0.300	0.500	0.100	0.400	40		
Selected (non- standardized)	M	0.641	0.359	0.436	0.154	0.410	39		
	Total	0.671	0.329	0.468	0.127	0.405	79	0.45	0.089
Total		0.628	0.372	0.414	0.157	0.429	261	0.99	

¹⁻Site A is a 'G' to 'A' substitution in intron 16, which is in linkage disequilibrium with an 'A' to 'G' substitution in exon 21 (site B).

Table 3. Pairwise comparison of the lines for allele frequency (above diagonal) and genotype frequency (below diagonal) for site A in breeder mice.

Sellotype frequence	y (below diagonal) lo			
Line	SA1	SU1	C1	C2
SA1	<u>-</u>	0.001	0.020	0.436
SUI	0.001	-	0.000	0.003
Cl	0.020	0.000	-	0.002

C2	0.446	0.006	0.001	

Table 4. Distribution of allele and genotype frequencies at site A of the IGF-1R locus in juveniles, test for Hardy-Weinberg equilibrium and F_{IS} estimates by line.

uveniles, test for i	Taruy- W	childerg c	10111011011	min 1 12 0	3137274140	-,			
Line		Allele frequency		Genotyp	Genotype frequency			H-W	F _{IS}
		A ₁	A ₂	A ₁ A ₁	A ₂ A ₂	A ₁ A ₂	of mice	prob.	
Selected	SA1	0.552	0.448	0.345	0.241	0.414	29	0.45	0.180
(Standardized)	SA2	0.458	0.542	0.167	0.250	0.583	24	0.68	-0.154
Selected	SUI	0.825	0.175	0.750	0.100	0.150	20	0.07	0.500
(Non-standardized)	SU2	0.885	0.115	0.769	0.000	0.231	26	1.00	-0.111
Control (standardized)	CI	0.481	0.519	0.222	0.259	0.519	27	1.00	-0.020
Control (non-standardized)	C2	0.611	0.389	0.444	0.222	0.333	27	0.13	0.316
Total		0.627	0.373	0.438	0.183	0.379	153	0.46	0.109

Table 5. Pairwise comparison of the lines for allele frequency (above diagonal) and

genotype frequency (below diagonal) for site A in juveniles.

enotype rrequ	ichey (octow die	igoliai) foi site A	in javonnosi			
Line	SA1	SA2	SUI	SU2	Ci	C2
SA1	-	0.428	0.005	0.000	0.570	0.567
SA2	0.451		0.000	0.000	0.845	0.160
SU1	0.014	0.001	· -	0.545	0.001	0.038
SU2	0.000	0.000	0.592	-	0.000	0.001
C1	0.588	0.836	0.002	0.000	-	0.246
C2	0.617	0.189	0.079	0.004	0.279	-

Table 6. Distribution of allele and genotype frequencies of the deletion¹ at the IGF-1R locus in breeder mice, test for Hardy-Weinberg equilibrium and F_{is} estimates by line and

sex. Line	Sex	Allele frequency		Genotype frequency			H-W Prob	Fis
		D_1	D ₂	$D_l D_l$	D_2D_2	D_1D_2		
SAI	F	0.796	0.204	0.593	0.000	0.407		
Selected (standardized)	М	0.862	0.138	0.758	0.034	0.207		
·	Total	0.830	0.170	0.679	0.018	0.304	1.00	-0.069
sui	F	0.929	0.071	0.857	0.000	0.143		
Selected (Non- standardized)	М	0.857	0.143	0.714	0.000	0.286		
	Total	0.893	0.107	0.786	0.000	0.214	1.00	-0.108
CI	F	1.000	0.000	1.000	0.000	0.000		
Control (standardized)	М	1.000	0.000	1.000	0.000	0.000		

	Total	1.000	0.000	1.000	0.000	0.000	-	-
C2 Selected (non- standardized)	F	1.000	0.000	1.000	0.000	0.000		
	M	1.000	0.000	1.000	0.000	0.000		
	Total	1.000	0.000	1.000	0.000	0.000		-
Total		0.946	0.054	0.897	0.003	0.100	1.00	<u> </u>

¹⁻A 12 bp deletion (D₂ allele) in exon 21 of the IGF-1R gene.

Table 7. Pairwise comparison of the lines for allele frequency (above diagonal) and

genotype frequency (below diagonal) for site D in breeder mice.

enotype frequency (below diagonal) for site D in breeder inice.								
Line	SA1	SU1	C1	C2				
SA1	-	0.305	0.000	0.000				
SU1	0.218	· -	0.000	0.000				
Cl	0.000	0.000	-	1.000				
C2	0.000	0.000	1.000	-				

Table 8. Distribution of allele and genotype frequencies of the deletion 1 at the IGF-1R locus (site D) in juveniles, test for Hardy-Weinberg equilibrium and F_{1S} estimates by line.

Line	Allele fre		Genotype frequency			No. of	H-W	F _{IS}
	Di	D ₂	D_iD_i	D_2D_2	D_1D_2			
SA1	0.804	0.196	0.714	0.107	0.179	28 .	0.04	0.449
SA2	0.804	0.196	0.652	0.044	0.304	23	1.00	0.055

SU1	0.900	0.100	0.800	0.000	0.200	20	1.00	-0.086
SU2	0.808	0.192	0.615	0.000	0.385	26	0.54	-0.220
Cl	1.000	0.000	1.000	0.000	0.000	27	-	-
C2	1.000	0.000	1.000	0.000	0.000	25		-
	0.886	0.114	0.799	0.027	0.174	149	0.46	0.083

Table 9. Pairwise comparison of the lines for allele frequency (above diagonal) and

genotype frequency (below diagonal) for site D in juvenile mice.

schotype ned		igonal) for site 2				
Line	SA1	SA2	SU1	SU2	C1	C2
SA1	-	1.000	0.257	1.000	0.000	0.001
SA2	1.000	-	0.261	1.000	0.001	0.001
SU1	0.333	0.250	-	0.261	0.031	0.036
SU2	1.000	1.000	0.211	-	0.001	0.001
C1	0.005	0.001	0.029	0.000	-	1.000
C2	0.005	0.001	0.032	0.000	1.000	-

Table 10. Distribution of haplotype and genotype frequencies for the joint A and D sites in breeder mice, test for Hardy-Weinberg equilibrium and F_{IS} estimates by line and sex.

Line Sex Haplotype frequency Genotype frequency

		A _i D _i	A ₁ D ₂	A ₂ D ₁	A ₁ A ₁ D ₁ D ₁	A ₁ A ₁ D ₂ D ₂	A_1A_1 D_tD_2	A_2A_2 D_1D_1	A ₁ A ₂ D ₁ D ₁	A ₁ A ₂ D ₁ D ₂	prob	Fis
SAI	F	0.444	0.204	0.352	0.185	0.000	0.222	0.111	0.296	0.185		
	M	0.466	0.138	0.396	0.172	0.034	0.172	0.172	0.414	0.034		
	Total	0.455	0.169	0.375	0.179	0.018	0.196	0.143	0.357	0.107	0.82	-0.051
SUI	F	0.810	0.071	0.119	0.667	0.000	0.143	0.047	0.143	0.000		
	М	0.667	0.143	0.190	0.429	0.000	0.190	0.000	0.286	0.095		
	Total	0.738	0.107	0.155	0.547	0.000	0.167	0.024	0.214	0.048	0.90	-0.009
Cl	F	0.488	0.000	0.512	0.190	0.000	0.000	0.214	0.595	0.000		
	M	0.476	0.000	0.524	0.262	0.000	0.000	0.310	0.428	0.000		
	Total	0.482	0.000	0.518	0.226	0.000	0.000	0.262	0.512	0.000	1.00	-0.019
C2	F	0.700	0.000	0.300	0.500	0.000	0.000	0.100	0.500	0.000		
	М	0.641	0.000	0.357	0.436	0.000	0.000	0.154	0.410	0.000		
	Total	0.671	0.000	0.329	0.468	0.000	0.000	0.127	0.405	0.000	0.45	0.089
Total		0.575	0.054	0.371	0.341	0.004	0.069	0.157	0.399	0.031	0.98	

Table 11. Pairwise comparison of the lines for haplotype frequency (above diagonal) and genotype frequency (below diagonal) for joint A and D sites in breeder mice.

C2 SU1 C1 SA1 Line . 0.000 0.000 0.000 SA1 0.000 0.000 SU1 0.000 0.000 0.000 0.000 C1

0.001

C2

0.000

0.000

Table 12. Distribution of haplotype and genotype frequencies for the joint A and D sites in inveniles, test for Hardy-Weinberg equilibrium and Fig estimates by line.

juveniles, test for Hardy-Weinberg equilibrium and F _{IS} estimates by line.											
Lines	Haploty	pe freque	ncy	Genotype frequency						H-W	F _{IS}
	$A_1 D_1$	$A_1 D_2$	A_2D_1	A_1A_1	A_1A_1	A_1A_1	A_2A_2	A_1A_2	A ₁ A ₂	prob.	
				D_1D_1	D_2D_2	D_1D_2	D_1D_1	D_1D_1	D_1D_2		
SA1	0.357	0.196	0.446	0.21	0.11	0.00	0.25	0.25	0.14	0.04	0.341
SA2	0.283	0.196	0.522	0.09	0.00	0.00	0.22	0.35	0.26	0.63	-0.048
SU1	0.725	0.100	0.175	0.55	0.00	0.20	0.10	0.15	0.00	0.17	0.218
SU2	0.692	0.192	0.115	0.46	0.00	0.31	0.00	0.15	0.08	0.68	-0.125
CI	0.481	0.000	0.519	0.22	0.00	0.00	0.26	0.52	0.00	1.00	-0.022
C2	0.620	0.000	0.380	0.48	0.00	0.00	0.24	0.28	0.00	0.08	0.423
Total	0.520	0.114	0.366	0.33	0.00	0.09	0.18	0.29	0.08	0.16	0.135

Table 13. Pairwise comparison of the lines for haplotype frequency (above diagonal) and genotype frequency (below diagonal) for joint A and D sites in juveniles.

Line	SA1	SA2	SU1	SU2	Cl	C2
SA1	•	0.696	0.001	0.000	0.001	0.001
SA2	0.749	_	0.000	0.000	0.001	0.000
SU1	0.008	0.001	-	0.415	0.000	0.010

SU2	0.001	0.000	0.442	-	0.000	0.000
C1	0.005	0.001	0.000	0.000	•	0.171
C2	0.003	0.000	0.018	0.000	0.217	-

Example 2
Identification of Polymorphisms in the IGF-1R Gene in a Line of Pigs for the Development of DNA

Animals from a single commercial operation were used to find polymorphisms in candidate genes for reproductive longevity in pigs. Sourcing all animals from a single farm should ensure a similar environment for both high and low reproductive longevity groups. Five living sows with very high parity numbers were chosen as representing high reproductive longevity and five animals culled for reproductive reasons at low parity numbers were chosen as representing low reproductive longevity.

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DNA was extracted from tissue samples from these 10 animals and the DNA used to amplify regions of candidate genes using PCR. PCR primers were designed from pig DNA sequence, or from exonic sequence of the homologous gene in other species such as mouse or human. The DNA sequence of these PCR products was then determined and the sequences compared to identify any polymorphisms. Each polymorphism was then assayed over a larger sample of animals from the same commercial population to look for evidence of association with increased reproductive longevity.

Five polymorphisms were found. Of these five, 2 were in intron 16 (SNP16i27 and SNP16i73); one in exon 8 (SNP1772); one in exon 16 (SNP3085); and one in exon 21 (SNP3757).

The polymorphism designated SNP1772, was characterized as a G/A SNP. It is a TaqI RFLP. Polymorphism SNP16i27 (position 27 from the end of exon 16) is a G/A SNP. It is an AvaII RFLP. SNP16i73 (position 73 from the end of exon 16) is a G/C SNP. It is a MnlI RFLP.

PCR-RFLP Protocol for SNP16i27

Primers used in RLFP analysis were as follow:

Primer 16 5' - CCT CCG TGA TGA AGG AGT TC - 3' (SEQ ID NO:14)

5 Primer 17 5' - TCA GTT CCA TGA TGA CCA GC - 3' (SEQ ID NO:15)

PCR was carried out using the following conditions:

	10X PCR Buffer	1.0 ul
	2mM dNTPs	1.0 ul
10	25mM MgCl ₂	1.0 ul
	5uM Primer 16	1.0 ul
	5uM Primer 17	1.0 ul
	Amplitaq Gold	0.1 ul
	QH ₂ O	3.9 ul
15	DNA	1.0 ul

Thermal Cycling conditions on the PE9700 were as follow:

94°C - 12 min

20 94°C - 30 sec

58°C - 30 sec

72°C - 30 sec

(repeated for 39 additional cycles)

25 $72^{\circ}C - 7 \min$

4°C - hold

Digested with AvaII restriction endonuclease.

30 The expected product sizes were: allele 1: 141, 122, 44; allele 2: 122, 81, 60, 44.

PCR-RFLP Protocol for SNP16i73

35 Primers used in RLFP analysis were as follow:

Primer 16 5' - CCT CCG TGA TGA AGG AGT TC - 3' (SEQ ID NO:16)

Primer 17 5' - TCA GTT CCA TGA TGA CCA GC - 3' (SEQ ID NO:17)

PCR was carried out using the following conditions:

40	10X PCR Buffer	1.0 ul
	2mM dNTPs	1.0 ul
	25mM MgCl ₂	1.0 ul
	5uM Primer 16	1.0 ul
	5uM Primer 17	1.0 ul
45	Amplitaq Gold	0.1 ul
	QH ₂ O	3.9 ul
	DNA	1.0 ul

Thermal Cycling conditions on the PE9700

94°C − 12 min

5 94°C – 30 sec

58°C - 30 sec

72°C - 30 sec

(repeat for 39 additional cycles)

10 $72^{\circ}\text{C} - 7 \text{ min}$

4°C - hold

Digested with MnII restriction endonuclease.

15 The expected product sizes were: allele 1: 241, 55, 11; allele 2: 137, 104, 55, 11.

PCR-RFLP Protocol for SNP1772

20 Primers used in RLFP analysis were as follow:

Primer 9 5' - GGA GTA TGA TGG GCA GGA T - 3' (SEQ ID NO:18)

Primer 8 5' - GAA GCA TTG GTG CGA ATG TA - 3' (SEQ ID NO:19)

PCR was carried out using the following conditions:

25	10X PCR Buffer	1.0 ul
	2mM dNTPs	1.0 ul
	25mM MgCl ₂	0.6 ul
	5uM Primer 9	1.0 ul
	5uM Primer 8	1.0 ul
30	Amplitaq Gold	0.1 ul
	QH ₂ O	4.3 ul
	DNA	1.0 ul

Thermal Cycling conditions on the PE9700

 $94^{\circ}C - 12 \min$

94°C - 30 sec

56°C - 30 sec

72°C - 30 sec

40 (repeat for 39 additional cycles)

72°C - 7 min

4°C – hold

Digested with TaqI restriction endonuclease.

The expected product sizes were: allele 1: 219; allele 2: 135, 84.

Example 3 SNP 3832

Samples from old surviving sows and from young sows culled during the first 4 parities.

996 sows from four different farms were genotyped and tested for the effect of SNP 3832 on the number of parities. Allele "2" was found to be positively associated with longevity. In average sows of the 22, 12 and 11 genotypes were culled after 7.4, 6.7 and 5.1 parities, respectively. The additive effect of SNP 3832 was estimated to be

1.11/parities/allele (P=0.004) with no dominance effect. The effect is significant, but over estimated due to the data structure.

Germany (GER): Longevity (reproduction) data from sows with known pedigree with DNA samples from their sires.

Data of over 19,000 sows, daughters of 179 sires were used in the analysis. Each sire had at least 50 daughters. There are 76 litter farms represented and the litters were from 1996 to 2001. Phenotypic performance of each sire was estimated based on the daughters' performances, and genotypic data was collected for the sires. Allele "2" found to be positively associated with longevity. SNP 3832estimated additive effect represent a contrast between homozygous sows of 38 days to culling (P=0.062).

A large number of animals were genotyped for the SNP 3832 marker. Animals carrying two copies of the "2" allele (homozygous) are expected to produce more parities and stay in the herd longer.

PCR for SNP 3832

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25 Primer 22 5' - AAG ATG AGG CCT TCC TT - 3' (SEQ ID NO:21)
Primer 23 5' - GAT CAG CAG GTC GAG GAC TG - 3' (SEQ ID NO:22)

	PCR Conditions:	
	10X PCR Buffer	1.0 ul
30	2mM dNTPs	1.0 ul
	25mM MgCl2	0.6 ul
	5uM Primer 22	1.0 ul
	5uM Primer 23	1.0 ul
	Amplitaq Gold	seem to 0.1 ul
35	QH2O	4.3 ul
	DNA	1.0 ul

Thermal Cycling conditions on the PE9700 94°C - 12 min

- 5 94°C 30 sec 58°C - 30 sec 72°C - 1 min (repeat for 34 additional cycles)
- 10 72°C 7 min 4°C - hold

Digest with Fokl

Expected product sizes: allele 1: 347; allele 2: 292, 55.

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What is claimed is:

- A method for genetically identifying an animal with respect to its potential to reproductive longevity comprising:
 obtaining a sample of genetic material from an animal; and
 assaying for the presence of a polymorphism in the insulin-like growth factor 1 receptor gene (IGF-1R), wherein the polymorphism is associated with reproductive longevity.
- 2. The method of claim 1 wherein said polymorphism is selected from the group consisting of: a single nucleotide polymorphism (SNP), a deletion, and an insertion.
 - 3. The method of claim 1 wherein the animal is selected from a group consisting of: a mouse, a pig, and a cow.
- The method of claim 1 wherein a step of assaying the polymorphism is selected from the group consisting of: direct sequencing, restriction fragment length polymorphism (RFLP) analysis, single-stranded conformation polymorphism (SSCP), PCR amplification of specific alleles, amplification of DNA target by PCR followed by a mini-sequencing assay, allelic discrimination during PCR, Genetic Bit Analysis, Pyrosequencing,
 oligonucleotide ligation assay, and analysis of melting curves.
 - 5. The method of claim 4 wherein the step of assaying the polymorphism is RFLP.
 - 6. The method of claim 4 wherein the step of assaying the polymorphism is SSCP.
- 7. The method of claim 1 wherein the step of assaying for the presence of the polymorphism comprises the steps of:
 digesting the genetic material with a restriction endonuclease that cleaves the gene in at least one place, wherein a particular restriction endonuclease pattern indicates the presence or absence of a polymorphism;
 separating the fragments obtained from the digestion;

detecting a restriction pattern generated by the fragments; and comparing the pattern with a second restriction pattern for the gene obtained by using the restriction endonuclease, wherein the second restriction pattern is associated with reproductive longevity.

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- 8. The method of claim 7 wherein said separation is by gel electrophoresis.
- The method of claim 7 further comprising:
 amplifying the gene or a portion thereof which contains at least one polymorphism, prior to
 digestion.
 - 10. The method of claim 9 wherein the amplification includes selecting a forward and a reverse sequence primer capable of amplifying a region of the gene which contains a polymorphism.

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- 11. The method of claim 1 wherein the polymorphism is identified as an A to G nucleotide substitution at position 3876 of the gene.
- 12. The method of claim 1 wherein the polymorphism is identified as a G to A20 nucleotide substitution at position 331 of the gene.
 - 13. The method of claim 1 wherein the polymorphism is a 12 base pair deletion at positions 3896-3907 of the gene.
- 25 14. The method of claim 7 wherein the restriction endonuclease is *Hpall*.
 - 15. The method of claim 7 wherein the restriction endonuclease DpnII.
 - 16. The method of claim 7 wherein the restriction endonuclease is Taql.

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17. The method of claim 7 wherein the restriction endonuclease is MnII.

- 18. The method of claim 7 wherein the restriction endonuclease is AvaII.
- 19. The method of claim 10 wherein the forward primer is SEQ ID NO:8 and wherein the reverse primer is SEQ ID NO:9.
 - 20. The method of claim 10 wherein the forward primer is SEQ ID NO:10 and wherein the reverse primer is SEQ ID NO:11.
- 10 21. The method of claim 10 wherein the forward primer is SEQ ID NO:12 and wherein the reverse primer is SEQ ID NO:13.
 - 22. The method of claim 10 wherein the forward primer is SEQ ID NO:14 and wherein the reverse primer is SEQ ID NO:15.
 - 23. The method of claim 10 wherein the forward primer is SEQ ID NO:16 and wherein the reverse primer is SEQ ID NO:17.
- 24. The method of claim 10 wherein the forward primer is SEQ ID NO:18 and wherein the reverse primer is SEQ ID NO:19.
 - 25. A method of screening animals to determine those more likely to have reproductive longevity, the method comprising:

 obtaining a biological sample from an animal; and
- assaying for the presence of a genotype in the IGF-1R gene, wherein the genotype is associated with reproductive longevity and characterized by a restriction fragment pattern, wherein said pattern when compared to a second restriction pattern is known to have or not have a desired polymorphic marker, the presence of said marker being indicative of an animal more likely to have reproductive longevity.

- 26. The method of claim 25 wherein the assaying step comprises amplifying the gene or a region thereof containing the marker with a forward and a reverse sequence primer.
- 27. The method of claim 26 wherein the forward primer is SEQ ID NO:8 and the reverse primer is SEQ ID NO:9.
 - 28. The method of claim 26 wherein the forward primer is SEQ ID NO:10 and the reverse primer is SEQ ID NO:11.
- 10 29. The method of claim 26 wherein the forward primer is SEQ ID NO:12 and said reverse primer is SEQ ID NO:13.
 - 30. The method of claim 26 wherein the forward primer is SEQ ID NO:14 and the reverse primer is SEQ ID NO:15.
 - 31. The method of claim 26 wherein the forward primer is SEQ ID NO:16 and the reverse primer is SEQ ID NO:17.
- 32. The method of claim 26 wherein the forward primer is SEQ ID NO:18 and the reverse primer is SEQ ID NO:19.
 - 33. The method of claim 25 wherein the marker is *DpnII*.

- 34. The method of claim 25 wherein the marker is *HpaII*.
- 35. The method of claim 25 wherein the marker is Taq1.
- 36. The method of claim 25 wherein the marker is MnII.
- 30 37. The method of claim 25 wherein the marker is AvaII.

- 38. The method of claim 33 wherein a G to A nucleotide substitution results in a restriction pattern characterized by a 328 nucleotide fragment, a 125 nucleotide fragment, and a 32 nucleotide fragment.
- 5 39. The method of claim 34 wherein an A to G nucleotide substitution results in a restriction pattern characterized by a 373 nucleotide fragment, a 134 nucleotide fragment, and a 127 nucleotide fragment.
- 40. The method of claim 34 wherein the deletion is characterized by a 12 bp fragment having SEQ ID NO:20 appearing once in the IGF-1R gene.
 - 41. The method of claim 35 wherein a G to A nucleotide substitution results in a restriction pattern characterized by a 135 nucleotide fragment and an 84 nucleotide fragment.
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 42. The method of claim 36 wherein an G to C nucleotide substitution results in a restriction pattern characterized by a 137 nucleotide fragment, a 104 nucleotide fragment, a 55 nucleotide fragment, and an 11 nucleotide fragment.
- 20 43. The method of claim 37 wherein an G to A nucleotide substitution results in a restriction pattern characterized by a 122 nucleotide fragment, an 81 nucleotide fragment, a 60 nucleotide fragment, and a 44 nucleotide fragment.
- 44. The method of claim 25 wherein said animal is selected from the group consisting of: a pig and a mouse.
 - 45. A method for screening animals to determine those more likely to exhibit favorable traits associated with reproductive longevity, said method comprising: obtaining a genetic sample from an animal; and
- detecting the presence or absence of at least one allele in the IGF-1R gene wherein the presence of the allele is predictive of the animal having reproductive longevity.

- 46. The method of claim 45 wherein the allele is defined in intron 16 of the gene.
- 47. The method of claim 45 wherein the allele is defined in exon 21 at position 3876 of the gene.
- 48. The method of claim 45 wherein the allele is defined in exon 21 at positions 3896-3907 of the gene.
- 49. The method of claim 45 wherein the allele is defined at position 27 at the end of intron 16 of the gene.
 - 50. The method of claim 45 wherein the allele is defined at position 73 at the end of intron 16 of the gene.
- 15 51. The method of claim 45 wherein the animal is selected from a group consisting of: a pig and a mouse.
 - 52. A method for determining the haplotype of the IGF-1R gene of an animal comprising:
- obtaining a genetic sample from an animal; and analyzing the genetic sample for the presence of an IGF-1R gene A_1D_1 , A_1D_2 , or A_2D_1 haplotype allele, wherein the haplotype effects reproductive performance or the ability to sustain stress factors.
- 25 53. The method of claim 52 wherein the A₁D₁ allele is indicative of having a favorable effect on lactation and pregnancy stress.
 - 54. The method of claim 52 wherein the A_1D_2 allele is indicative of having a negative effect on reproductive performance.

- 55. The method of claim 52 wherein the A_2D_1 allele is indicative of reproductive longevity.
- 56. The method of claim 52 wherein the animal is a mouse.

57. A method for genotyping an animal for reproductive longevity, the method comprising:

obtaining a sample of genetic material from an animal;

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detecting a polymorphism in the IGF-1R gene of the animal;

- determining whether the animal possesses a marker, wherein the marker is indicative of the animal having two copies of allele 2.
 - 58. The method of claim 57 wherein the step of detecting the polymorphism comprises: digesting amplified nucleic acid with a restriction enzyme; and
- separating the nucleic acid fragments according to size such that a restriction fragment pattern is generated,

wherein the restriction fragment pattern generated is indicative of an animal reproductive longevity.

- 20 59. The method of claim 57 wherein prior to digesting the nucleic acid with a restriction enzyme, amplifying the nucleic acid with a forward primer and a reverse primer.
 - 60. The method of claim 59 wherein the forward and reverse primer is SEQ ID NO:21 and SEQ ID NO:22.
 - 61. The method of claim 57 wherein the restriction enzyme is Fokl.
 - 62. The method of claim 58 wherein the restriction pattern characterized by a 295 nucleotide fragment, and a 55 nucleotide fragment.
 - 63. The method of claim 57 wherein the marker is positively associated with longevity.

- 64. The method of claim 57 wherein the animal is a pig.
- 65. A method for genetically identifying an animal comprising: obtaining a sample of genetic material from an animal; and

identity to SEQ ID NO:1 or a fragment thereof.

- assaying for the presence of a genotype in the IGF-1R gene sequence as set forth in SEQ ID NO:1 or a region thereof in the sample, wherein the animal possesses a nucleic acid sequence having at least 95% sequence
- 10 66. The method of claim 65 wherein the polymorphism is identified by a G to A nucleotide substitution in intron 16.
 - 67. The method of claim 65 wherein the polymorphism is identified by an A to G nucleotide substitution in exon 21.
 - 68. The method of claim 65 wherein the polymorphism is identified as a 12 bp deletion in exon 21.
- 69. The method of claim 65 wherein the polymorphism is identified as an insertion of a 20 G nucleotide in intron 16 at position 176.
 - 70. The method of claim 65 wherein the animal is a mouse.

identity to SEQ ID NO:7 or a fragment thereof.

71. A method for genetically identifying an animal comprising:
25 obtaining a sample of genetic material from an animal; and assaying for the presence of a genotype in the IGF-1R gene sequence as set forth in SEQ ID NO:7 or a region thereof in the sample,
wherein the animal posses a nucleic acid sequence having at least 95% sequence

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- 72. The method of claim 71 wherein said polymorphism is identified as a G to A nucleotide substitution in intron 16.
- 73. The method of claim 71 wherein said polymorphism is identified as a G to C nucleotide substitution in intron 16.
 - 74. The method of claim 71 wherein said polymorphism is identified as a G to A nucleotide substitution in exon 8.
- 10 75. The method of claim 71 wherein the animal is a pig.

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- 76. The method of claim 65 wherein the polymorphism is an A to G nucleotide substitution in exon 21 at position 3876.
- 15 77. The method of claim 65 wherein the polymorphism is a 12 bp deletion in exon 21 at positions 3896-3907.
 - 78. The method of claim 71 wherein said polymorphism is a G to A nucleotide substitution at position 27 from the end of intron 16.
 - 79. The method of claim 71 wherein said polymorphism is a G to C nucleotide substitution at position 73 from the end of intron 16.
- 80. A method for genetically identifying cattle with respect to its potential to
 25 reproductive longevity comprising:
 obtaining a sample of genetic material from a cow; and
 assaying for the presence of a polymorphism in the insulin-like growth factor 1 receptor
 gene (IGF-1R), wherein the polymorphism is associated with reproductive longevity.

ABSTRACT OF THE DISCLOSURE

Disclosed herein are embodiments for genotyping an animal for the presence of polymorphic alleles in the IGF-1R gene that are associated with reproductive longevity and/or ability to better sustain stress, and preferably selecting those animals for future breeding purposes.

Application Data Sh_t

Application Information

Number of copies of CRF::

Application Type:: Regular

Subject Matter:: Utility

Sequence submission?:: Yes

Computer Readable Form (CRF)?:: Yes

Title: INSULIN-LIKE GROWTH FACTOR-1

RECEPTOR (IGF-1R) POLYMORPHIC

ALLELES AND USE OF THE SAME TO

IDENTIFY DNA MARKERS FOR REPRODUCTIVE LONGEVITY

Attorney Docket Number:: P05562US00

Request for Early Publication:: No

Request for Non-Publication:: No

Total Drawing Sheets:: 83

Small Entity?:: No

Petition included?:: No

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State of mailing address:	lowa			
Country of mailing address::	USA			
Postal Zip Code or mailing				
address::	50309-2721			
Phone number::	. 515-288-3667			
Fax number::	515-288-1338			
E-Mail Address::	patatty@ipmvs.com			
Representative Information				
Representative Customer	22885			
Number::				

Assignment Information

Assignee name::

Street of mailing address::

City of mailing address::

State or Province of

mailing address::

Country of mailing address::

Postal or Zip Code of

mailing address::

Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 1 of 83

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Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R)
Polymorphic Alleles And Use Of The Same To Identify
DNA Markers For Reproductive Longevity
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 4 of 83

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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R)
Polymorphic Alleles And Use Of The Same To Identify
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 6 of 83

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Applicant: Farid, Abdol Hussain et al.
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Title: Insulin-Like Growth Factor-1 Receptor (IGF-IR) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 7 of 83

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ID	5:	421	caccctggtcatcatggaactaatgacacgcggtgatctcaaaagttatctccggtctct 480
ID	6:	56213	caccetggtcatcatggaactaatgacacgcggtgatctcaaaagttatctccggtctct 56154
ID	5:	481	gaggcc 486
ID	6:	56153	gaggcc 56148

FIGURE 5

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2581 caggactcaa tgcctaggga atgacactgc ccatagtgaa ctgggttccc ccacatcaat
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14761 gatetgaaga etgetttgae ateaggeacg gagatgeaga gtttggagtt tacacagetg.
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16861 caggtagett cecaaceatt tetecatett tgggtecatt gttgeetgte tgaatatett
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16981 gcacctggaa agtggagaaa catcctagag agacacagcc ttgcaaggct ggtccagggt
17041 gtgggcagtg tacagctgtg actgcagatg tcagcacaga tgctgggtac ctgattcatg
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19201 caggecagae etetacaaaa aaaateecaa caacaacaac aacaacacca ecaceaacaa
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19261 cacctttgca ggaaggtgtt ggtaaaggcc ctagtttaaa aatggtgtta catatacaca
19321 ccatgaagtt cagagagtcc attcgcaaca ttatttgacc ttcagcacga tttgtttctg
19381 gaactttccc accccattct tgcctcgttc ccccaggtcc tggtgactgt tattattttg
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21541 cttcctctct ttctttttta tttttattt ttctacttat tttcaatagc agccacctcc
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24001 cctttcttgt cccccctccc aacaacaaca acaaatccat ggcttttgtt tctaaaatta 24061 tatccgcaca ataacctctc tgccaattcc aggcacacag agccaatgga ccagctccct 24121 ttggataatt caaattgacc ccaagatagc cctaccccac ttgtacctcc tccctttgtt 24181 taagaggttg ctctaagctc tttgtttcct gaccgaaatc cttccatggc ttcctattat 24241 cccagagaag tccacactg actaagcatt ccacccagcg tctgcagcct tctcaagttt 24301 caatccccat ccttcctggg gtggagtgtc cctgttgtac ctttggtaga gatgactagg 24361 gactcatcag attacaccag gacagttgat gtttggatta gcagcagggg tgtgaggcta 24421 gatgcttatt tgccttgtgg catgcaggac acctgtcagc tgcggagaac ggctcgccat 24481 accaatttcc cgaagggcct cgggctcttg tgcttccttc tgctatgatg ctagcctcgg 24541 etgetgecae gtgtgtteee ageeteetge tetggeceat etteatetgt ceacagacet 24601 cagtgttgca aacatggctt cttcagatat gtctccctag ggtgcccccg cttcaggaag 24661 tacggctgag agagccatgg ctctttggtt cctggtcact ttgagttcct ccataccagg 24721 agggatgggt tggagctcag agaagaccct gggcatacag tcttgggtgc ctttgtgtcc 24781 aggcaactaa ggcacctcac ccgctatatt ttgcaggaac tttctaagct gggcttaggg 24841 gtggacactg acatagaact tcgaactctg cagctgcctg tggattacag ggaggtaaaa , 24901 cggaggctta ccacaatctg ggaagatttt caaccacaag taagtgatac ctggagaggc 24961 tgtgggtetg ggaacetgat gatgggeea gggeetagga aetttetgga eettgateta 25021 gatttgaagg ctggagtaca cctgaattaa agcttctgtc gtgagattct gggtctgtgt 25081 ataagtttat gctgaaacac gaggtagata gtaaaagatg actgggggca agtgggccgg 25141 acaaaatggg aatctggtat taaagccagg aggactctag gatagctcac acctatccta 25201 aaaaaagaga ggccctggat taagccctgt gtatggctaa ttctggctgt tctgaggaat 25261 ttaaggtaaa gegetttggg eetaggaagg tetgatgatg atteacceag ggttggteca 25321 ccttcaaaac atcctgccag gggcctggtg tgatggctca cacttttaat cctggcacct 25381 gtgagtttaa ggccagcctg gtctagttta tgagttcaca gccccattcc aaactaacaa 25441 aagtaactac atagtaactg aatataagca ctgtgtaaac aggggcacaa tgccaggtgt 25501 ggtageteae geetttaate ceageaettg ggaggeagag geaggtggat ttetgagtte 25561 aaggccagcc tggtctacag agtgagttcc aggatagcca ggactacaca gagaaaccct 25621 gtcttaaaaa acaaaaaaca aaaaaaccaa aaaaacaaaa caaaacaaaa aaaggacact 25681 gaccaaacac tgcagcaaaa tcctctgcaa ctgttttcaa aagaggacaa ataaagtcta 25741 ataagattcg attagactgt tgctagcttc ctttatgaaa gtttcattaa atacaggatc 25801 ctgttaaaag ccagcatctc ccagggtcat cttggatact tgctttcccc caactcctgg 25861 ggggaaagag agcacacaac ctcggatgag ccatttgaaa tacaaaagtc ctttttaatc 25921 cagaaaaaat ggggaaaagt ccttaaatgc agatgtagca tgcctaattc aacccaaaga 25981 tgtgttcctt ttgtatcagc accattactt aggggcctaa gcagggctcc tggaggactt 26041 ggagctagat ctgtttagcg gggctctgtc atcactggac ctctcttcac atctagtgat 26101 ctggtgtctg tcctgggatt tctgatatgt atgtccctac agcttgttgt gcatgtgggc 26161 atggactett etgecaagge catetttetg gaacagtgtg gtaagaaceg aggetategg 26221 gattcagatg taagaggett ccagceggag gatggagtgt geeteecegg tggeecegaa 26281 gtgaggctgt ctgttgtcaa catgaaggag gtctgccggc gtgtggctgt tgagaatgtc 26341 gaagtggcct tttcccgaga tgcgggcagg tacttcagga ctccgtgtgc caagtgccaa

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26401 acagatggga gagagggatt atcettcaga gtecettete tetectcaag aggetateet 26461 atgttgtcca tgtggctgct ttggtcacag gcatcctatt tgtcacaggc actgcagatt 26521 aatttaacct cttctccaaa cgctgacagc ccctcccct aagcaccttt cctctctct 26581 agggataaag aaatacccga ggggagttgg cagtaaggga gtcattgtgg tgaaggtgtc 26641 ttaccactag ttgtttgaaa ggccggctca attctgtcat gaccgcttgc tggtcattag 26701 actetgagee caagaaceaa tgtetgtetg gagtgttett aggeeatgte etttetetat 26761 cagaagagcc aaactcaagt ggtactaggc atgtgggcat ccataaactc atgcttcatg 26821 aaacctgtac ttactgtgtc tggcatcgac aagatctggg agtaatgggg agtttcatgt 26881 ggttgcaggt acatctgcga ttacacctac tacctgtctc tgcaccttgg gactgggcat 26941 gcggctctca tccatgtccc tcctctgtcg cactggctct cggccagcct tctgggcaaa 27001 gccttgcgag tcatcatcca agaaatgctg gaagaaatcg ggaaagtcca gactcaaagt 27061 acagcagctt aagggaagca gcagggggat gccttcctta gatgaaatgg agtgtttcag 27121 gcttggacgg agaagccagc tgtgctatga gaggctctaa aaaagggaaa cagttacaag 27181 gctgggggtg ggggtggggg tgggggtgtg gactcacaga atcacaatct tactttagtg 27241 totaaagcaa gaagaaaato oocaaaggog gatgottoat aaaggattaa otaggggaag 27301 ttatgacett tgtaacteca aagagecace aaaacetagt gtcaagacea aatgaactga 27421 atttattata caatgttctc cctgcatgta tgcctgcatg ccagcccagg gcaccagatc 27481 tgattataaa tggttatgaa ccaccatgtg gttgctggga atcgaactca ggacctctta 27541 agctctcagt catcttccct gctttgtttt gttttatttt gagacagggt ttctctgtgt 27601 agccccgcct ggctgtcctg gaactcacag aaatctgccg gattctgcaa gaaaatgact 27661 ttacaagatt teetttgeaa ataaaccaag gacattttac atatgacaga gecaagceac 27721 actaaggcag atcaaagccc aaggtggaag ccgggcgtgg tggcacacac ctttaatccc 27781 agcactcagg aggcagaggc aggcagattt ctgagttcca ggacagcctg gtctacaaag 27841 tgagtgccag gacagccagg gctacacaga gaaaccctgt ctcgaaaaaa acaaaaaaac 27901 aaacaaaaa agcccaaggt ggactgggat ggcacaggcc accagcacag aacctagctg 27961 cttggctggt gccttggctg gagacccctc tctgtgagca aagagctcct gctccaaacc 28021 ttaggtccct ttgttccagg agtgggctct agtgacactt caggtcactt ggcacctggt 28081 cagacacatt aaaagaaaac gccgggcata cgcctttaaa cccagcactt gggaggcaga 28141 ggcaggtgga tttctgagct caaggccagg gctacacaga gaaactctgt ttcaaaaaac 28201 agaggeccag accecaacce ceatgageag etettecace ecetetgege etetecagtt 28261 tetcaagace ctaagegeee atttgeattg tattttttgt aaatagettg geateeeeg 28321 acaatacaca ctgcatcctg ttcagtcaca gctgatagtg ggtggtttgt atgcttgcga 28381 gccagtgtag aacactgttt ttttgctggt catttaatgg ttgggaggtc tgaagcatgg 28441 ctctgtgttc atctataaac ccgactaata aaaagtcctg gttctgacta cccagagaat 28501 gtgtcaccaa caacacacca gccaaaaaaa ggaaagaaaa aaaagaaaaa ggtgtagtat 28621 ggtgaggtgg gtgggcagca gtatattatg aatctagcta gtaagcaggt gtttaaaaaa 28681 acgaataata aattaattet agaacgeteg taataggtga ettagetggg caggggtgge

FIGURE 6L

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40021 ggtattetea etggetetaa gtgaetggte etagtggeae atgaeaggtt geaggtaggt
40081 actaaaagct ctaggcccgg gagtcacgtc agtccccgag agccagagat gtctctttac
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40201 tgatcatttg tctgaagtca cacagggagt gagctggaac ggcgtgaggc tgtcagacaa
40261 tgcagettta etgtaagtaa eeatggtett getageagae ggegetetee egaetgteta
40321 cagacagtca gateteetga geeggettge tetggttage tgetgeagaa gettgttete
40381 cccggtcccc atcatttccc gaaggattaa tcggaatggg aagcagacac aaggagctta
40441 gctccgtggc caggctgcaa tttcctctct ggctgacagg cctagccctg aatagaaaag
```

Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-I Receptor (IGR-IR) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 25 of 83

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40501 tgcaaccggg ggcaagaaca tgagatgtgt ctggccccaa gccccgtggc caggggtggg
40561 gggtggggct gaggaggtgc cacgggcagg gtagggatga ggtgacatca gtaaaaacaa
40621 gccatgtgat gcctctttta aatcatccca gagtcagccc ttccagggct gcccataaaa
40681 ggaagtettt tttgetetgt gegatttaag acettgtttt ttttttgttt tgttttttt
40741 aaatactctt tccacaagaa aactctccag tagactctgg gattaataag ctgataaaat
40801 gctccctcta aaaagcacac tttaaaaacca ctggacagct ctgatgagct ggagccgcga
40861 tttacagaat atgtccctct gtgaaggtgc acacctatcc ctctagaagt gagtgctatc
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40981 ccccgtgtct tcagctggac aggctgccat ctatgtgagg cactggcaaa ctgaaatatg
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41101 gtgactetta gcagggatca gactttcaga agggcetacg ggactgagtg ggtcctcctc
41161 tagcatetea ageaactaag gttgttttag ggecaggace tagacgggga tgetatatag
41221 cccagttgct ggcacacagt cgatgccagg gaccgtccag cagacactgt cacctgcctg
41281 tgatgactet atteaattta tetegacaaa accagactgg gatteccage cteetggggg
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41581 gtgaactctg ggactggccc ctgacctcag cagggttcac tgacaaatct gtgtccaaag
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42061 tecetactgg ageageeegg etecetteee tteaetetgt tetttetta gaaaggagae
42121 cgatctgcca agggggttct gaatggcaaa gcaaaaaggc ctcctgaatc aatcatttcg
42181 catccaagag cggcctccgt gtgtaagctt cctcctctct atgctgggaa caggaagtcc
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42301 aaaaaaaaat ccaggcagct ggggatggat caacttcagg agttccatga gtttctcaga
42361 caaatgtttg ctccagcata aaagaaaaga aaaaaaccaa caatacctct cctgtgacca
42421 ggcccaaacc acaggaacaa aactcacacc tgctcttttg aacggtttct cagctcgtga
42481 gacagtgctg atcctggtat cctaaccaag gctggtgggg cacaccctta cacagactca
42541 gaggcagcca tgctcctact ccatcctttc acctttgccc acatactgac tattctcgca
42601 atggacgagt aggactgccc tgcaccactg aggtcaaagt aaagctataa cagcactcag
42661 aacccagage ttetetagge aggeeectg cateetgtac geetteggag gteagtttag
42721 aaaggccaca gtggcagtgt teteegtgaa gatgetgeag ggcacageag gecaeggace
42781 agtcagtggc tcaatatggc ctgctgtcta ttcagttttg taaggccctc ccaagcaaga
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42841 gggttttttt tttttacact tgccaacagc tggggcaaaa acaaacaaat taacaagtcc
42901 cccacgccc accccaaat aggagtattt tataagaaga aaacgagagg aattcaaatt
42961 ccaatataaa gttttagttg ggcaccagca gtaacactgg ctgcaatcca agcactaggg
43021 gagtttgagg caggaagatt atcctaaata cctgggctac actgactcca tccttgttga
43081 ttcctgaatc atagtggcag agcagagggg ctggaaggaa gagtgagcgt gcggtcccca
43141 gcaccaccaa tcctctccag tgtattagag aaatcctctc agtctatgga gtggtccaca
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43261 atccgatctc cttacaatca ccccacgtac cggagtatct tcatctgcgg gaagccctgc
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43561 catccaaggc gccacgagag tgagcgctgc gctggggaag agaaggccag gggtaggcag
43621 agtettttgt gtagaagtga agaaatettg acaaccccc tgggattett teactagtgg
43681 agctgaacct gttttgaagt gtttctggga aactgtatgg gtcctgacat atcagtggga
43741 acctgctttg atgcttaacc ccgagatgca gagcaacagc cgagccccac cttaacgtgt
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43921 cagcgggtaa gagcactgac tgctcttctg aaggtcctga gttcaaatcc cggcaaccac
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45181 taqtaactaa cttgcatttt tgtcttaaaa attaaatttt gcatgaaaaa gatgagccag
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45301 cacgccatgg tgcaaagtgc caattagcac tggggggacc aagggggcac aggatgagat
45361 taaggagcag cacaaagtca cagatccaac aagctcagca gcagcaggat agagcctttc
45421 taateetete etgtaceeae acacaaegga gtgggtgget etggaaatae atacageata
45481 tcagggcagt tgtccggctt gtccagaagg ccaccttca tgacgaaacg aagaacttgc
45541 tegttggaca agecetggta gggetgetea gecagegtgg egateteeca gaggaegace
45601 ccgaaggacc tacaggtgga acagaacagg aacagctcag aaagggatgg gaagggccat
45661 gtccctcct tgccagcatt tcttagactc tgggtcctga aaaaaagtgt cctcaatgtg
45721 acgactgcac tttgtggaca tcaaaccagc cagcttgcta cgtgtgggac atctgttccc
45781 ctttgtcact gctaaggctt cctatagggc tactgcaacc cagggctgag gtaattaatg
45841 ttaggggctt ctaacatctc tcacagtttt aagagcccga tctacagcgg acacattcag
45901 tgtctggtcc cagccattac ttggacataa gccaaattag ctctagttca gcaaggaatg
45961 cagaggacca ggtcccagcc tggccactga cctgagacag ctaagcatct caactatctc
46021 ctgcagggat ttctttcatg ccagcttaga aatactgatg ctaatggttc tccacgcctc
46081 cgcaagaaga atetgatcag ggttettaaa acaatgggta tettgggete tecaaagcaa
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47581 tcattctaaa atgactgcaa ttcctctagc aagcagaggt tcaggcagag ctgagcatcc
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FIGURE 6T

Applicant: Farid, Abdol Hussain et al.
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47641 gtttatacgt tctagtgtcc ttgatgtccc cagctgtgtg ctcaaacacc ccagcacttg
47701 gccttttctc ttcacgcatt ttcttattgg tttgtaatgg ctacgtaatc caagtggagg
47761 aaacctgcag gaagcaccct gtgcgcctct ggctgtggca tgcagttgtt acccagagtc
47821 gggggagaat gagtctgtgc tcactctttc cctatcctgt aaggtgatct ccaagcttcc
47881 agaaaaatga agcttagaac aaacagatgg aagtgtggcc ctaggtgtgc cctctcctgc
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49921 gggtgtgtaa tccctgagat gggggcactg ggggacactg aggaatgcag aactcgcccg
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Attorney Docket No. P05562US00
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50041 gaggatetge tttgacttge ataaaagggg aattetagaa aacaatetet etecetaaat
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50161 aagaaaaatc ataattataa tagaaaatat cctcataaat gcgccacatg gttaagcaga
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52381 gcccagcctt tacctggctt ctgacgacct gaactctggt ccttatgcct gtgaccaccc
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52441 cctccccct agtctttct atttggagca ggacaaaaat gatttgaagt gaatcagcaa
52501 gtttaggcat taagcattct cagctccagc tagtttccat gttcctttgg cttccgccat
52561 caccaagtgt gagtaagagg aaggacagag aacccgtaca aaggccacgt gaaacagaca
52621 cacttectaa gtggattace acgagetttg ettecegtge aagetgecag gaettatteg
52681 tetteattee ettettetaa ggtacaaaga taggaagaet aatgetttee tettggatta
52741 ctgttttcta cgggcactga ggataaccac agttcaggtg gagttcacct tggaggggac
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52921 aaacaaaata agataagatt tatgtctaag aggctgatga gaatggctta gtgggaaagg
52981 tacctgctgt caagacaggg ggcctatgtt catgtcccaa gacaggtatg gtaaaagatg
53041 aaaaccaact ccctcaaggt ctaagtaggt atgcacatgc atatgcatga acacacacac
53101 acacacaca acacacaca actacacaca tacaaacata aaaaattccc aaagctccgt
53161 caccccactc ccatgtgcaa aaccaaccat gacccttatg cctcagtttc cctaactgaa
53221 aaccttctca tacaacacta tgaagccatc tcatttccta atgtatctca agtcacagga
53281 gagcatgacc aggcttggag atcacactat ttgggctagt atataggccc tttgtgctct
53341 gaccacgatg aacactcaag agcattctgg ctcttggcca gggttgctaa gttctcagca
53401 gatgccagac totaggaggc cagcaacagc cototagcac gtocaggagt actotocaga
53461 cgacatggca tgtgaggacc aagttgagct catttatcta tttaaccatt ttcactctga
53521 tecagggtet etteetataa tetaggggat gteeteagag gatgtgeeat cacaggtget
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53641 ctctgcagtt ccgtgacatg tacttatagg ctcatgacac atactgcagt gtatgtgtgt
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53761 ggggacttaa ttatgtgcag gaaagggctg ggctgacatt ttgtcagttc atggtaatat
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53941 tagaatacgc tgaagccggg gtgtatacac tgaagctggg tatatatact ggagctaggg
54001 tatatacact gaactgtgga gtatacattg atgctaagat atatgtaagt tatttttaa
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54121 ttccgaaaga gactttcgta atccaacctg tctctttaca ttgaattaaa attttgtttt
54181 atgtttgtgg gtgttttgcc tgcatgtgtg tttgtgcact acctgcccac ggaggcccat
54241 ggaggccagc agaacagtga ataccgacag ttatgagccg ccacgtgggt gctgggaatc
54301 cacccagggt tetttggaag ageagecagt getettaact getgagecat etetatagee
54361 ctttctctct ctacatgagg aagcggaggc ccagaggcat ctgttaactg aggactggag
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54481 caaaggcatt aagcacattc tgatgttagg aggtgcctta aaggaccact ctccagtctc
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54601 ttgtggctct gctatcaatg catacaccgc cttgtggctc tctcctgaaa cttcagaaac
54661 acacatgagg acatttgtga acaggtactc agaactaaaa tgaaggaacc atccatcact
54721 tggtagatga ttacgttgtg gaggaaaaga aaacgaaagc ttctacagag aaggggagca
54781 aacccagaaa gggcccgaga agaggaagtt ccccttacag cgaatgcaga gacaagctgc
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Applicant: Farid, Abdol Hussain et al.
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54841 ttagttagcc acaggtatca aagccagggc cgaagtgaga gcctcagagc ctgaggcagc
54901 cccatgaaag gcatttggtc cagcaatatt tctcaacacc cccatcagtt aactcttagt
54961 gaggacttag aagccaaact taagccaggt gcttgtggtg cataccttta atgccagcac
55021 ttgggagaca gaggcaagcg aatctctgag agttcgaggc cagcctggtc tacgaagcga
55081 gttccaggac agccggggct gcacagaaaa aaaaacaaaa caaaatgctg tctcaaaaaa
55141 caacaaacaa acaaacacaa aaataacaac aacaaaaaag aagtcagact cacatatgct
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55261 attacacagg acctactcag gttcacagtt tgaaaatctc actagcctta agcagaatca
55321 tgcagctgga atggttatca tttaaaagcg caggcacaat actggcttat tgcctagcct
55381 ggggagtagg ttcagggttt gagaagtgtc tgaagcagtg tcggggcaag gggagagcag
55441 tgtgtgccat caaggtaagg atagcatgct cacatgtaca cgtgtgtgtg tgtgtgtgtg
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55561 tgtgtgtgtg tgtgtgtgt tgtaatgggc tcctgaggaa acagtggtct cgtgcacaga
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55681 tacttactga aaaggttacc agccacggtc atgaccctga ggtaaagaag aggctcaccc
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56581 cgattette acgeatactt geageetegt ttacegtett gatggeeact etggtttegg
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56701 cgageteceg gttcatggtg atettetete gagetacete ccatteatea ggeaegtaca
56761 ctagtaacag acaggatgga aacggccett agacagagat tacaagatge tgttccctaa
56821 gacccaccaa ggagaacatc aaggtacaaa cagtccagca cgggtactca atctctgagg
56881 actccatgac actttgaggg tggtactcgt agctaagaac cagaaggaaa ccaagaaacc
56941 taagatagga aaaccaaatc aaaccaaaca acagtggttc ctttttcata tccttggcaa
57001 aaccccaacc caacaaaata aaaaaagcaa attagcccta agagacaaag aaagtgggac
57061 ttctgttttg cctgtcttca tcaagggatg gaaatgtagg tcagaagggg agaacagtgg
57121 gaggggaaag gaccaaaggg ggtcaaatct cctcctccct ggatgaagcc agaggtgaga
57181 aacateetet geatggeetg ggtgeeagtt tacaggeeac ceageeceac tgtaggeett
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57241 cagcgcttga agtcgtgctg gaagtaagac atatactgca ttttaaagac ttgcttaaaa
57301 caagtaaaaa aagataactt ttaaaagatt ttaaaattta cgtatacatg tgaaaatctc
57421 tgagcacaca cacatgcaca gatgtgaggc accacccgtg tttgggtgcc tgggaaaccc
57481 agaagacagt'gttggcttct cggacctata gttacagatg gcgggaggtc tcttcacaga
57541 ggtgctggga actgaattag agttctccag aaggtcagca aggctctcaa ctgctgggcc
57601 cattttttt tccaqctttg tattccaaag acttggttaa agcaagttta aaaagatagc
57661 ttttatatta actgttggat gttgtgggct aaatgaaaat tctaacagat ttatctcatc
57721 agcttttaca cttaaaaaaa catggctact agctgggtgt ggcaacagga aactgcagtc
57781 ccaccccagc ccactgggca tggaggttgg agatggggtc tgctgaacta aaggcatctt
57841 cagattgtgt ttcaaaacac caagacaatg tgtcagccgc ctgggatgtc tatcagttca
57901 cgcacttcat tccacttctt ccatagccat gtccgacaac aagctcaagg tggctgagag
57961 ggagaaactc agtcaacact aaaggcagag actcaagaga agagtcctca gccatcaaaa
58021 gaaaccgcag aaaaggagat gagaggccag ccccaccc caccccgca gagcgctgag .
58081 gtaggtettg tgcactacaa gcactgtete cetatattee tecatacage tgtttaacte
58141 tggaagctgc agaggttgga tccagtttaa cacacctgc tgcccctttc accgagatct
58201 agtgacgata cagactgcct gccttttcca cctgagagga aggagaactc acacgttgat
58261 gaagaaaaa gcagggttgg gagatgaaac ccagggttca aatcccagat gattcaaggt
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58381 atttgactta gtggaatgca tactgtttta aaaccatcaa ttttgtgaat ggctgttttc
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58681 ggatggaggg ggactcagta gtgaccccac ggctttggcc tgcactctgc aatgccagga
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59461 tgctctattt ttgctgcctg atccagatgt agaactctta acccctctcc agcaccatag
59521 caccatgtct gcctacatac catcatgttt gctaccatta caacatgggc ctaaacccct
59581 gaacctgtaa gctcctttaa gccagccact ttccatgaga agagctgctg tggtcaacgg
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59641 tgtctcttca cagcagcaga acactaagac acatacttat ctgtctactc acttactctt
59701 acttactggc tgatttgggg acaggactgc tctatcttgt tcatacttgc cttgaacttg
59761 ctatgtaatc caagctagcc ctgaattgac aacttttctg cctcagtgtc cccagtacgt
59821 ggattccagg tttgtgtggc cacatttggc tcctgtgcat gtttttgtat ttcatggaat
59881 atgtatgtet ggtaatttet caactagaat tetgtettee atgggeetea gacaacttgt
59941 gaccetgeag aggetgaget tgetgaggag ageagagggg gaaaggtett tataacacet
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60061 taatgagaat caactacacg gcttgcttgc tgcagagcat ggagttaaaa gcccatcaca
60121 tccattccct gggaggtccc tgaagttagt ggtctcaacc ctttttctaa actcaatcca
60181 gctgccgact ttagccagag tacctcagct ttgccagcca gagctgttca taaacttcgt
60241 acaatgttat cagcactttc ctgtagggca ttctcagtgc actcttgaca acagcaacat
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60361 tagctetttg aggeaaaagg ettgtgetge tgetgtgett gteaaaaagge caaacaaace
60421 aagtgcagat gaggtgacag tgttctcacc tgttgacctg ggtagtgaca agtttgttct
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61261 cagetagete ttecatacae cacateatga ggatteagga ggaaggettg ggatgteeet
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61921 gaaggaaatg gtaccaaagg cagatcctgg gacaaaggta tctcaaaatt acgcaacgct
61981 tttctccagt cagcacctga cagctcccat attcccacac cttggccagg acgcttacca
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FIGURE 6X

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62041 tcagctgcgc tgaaatactc ggggttcaca gaagcataca gcactccatt gcccaacctg
62101 ctgttatttc tgtcaaaaaa aaaaagatga aaaccaggtc aacaaatccc atctctatac
62161 atttctgtaa ataccttttc ttttcctttt tttctttctc acgagttatc ttgagcaagg
62221 gacgcaaacc atgatcactg ctgacgggac ccttgcctgc actcctgatc caggccccgg
62281 tatctgagga caaatctttc cacttttgcc ctggcctcag attagagctt ctaaagaaac
62341 atttgtttct ttgcttcact ggaaataaca aggcccactc catgggaagc taaactggca
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62581 tececeacag acgagetatg tteaagttea aateateaag ecaatgegee acaagtactg
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64381 aataattaaa gtagtcatgg aactgagaga ggtgggcatg gaagaagctg ggagagggag
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64441 gagecagatg gtgtaaatac agtacteatg teagaagtte taaaaggeta aaactaaaac
64501 aatgaagaag aagcatcagt gaactcttcc aaggccagac tgaaaacgga ttcatgtact
64561 gggaagatga gtgtgtctga ggacacactg ggggcctctg cacactgcca ggtctagagg
64621 agttaaggga acgtaccagt ctcctgtggc atgctatgca tcgctccagt cagacctcag
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64981 agtgcctcca tgaaagcact ttagatttca ggcttataag gagtctatag caactattgg
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65401 ccctgccaga gagaagctcc gagctttcct tttcccctag gagaattcgt ggttttgtgt
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66181 gtgaaacccg ggccctttct gccttttgga ggtaaccagc ttggttccat catgtgcttc
66241 ctgcttctga cattttgtct cagaacagcc caaaagtaat agaatcaaca gacagagcac
66301 gacaaccccc gaaactgtgg gggccacccc tctaatcatc tctgggtttt gccacatcgg
66361 agagetacag taacatcaca atggagagtt agcacataag etaactgatg acgetagget
66421 acccagaaaa aaatgagaaa attcagagtc tacctgacag aactctcagt ctcatcatct
66481 ctcaatctag acacagtagc agcagataca gggaatccta gcttcctgct tacggttcta
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66661 acatatttac cacatacaac atatcttaaa gcatgtgtac attaacccaa tcagcatcag
66721 cattaagtcg cattgttacc atttctgtgg tggtaacact ttcttagcaa tattcaataa
66781 tgetttatta accatageta gtgtgttata aageteaete ttetatetae etaaaattgt
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity
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66841 gtaccttttg agcaacaccc cctaagttat gttcccctgg taacctgcaa gactctgagt
66901 tcaccctttt agatttattt catatatacg tgagatcacg tggtgtttcc ctttctgcac
66961 cttctattaa cagtattcca ttacatatat ggaccacatt tttacccagg cacctgctgg
67021 aggatacttc attgtttggc tactgtgact agtgggccaa aaaaacaagg ggggtgtaga
67081 tttcaacaca actaacttcc atttcttttg gactgggatt gctggatctg ttgagacctt
67141 gtctgaatca agaatccaga agagaccacg aagaacataa gacattagga aacaacagtt
67201 ccccacacc tagattctag cctcaaagct gagctgagat tttggggggt ggggagtgca
67261 aaaggaattc acatttgatc ctgtatttgt ttggatatta catgaacacg aaataatcta
67321 aaaaaatttg gcttctcata agacacgtaa aaattcacta tgtattattt caccaaaaag
67381 actcaatttt ttttttttt cggtcaaaca agcatgcagc ccttgcattt aacatttcac
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69061 caagcagtca aattgggttt aggtccccag aaagaatgat aaacggtgtg ctaacttgag
69121 aacaactgtt cccccaatcc ttgtatatac aggactctgg acaggggacc ttgtaggagt
69181 tettteetga gatggetget ggcacagege etceatecee tetgaetgaa aaaacetgtg
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity
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69241 gccaagtaca agtgatcacg cctgaaaaac ctatgtaaca ttctccaggt ccattttcct
69301 acctaaaaag ttaaaaaaaa aaaaaaagaa cccttcaaac agagcggccc tggggaaggg
69361 cagtgagega cagegtgetg agectecage tgtggcaetg acacatteae ageceacace
69421 ctcacacggg agggaggaag ggccagttcc gaggctggcc tctgaggcag ctgttccggc
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69781 gcccacaggc tgatctagct gaccacgagg acaacgcttc ctctgcatca agctttgaga
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71581 agagtacggt gctcgaggcg ccttggaagc catgcccaag gcctatggat gctccctagg
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R)
Polymorphic Alleles And Use Of The Same To Identify
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71641 ccaacatctg ggagagtaca gtagggagtt gggggaaagg gccgcatgct ccctggtctt
71701 cagcagatgg atctgctacc aaaaccatag ttctttgtcg ggtactgaca tgccagtttc
71761 cattgaaagt atatcaactg ccataccagg gcccatctca aaagttagga atggggattt
71821 gttatcataa tgctggggaa agagatagga aatgagggta ctattctacc atgctcacac
71881 ccagcaaacc ataaaccgac caatcaacaa accatggaag gaaagataaa ggaaactact
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72241 tattcaacag ttactgtgga cacagagtca ggaccatgaa gggaggcaat ataggcatca
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72481 tgaaagccat ttactgatac caagtttagg aagtaacttt ataaatagta gcctccactc
72541 tactataaac tcacaggcaa agtcagacaa tgcagaaaga ctaaatctga tatatttgcc
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73921 teetteteag cetgettete agetteagtt ttagggeaag egeageatgg ecetttatea
73981 ccaccacaca cttctgtctt gggattttcc gtcacctcct ccacgtcgat ggtaccatcg
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity
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74041 gcgtactttc tgatgggtat tttgtctgct tggggaagac cagagatggg aatgaaagcc
74161 agaagcaagc cctgcaacgt gacaggtgtg accctggggc tcaggttctc tcagacaaca
74221 gacaaggcca gtggcactgc ctgcctgctg cctgccagcc cccagctcac ctttggagca
74281 gtagttgtgc cggtacaggt aaccatectg gggetgeege tgecacetea caatgtagta
74341 actcaagtta ccattgggca gagttggagg attccacttc acaatcagct gagaggaaga
74401 qtttgatgct gagaggacat ctaggggaat ggaagggact ggaaagaaga acaaagaaca
74461 tttcagagaa tagaccggaa tccgaggttg aggttatcag gggctcgatg caaaccaaca
74521 ggaagctgca ggcttgacct gggcacagag atagcaacgg accaaccaga gcatgaggtg
74581 aaccagaget ttggaaagat taccaactgg gaggecagee eteteteaga geaettgete
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76321 gaagggatta gtcaataatc cgatatcata gtactctaag cctgcgttcg cctgttgaga
76381 aagctggtgg ctacggaata aaggtggaaa gccagcaagc cactagaggc aaggggtgca
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FIGURE 6EE

Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 40 of 83

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76501 gtggctacat cctgggcatt cagagaaacc atcctactgc tctggaggac accgattcaa
76561 ttgcgtagag cagagatgtt gacatttaag atggtaccaa gtctaggaaa tggctcagga
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76921 gctcgccctc cttgttcgga ggcaggtcta catccaccat gttccagctg ttggagccac
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78781 gaacagagca ccccatgggg tattttgggg gacagaggct aagacaatat ttaaccaagt
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FIGURE 6FF

Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R)
Polymorphic Alleles And Use Of The Same To Identify
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78901 cccttggggt gaacttactc aaacaccata taaacaatga aagggctgag gcgggacact
78961 ctcctgacta gctgctgtct caggttggga cagtttaata tgacaactag agaggtcaca
79021 tgaggatgag gtgacggtcg cctgacttga ttctttatag gtacaccaaa gactctctat
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79141 ggcccaaact cacgcctcct tgtagtaaac tgtgaagctg atgagatccc ggtagtccgg
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80881 cctctattac caacacacaa agcttctccc accaaaggat gttccttgat tcattccctg
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81181 ccetgagtac gtgggagagg acagacatcc agtgtctccc tgggctctca catcagggta
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity
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81241 gctagatgca gcgtgtaaca tggtggcttt ccacatttag tatgtacaca gcttgttctc
81301 agtateceet acceptagag ettetggtge agaggtecag agtgeaceag gaatetegat
81361 gtctagcagg taatgctgat aaacaccagt cctgggtcca cccagaaact accaggccgg
81421 ccacggcact gtttccaagg cctaacttcc actagagggg agatatgggc atggccaaag
81481 ggaagatage tatettaggg ggecaagggg agageaagga ttetaettgg gtgtetgeat
81541 aagtttttt tttttttta aacagttatt tctaaatcca cccacagacc agacaaacat
81601 gtcccaaagt cctgttctgc cctaaaggct gtgtccacct aggcaccacc tgctgagagt
81661 cctcacccac attagaacta ctgcctaggg ggtctgcttt caacatgacc ttcacatact
81721 gagaaatgag ttacagctgc agggttgcca agcctctccc gacacttcct ggagaactca
81781 ttagggaget cagaggecag gaeceaggae eggaagagee atgaatteee cacaggaeaa
81841 gccacaggca gtttcctcgg tgctatgaag agcagcgttc atgtgcgtag gtgaagtctg
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83581 ggccctgtca catgataagc tcacagccaa cacttagcta tgatctgtgg ctaccatttc
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FIGURE 6HH

Applicant: Farid, Abdol Hussain et al.
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Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 43 of 83

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83761 teagettete tagggetget ggtteteage tgeceatgga getatetttt etggtetett
83821 cgggccccga ggccaattga gaagaactgg atgctaaata taaaaggacg ttataagctt
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83941 ccagaacaaa gtcacagaaa ataggtaagt cctgcacctt gatacaggca gggctgagct
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84181 cactacttag cccatctgta gaatattggt tcacttcagg tccacagtta taccatctac
84241 aaaggeteag ttagtatgga aatatataee caggeaaaae taaaccaage accacataga
84301 ttgacaaaat aggccgtttg ttcctgagtg ctttttaaaa acagctttgc tgaggtacgt
84361 tttaccttct aaaattgttc ccactataaa tatacagctc catgggcttc aataaattta
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85921 tcagtagcca tacaaaaagc tcaagaaata gattccttca acactgagtt gaggcaagat
85981 tgcttaaaaa taaaataaaa tataaaataa aataaaaact cagaagtggc ctgggactat
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Applicant: Farld, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R)
Polymorphic Alleles And Use Of The Same To Identify
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86041 atacatttga aggatcacag ggatagcgaa gtcatttcta atctttgcca tgtgccaaac
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86161 tttctgtctg acttggggtg aggtgaacct ggtgaagttc cacagggctc ccctttccct
86221 gtaactgtga tgtatccaca cattctggga aacactgggg ggagggggag gaggctatca
86281 gtataaacga cagcaccctg aaattaaatg tcagataact ctgctaatac acccctcctc
86341 aaaatctcca gtgcgtccca aggcttacct aaatgataac ctctacatct gcatgcaaca
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86461 aaatgtcatt ctgtgctctc tectettetg tecetgtget ttetagtece ccaaaccett
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86941 gtaaaattac agatatgacg tagcaatgaa tagattatgg ttgggggtca ccactacatg
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88321 aaacaaataa aatactcatt taaaaaaaaa aaagacttgg ctggtacttc atacctaagt
88381 tttgaataat gaaaaaatag gctataaata ttcttggaaa taaaaaccag ccttcaattt
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 45 of 83

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88441 tcagcaacat ctaagtgtat gatactttta gttaaggcat aatgataaga taatcatacc
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88681 actgggggga cataatgaaa cgagatggag ttacagttca ctttaaaatt taatacaagc
88741 aacaatgaat taagggtgac cacggcctca tgtacgggag gccccagaat gattgcgaca
88801 ctctgggcag ggtacctggc acatgcagaa ggattcgcaa ggagaaggag gtttttctga
88861 agcttctctg ctcgcaatgg gaagatggac ggggtgagca gctctgtttc tggctgtatg
88921 taccagggga agacccaggc cagagttggt gtgagaaaaa ctggaggttg taaagaaacc
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90661 tgagacgaag gttcttcagg aaggacaagg agaccaaggc atgagaatgg cggatcttca
90721 cgtagccggt caccacctcg atgagcccca tgaagttctc caactccgag gcaatgttat
90781 ctagagaaag aggagtccat gtcaccgagt cttcctctgg tcaggtggcg cactatactc
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FIGURE 6KK

Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity
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90841 agattcacac acactggtgt ggatgcgaag gggaagcagg tcttgcgtgg gggaattgca
90901 gtgagaaggc tggcatagca ccaaccgtcc ctgtctccac tctccgcaag ccacaccggg
90961 acaaggcctc tgaggcggga aagcagactt ctaggaggct ggctgctgca gttgccaagc
91021 tgattaacag aacgtgaaat cagaaatgcc aggctgctaa ttagccccag ctgacaaatg
91081 gaggcaacat ctcttagata tgctcagtga caggcagtcc tggctccaca gtgccatggc
91141 aggactgggc aaacctgtcc cagcctggta agaatggtct ttgctggacc tggagttcca
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91261 gcaggtagtt gatctatctt tccccacct ggatcgagac agggtttcat gtatgctaga
91321 cttaagatga ggctgaactc ctgatcttcc tgcttcctct ccctgatccc tattcctggc
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91501 accagaaagt tgccaggaaa agttgctaat gtaacaggac cgagagccag tgaatttctt
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92161 tgtagaccag gctagcctca agttcagaga tctgcctgcc tctgcctcag aagcactagg
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92281 ctgcctcaga agcactagga tgaaagttgt ctgtcaccac acccagtaag taattcttct
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92881 aaaagaaacc ctaaaccaaa aaaaaaaaaa aaaaaaacaa caaaaaaacc caccttgttt
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93001 aatccactat aaataaagag gacactttgt ctctgaagta aagaaaaact ttctttccct
93061 aaagatttac cagtaaagat taaaataata gtcttctgcc tatagtattt tacctcatga
93121 aactactagt agctcaacct agaagttcta gtggcaggaa ctgaagggca tctatctagg
93181 ggactttctc tgtcagtcca agactagaga agataagatt acagggctct gctgccacca
```

Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R)
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93241 tgggagatgt tgttcatgaa tacaattctg aagcaacatc ccaggagcgc tgtctacacc
93301 aagtgaagaa agggagtctg caggctcaga ccacagcttg aggtgtggat ggcacatctt
93361 tctggctctg agaggtaaaa caggagatca ctctttaatc tttcctttgc tcagtctcct
93421 caacggaggc aagcaaccca aactctggcc ttacagaagt gtggactctg cagcagctag
93481 catgagecag ccatgggtct gagecateag aggtgtgtgt gtgtgtgtgt gagagagagt
93661 qaqaqaqaga gaqaacaggc ctttcctgcg taagatgtga cttaaagaaa gacgagggct
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93781 tqqattttat gtttccqaqa aactcacagg ggaggcgtca cacaagaggc ttacttacat
93841 cccgtgctgc ttttctaagc ctgtattctg cttgtgacta aagggacaga gccaggaaga
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94081 ggggcagggg cottogcagg ggatacagta catgotggtg aagataaaga gtggaccaaa
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94801 ccctcagtgc cacacacac ctggagccag aagcagtggc atgcttcctg cttcctgccc
94861 atcatttagg aaacggcaga gaagatgcta agcctggggc tataccttct gagaccttag
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95101 catecettee caaaatacta tetatagtet gttttgttta aaaacettaa tagttetetg
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95221 gtggacctag ctagccaaag tgggggacat ctggcttcct ataataccac atgcaacatg
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95341 gctaacagaa ctgcttggga aggattagga ggcgtctggt gctgagctct cagttatttc
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95581 tttctcattt aactacatga cgcataaaac ccagctctct ctaaaaacaa acaaacaaac
```

Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 48 of 83

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95641 aaaaacatac ttggaataac tgcagcttca tacacagacc cgtgcagaca gtacagagca
95701 cccgtgtact ttccacccaa cgctcccaac tgctgctaca tccaacataa cgataacaca
95761 gtgacctggc acgegtgtca ggttctgtgc atttctccat ctgtctacat agacctgtaa
95821 attocctact gcaatcgagt togaggacaa cttcaacaaa gtcctcctgt gtgttgcctg
95881 tttgtagetg ccetetgace tgececagae tteactatee etaacteceg gtaactgeca
95941 aggtgtcttc tactgccacc ttgggaaatg ttacagaaag gcagtcatat agcacatggc
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96061 ctcctggtga tgttctgtgg ggaggcctgc tttcctactt attgtaccag atatctgagt
96121 tgtcctcact tcctggtatt acaaacaaag aggctacaga caagtacatg aaataaacgt
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96241 cctggagcca cctcccgctt tgttaaaatg tctaggaatc atttagcaat tgaaaacatg
96301 tetataaggg tgtgttetga ttatagetta agttteetaa aacecaetet tgtatagaca
96361 gagcacaaac agtattttca aaatactttc aagcaagtgc taaataggct tcaaaggcca
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96541 tectatgaag acacagcate etcactgtgt gttacaggac agttgaatge ettteagtag
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. 97381 ctcctcctcc tcctcttcct cttcttcata aactgatcac cctaagtatt ttgttacagt
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97861 ggacetteta caaggtgggg actggateca gggcatetea caagtgtggg cagtatteag
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGR-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 49 of 83

```
98041 caacaacctc agacctctga ctgtcctgtg tttccagagt tataaaatcc tgtcttcacg
98101 aacatcaccc atgacgagtt tgtggaagta cagcttgttc agtcagaaac agctctgctt
98161 ccacagtgaa gcctcagagt ccaggaagga aaaccagagg ccacaaagat ggtggaaggc
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98281 gcccaggett gcaatgagac atgggagace tgtttccagg ceteatgaat getggtgagg
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98401 tgggaaggag tatctgtagt taaagtctcc accgctcatg cttcagcttt gcaggtgcac
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98521 aggcacctc aaagtttaga aataaaaaag caagctgaaa cagacccacc actccaagat
98581 tegeattaaa acaageteag ceateaceet etgacagace catetgggtg aacceetggg
98641 aggteetaca ettagggget atgeageaga gaacceteet gggeetaggg acaagageeg
98701 ctgacctctg ggtgctgttg cggatgaagc ctgagggaca ctcctgcatg cactcatcgt
98761 cgtggataac gaagccatcc gagtcactgc tctcagcgtt ggggatgttg gcgcagaaat
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100381 taacagtaca tgtacccaat tgggggaagg gattgtttgg aaatactaca gaactacata
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R)
Polymorphic Alleles And Use Of The Same To Identify
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100441 aatatgtata ttttaactgt gatgtaccaa agagtcaaga ttccatgttt tattaaaaaa
 100501 aaaaaaact actctaaaac caaaactaag ccaaataagc cccagtaacc acaggttaac
 100561 ataaagaaca cacaagttca tctgcaccca aaacagccgc tttgtcaacc cttcatgacg
100621 tggtetttee egggttette etgtgeacag gtgtatgaet acacatacet gaatgttaaa
 100681 ccttggacta cggagaaagc actggataga cttggagtgg ataccaccat acagcaatct
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/ 101341 agatggaaca tgaggcaaca tagtgagtte aacttggeet etcaagatgg etggtgtete
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Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity
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102841 gattatcaat gaggactaga tgccaacttt ggagcccttt acttacttac ataggaaaga
102901 aaatgataca tcacaataga tgatgaagtc aaatagctac atagtagtca aagcccatac
102961 catataaaag cagggggaaa taaccttgta cactgtcttg aatcagttac cccccaaatg
103021 caaatgctac atctttccag gatgagaaag agaaatagaa atccttgaca aaggatttgc
103081 taaatttgag acatgttgag ccctgttcca ggcctgtgat gggaacttct tctcaactaa
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103261 tctgtgggtg gaagaggaca aaaaaggggt atgcagagag ggagagagaa gatggaggaa
103321 ggccgacctg cttgctaggg gactgactct agagatctgt gtgtggctac agggaggaaa
103381 ggggtgtggc teteteteca geacagagga ttecaggagg aageaggaga aegeaggget
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FIGURE 6QQ

Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-IR) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-I Receptor (IGF-1R)
Polymorphic Alleles And Use Of The Same To Identify
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-I Receptor (IGF-IR) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 54 of 83

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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-IR) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 55 of 83

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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R)
Polymorphic Alleles And Use Of The Same To Identify
DNA Markers For Reproductive Longevity
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 57 of 83

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118021	gctaaaacag	agaaaacacg	taaataacat	gtctgccagg	tagcaagcta	ttccaagaca
					cacaaagaga	
118141	gtggctcagt	tagtgtagtg	cttatctcta	tccgagaagt	cctgggttca	attcccagca
					gtccctggag	
118261	ggggactcca	agttcaaagt	tactcctggc	tatatagtga	agaatgcaag	gcagactaga
					tccttccaat	
118381	actcaacttt	gcttcagctt	acattataaa	cgctggtgtt	agcatagata	cagaagctct
118441	ttcctgtgtg	gtgatgaatc	attactcaca	accaaaccaa	gtgaggaagc	agggcccctt
					tgtataatgg	
118561	aactgggttg	agtaccccaa	atactcaaaa	gaaacagaca	atgctaacct	gggagattag
118621	aaatgattct	gcaactagtt	ccctgctgca	tgtgcagctg	gaggccccca	caagttggag
118681	gcttggtggg	agactgagaa	agcatttgct	attcgttttc	taggtccaag	gcatttcttc
					taagaagcac	
118801	cacacactcc	tgatcatcaa	gatggggagg	gtatccagct	tgttggctgg	tcactagaac
					tcactgatct	
					atgcttgctg	
118981	tcagggcagt	attccaggaa	tgcacagaca	gcctgctcag	cttctctcac	ttcccctctc
119041	atagcccaga	tcgaggaaga	tggattcacc	catcctgtca	acaaagactg	ccagaatctc
119101	ctgtagttgc	aggtatacaa	ttgtgagttg	ctatttaggt	tatggaaggg	ctgtggcatg
					aggggcaaat	
					cacacacaca	
119281	catgcacaaa	ggctcacaca	cacacataca	cactcacaca	cataaacaca	aacacacacg
119341	tgctcacata	tacacacaca	tgcatgtaca	cacactcatg	catacacaaa	cacacataaa
119401	cacacacata	tataccctac	ttctgtacat	gtgtagtcct	gactcaggac	cttgggacct
119461	gctgctcatt	cagcatatgc	cgctttctgt	tctccacaga	catcctgcag	atgtcccagt
119521	gacctgcctg	ccgcctgcct	tccctgtctt	tgaggtggtg	acaactcctg	ccaccccaac
119581	attttatgtc	acagetteet	cttcagctga	cttgaaagct	ttattttgtt	caaggtctac

FIGURE 6WW

Applicant: Farld, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity
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119641 cgagcaacca agaaagaaga gctagtgttt cacgggtaca gggtggagtg ttctggggat
119701 aggtcagtca tcgtttccct tttagggaga cttggatcgg tgttaatgaa ctactcacag
119761 gaactggcca gaatcagagc ccttgaattt tatgcactgg tccagggagg tcctgagaga
119821 gtggatgcag agtgcactgc cttttttctg cctttgcttg gaatcccaga ggagccagat
119881 tccaggaagg aagtggggca gggaggcagc agggtataac gaacaaacag aaaaacaaaa
119941 caacaacaac aacaacaaca accacctca aacctggtct ggctcagcgg gtaagagcac
120001 tgactgcttt tctgaaagcc ccaagttcaa atcccagcaa ccacatggtg gcttacaacc
120061 acccataatg agateegatg ecetettetg gtgettetga agteagetae agtgtaetea
120121 gtataataat aaataagtct ctgggccaga gcaagcagag ttcctaaaat tcaattccca
120181 acaaccacat gaaggttcac aaccatcagt aaatccatcc atccatccat ccatccatcc
120301 aggcatggcc acagaaacaa tgtgtgccta ttgctattag taacaaggta ccttcttcca
120361 tttgggctgt gtgcggcttc atggtagttt accctgtgct gcctgtgttc atttccacag
120421 ggaccccagg atggcaggaa gacctgcagc cttctcacct tcagaaccag agccaagaac
120481 acagagcaaa tgtcatgaat ggtcagagtc acacagcctg caggaggcaa agctggaatt
120541 ctaagtggct gagccttgag gtctcaaagt ttgtgaccaa tacactgtgg tggacctgcc
120601 ccggttccag agtcactgga aatattctca tagcaagcag gaagatctca gggacagcag
120661 acgtagttct aaatgggcat gtcttgctag aaggcacagc tgatgtgcct tttccagtaa
120721 gcattacaag aagcccattt ccatggttga aaaatgcaaa ggattattct gagccatccc
120781 gatgataaag aggacaacaa tttgccttta aaaaaaaaat ggaagctggg cacagtagca
120841 ctcttctgta gtctcagaac ttaagatggc tgaggaagag gaggacttgg gaagaggagt
120901 tcaaaggctg gcaggggaaa acacatcaag accttgtttc agatccacaa cgtaagcaaa
120961 ccagaaggtg gtggcaaacc tggcggcgac gcttaggaca tctaaatact caattatttc
121021 tacttcaaac agtgtaattc gtacaaactg gtagagcaca gaagcagaag aacattaaac
121081 acaggattta ataatggata tgtgcacaca catatacact gttgaagagc ttgtcgtctg
121141 aaagtttcta tgaattaaat atttgttcat taactactcg agttcacttg caaggcatgc
121201 caggcgccaa gggagagcac agaagtgcac aaggccttcc acacaaagac caggaaagga
121261 gatgtgcaca tagataatga gtaaggcaca gacttatgtc ataaagatgc gtatcaggaa
121321 gccaagagca ctgccagcca agaccatggc gtggaaaggc atgtgtaaag caggtgctcc
121381 cggcagggtc catgtatgca tgagaatctg ctggagtgat ggtgggggtg ggccccttag
121441 atccatgcat ttcaagggaa aactatcaat tatcatcttt atttatttat tttatgcatg
121501 tgagtacact gtcactcttc agacacacta gaagggggca tcagattccc actacaaatg
121561 gctgtgagcc accatgtggt tgctgggaat tgaactcagg acctctggaa gagcagtcag
121621 tgcttttaac cactgagcca tctctccagc ctaattatca tctttactga taactaaaat
121681 gccttaaaag tttaacttag taatactctg tttaaaaggt atataactaa acacagccaa
121741 tctgtttaaa acaacacagt ttcctgagtt gactagaaga tgctagtttt tcattctgtt
121801 tetectggta caagaagatt teaaaetate tacaateeet catttaaatt taaaaaecaa
121861 ccaaccaacc aaccaaccaa cctccattgt gagaggtgcc atgccactgg tacctgcctc
121981 tgcctaacag tgtggaaggc tgcacaggca gtggagggaa aaaaccccca aacctctgtc
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FIGURE 6XX

Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R)
Polymorphic Alleles And Use Of The Same To Identify
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122041 gtttgttaaa acaaacgage attttcacat tcactcatac tccctgaccc catagatcct
122101 gtgaattota aaaaggacca tgcttgatga gaatgttttt taaacattaa cgcctacaag
122161 qtqqaqaqcc aqaaqaaqqq aqccaaqaqt ggatggcgac cctcttcctc tcatgcttag
122221 caatttctgg tttcactcct atgcttctga gcaccccaac aggaaggctt cacagctcct
122281 gaaggtetga getagaceag ggtacacatg aatatgetga ttatgtggtg etcatgeaag
122341 gagaccccta aaagatgact teeceaccta agteegaace gecatgacte tgagataatt
122401 tacccaaagt tacatttctg tttcccaaag tattaacaag agtgtttggc tcttttaaaa
122461 agaagettag tettteaggg etttttttt tteeetttte ttttteatt tteaacaaat
122521 tgcttctatc gtctgagatc tccattacgg aactgtttgt attgagtttt agtgaaggaa
122581 gaaaaaaaaa aaaaaaagaa aacagagcgg ctgggaagat tcaagtgtgc tggagactta
122641 acattaggat cgggcacacc ccaggcttca gggttaagtt aaggagggtg cttgtgtcca
122701 gtgtgagagg tggggatggg gatagaggaa gcttcctcgg aagacttgac cactcccct
122761 cttttgtgaa gcctggaccc ctgtcacagc gagtaccaga gtacagcact cacccatgag
122821 cctcaattcc atgtgagect attetttact ccaegttagt etgeetettt tgcaageaca
122881 ctctgcaact tttcatttta aactgtgtgg tgtgctaacg agagtcagca gctctctgag
122941 tgacctaggc aagttettte tteeatgage tgeeteeage aacagagaaa gggggcaata
123001 ctgttgcagg gtcacggcag ttgggaacac ttttcctata gttccagaat tactggtctc
123061 ctacccaaac ctcagatcat cattaagaca tgctaatgtg tagcgagaca ggttggaagg
123121 atcttttct atataaaata ctgctgactg gggcgtggaa cccctgtcct attgggtgct
123181 gtggccatgc aaaaagaggg tcttccaaaa atttaggatt aagagggcat ggtgggagag
123241 gaacaacaaa ctttggaagc cttctcaatg tactgtagaa ttaaaagcca accaaggcaa
123301 ggaaggtgcc ttccaaccta gaaatccagc caccctctgc aacgagccca ggcctgctgg
123361 agaccacgtg aggtggtagt gggtatgcct ggaacctaac ataccacttc tagtaccgcc
123421 cagctggcac ccaatcagga gcggcacagc attctctaag cccaggttgg cacctgcaat
123481 catcttaagc atacaatggc attgatttcc acttagggtg tagaaataac actcagaaat
123541 tttacctatc tctgctgttt tgctgccaaa tttcaaatac tctccaagaa acgcaaacac
123601 aacagaacac ggaggatgtg acacacaaat tetgcacaat etcactgeca atacetegat
123661 gttatactca gtgtattaaa tggaatcaga atcctgagtc ccactgtcaa acttcctcct
123721 cgtctagtcg gtatgtgtga ctgacagctg atgtcactga atgcttgatg tctgaaatgc
123781 ctaggattcc tttttctccc cccatccaaa tataacagct ccatttcata ctccctagaa
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123901 accgtcctta cagatttctg ggagggagct caagagtaag taagtgtgta agccttcata
123961 gctctaccaa acacatgcat aaaacctgat taaacacaga ggaagaacat aaatattctc
124021 agtettetaa ggggeagatg aagtetgggt aagaaggaet aaagaaaaag catagagtaa
124081 aattegetee egettittaa taaetettta agataaggag agtigtitaa tgttiteaaa
124141 ataggcagag agagagagag cgagctagag agagaaagta tgtggtatag tttatgtgtg
124261 tgtgtgtgtg tgtgtagcac tgtgacagag tgtgtgcttg gcatgtgaaa tcccctggtt
124321 tcaatccttc cccttcagaa tgtttccttt ggaacaaacc aatacatctg gtgacgctga
124381 cctttagtgc ccctcctca ctcaccctgt tttctacagc aaaagtatct acaaaaagga
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-IR) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 60 of 83

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124441 agccgtgcct caggaagtct tcaccaaagc ttgaagagca tggagagaag aagtaaggtt
124501 ccatcagatg ccttgtgttg aggtaagtcc tccctcccaa tcaacccaca atgaatattc
124561 tetgatgtea geceetetet etgageatee tgtetetgag acettggaag taagaettet
124621 gtgtctggga ccaaaactac tggggctatt ttggcagctg agccctctgc ctcctctgca
124681 gggaaaaaca gtaggaacat ttcaaataac attcatattt gagatatttt ctggctctgc
124741 gtatttttct acttcatagc acgtgtaatt gtcctccctg ataaaagtgg acctagcact
124801 gctattttaa aagttttgtg ttctacactt taaaaggata cacaaagtcc tagcaggaga
124861 acqtcaatta ccaagtacaa aaacaaaaca aaacaaaaat aggaaatcac catttggcca
124921 atgccacagc aatgggtgtt gaatgaggag gggaagaaat aatgaacagt gtgtttagag
124981 agtcccgaga tctgtctcaa agtacttctt aattagagat aaaaataaag atgttaagtg
125041 acgagaggac cttgcagaga cagcattaac caagtgtcac aattacctaa gcatggggca
125101 tcaggtgccc tggcaggctg acgtactgac tcttcacagc aggaaagagc acaggaggct
125161 gtgtgtgggg ctgcaaggaa tgacctccca tgaagtacct ggatcgagct actgaagtgt
125221 gggaaaagtg tcccggaatc ggagaaatca aggaggacag gaatcagagc tgggtgggca
125281 ccgctcctgg atcaggaacc acaaatggca caagagaggt caaagggcac tgggaagaag
125341 gggcagatgg tggcagtaca gtgcaggtgc taccttcatg gttcggacag cttcatcggg
125401 gttttgtaag atgctaatta gaagatgtca gacacctccc accccacgt gtgcaaaact
125461 gaagcagaaa tatttcatat aaaacaaagt gtttttttt taagttccta aggtaaataa
125521 aagcaactta gtgacatatg gaacacaatg tccccgacag tttgacaata aggcaacact
125581 cttccttcgg gaggagaga atctctactt tgcgtaggga cacatcaagg atatgcttct
125641 ttggaaggtc ttcaggctgg cagagctggt caccaatggg gatggtgtgg gggcgtttac
125701 gttctcacaa ttccccatct atctcatgtt accctccagt gcaacggaag tggagggtac
125761 actgagtqtt catgaagaga aagccgttaa cgagatgctc aacgagaata taaccatctg
125821 togcacttqt cocqcatqtc ccatqccctt ttcctaggct gcaagcacgc aagtggtaca
125881 cagatacaca cccaggcaaa acatccatac aaactaaaca cacacacaca cacccttttt
125941 gggtcctttg gagtccttgt ttatgggagc agacattgtt ctgtcttctg tcacagatag
126001 catcctctga gcttggaaag tttaacatga gattgatccc actctttcca agggggagag
126061 ggagactaat gtgtaagcag acagctccca gtatcacaag ccttagtttc ctcatctatc
126121 actatgtggt ggtagtgagg gcagtaaaaa gaccttgtca agctttgact agaacaacct
126181 gagattette attetgggag tageettaaa tgtaactata tteaetggea aagaagaaca
126241 aactaaggcc ccaggtgaat gtaagaaggc taccattttt acttcggtga tgctgagtgt
126301 taaggcgtgt atagcacaac agaggagtac tccaccgcaa agggatgcac tgtctctttg
126361 gagtcgaggt gcccctcca ggtaatgaga cagaaaggag tggatacagc tggtccaggt
126421 tettggetgg etcegagage tatgetaagt cetetgaate aatggtatga cacaagtttg
126481 ttetetgttg agetggtgte ttaetteate accagtgete atetetaaet eeegagetea
126541 agaaatcott ctgcaggatt ggagagatgg ctcagcggat aagagcactg actgctctcc
126601 cacaggtect gagttgaaat ggeteacaaa ceatecetaa tgagateega tgeeetette
126661 tggggtgtct gaagacaget acatgtactt acatataata aataaataca tetttgggtt
126721 ggagcgaacg gggctgggga gagagcgaa aaagtgtctg aagatagcta tagtgtactt
126781 atatagaaat aaataaatta aaaaaaaaga aaccettetg cacageette tgaatagetg
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 61 of 83

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126841 gggtttcagg cttgagcccc ctttagagac aggcagcaga gacacaaagg cctgcccaac
126901 atgtgtagga ttagatagcc tgggtatggc tgaagggaac aggtactcag gttggcccag
126961 gccaggagcc agactetagt atattatgca aggttgcctc ctaaattata atgcacttca
127021 tttgcagaga gtccaacact agactcatgg tgtctggtag cccagagggt aacaggctgt
127081 gttcttcaca gggaacatct gaggacagag acggaccaag agggctcttg acggtggctg
127141 gtggtaagga ctaggagcgc ttacggcgag ctgcttggga gaagcctttt caggtggtta
127201 tctgagagaa ggcacaggaa gtcatgaaat taggacctga ggacttgtcc aggataggcg
127261 cagtagtgag aactgcatcc ttgtgaagat gggtttgacc tctgctggag gctacaatgc
127321 agctaaatgt gtctttatct taatgactcc agaaacgcca tgctgtgaaa gaagcaggta
127381 ggtagaggga ggtcaggaga gcaaaccaac ctcacgtgaa gggtaactca cagacttgtc
127441 gttcactttc cagaacccaa ccctcaaaag aacatacttc tagggtgtag ctgggaacca
127501 ggaagcagcc agetetgage tgetgettge ttgetetete acacagtggt aacaagcaat
127561 gctcatctct tttcaaagca ctccaagcaa ataaacacct gcagaggagc tgatggagac
127621 agacttetet tactggggtg agtetattaa acagaaaagg aggeataaaa atgataaace
127681 cagaccgtca gaacacagtg agtgcaagaa actgtcagca gagagcgaac tgcttccttc
127741 tcacgccgga gggcactgct ggattaagca taatacaagt tatgtatgct cgctgatgtt
127801 cctgagcggg taagaaataa ccagtgttca ctagagtaaa aagtgcagcg gttctgaatg
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128161 agggatggtt tgatagtgaa gagcactggc tgctcttcca gaggacccag gctcaattcc
128221 tatcacacat ggaggettag aagcatette tteaggagaa gacetacagg ggaactaata
128281 tccttttcta gtgcacacat ttaaaactga ggcttagttt aaaccctgtt ttaccagggg
128341 caggggtgtc tgtgcacaag aaagacagac agagtttagt tttagttcta gacttctatt
128401 tgctcaccag gaaaataaca ttggtagact ttaccaaaat ctgcagcaaa aactaaatga
128461 taaatgtact ctcaaaacag ggctcaatat cccacaggtg atcagggtca acttttccag
128521 ggaccctggt gtcagtgggt tttatacttg gaatgagaac acagtaaagc tgggactgcc
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128641 ggcttctgtc cctcagggac tggacttctc aagagcagaa cactgctgta ggaaacagtg
128701 tcttccaaga gtcaagggaa ggaggaatgc caccactgag aggagatggc ttgctttcct
128761 ccctcaagaa aagagtgagc aaggctccat ccacggcgtg ttccaacacc ttcccacata
128821 gctggaccca gatgtcatag gggacttgca gaaaggaagt ttcaacatca gctagccttg
128881 gcagatgctc ccaagtcgtt caaaaataaa actcgggcga tgacggtaaa gatgggatga
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129061 tecagaaagg cagagtgtgt gettetgaag geagtgttet ceageettte tacettecat
129121 taaatggttg agtctgtcct tgtgtggatt gatgttattt ctaaagcaca caattcattt
129181 caagagggga gaggcaggat taagctggaa atagtttttt ttctattgag aaaaaattaa
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FIGURE 6AAA

Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-IR)
Polymorphic Alleles And Use Of The Same To Identify
DNA Markers For Reproductive Longevity
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129241 gtgtaagtgg aaacgttttc atgaaaatgt ttttatatct caacgtttcc tgcgagcact
129301 ctgatcccaa tatatcacag gactgagctg gatgagggag aatgaaggca aaatgaattt
129361 aaataaaatt gaaactgacc ggctaggctt tattctccaa atctcatgaa atagcaaagc
129481 aaaaaaacaa gaagcaactc aacgctttca gctcaaatga gtttcaattt tatttcaaaa
129541 tgttttaaaa tctagtttat tttcatagec acaaaagett ttgaataeet atgaatcaaa
129601 agccatttgg cagaaccatg gagcttgtcc tagttgcttt ctctcaaact cttccctggg
129661 cctgttaacc ccttgttagt cagtcacgca cagcaggtca ctctcagtac ccagttctta
129721 ggggcactct acagccttag tgtgtatgag caggcagagg ctctaggctg gacggcacat
129841 gaaaatgata tatacagact ttaactacta attactttag acaaagaaca aacagaaaac
129901 aaaaaacaaa aacaaccccc tccccaaacc caaaccaaaa aaaccactaa aaaacaaaac
129961 aaaaaaaccc caatccacat cetttagaga ttattttgtt aatctgcate ttgatetgtg
130021 ctgctcaggg gccacaggag gccacaggag gcctctgaaa cttgaaacag ttaagattaa .
130081 gtaaaaagtg caacgaagag agctcaccac atcctaaatg tttctcaact actcaactcc
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130321 gggactgcca ggtgtgtgtt actctacaca ggttacgtaa ttgtcaggcc tttgtgtttg
130381 tgtgcatttc cctaagcaac ttcatgtaca aagctaagtt gggtatgcat tctctcccca
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130501 ggtccctacg tacagagaaa ggagatattt acagaagaaa gggcaaatga ctgcttttac
130561 agggaaagag catcccaggt caacacacgt accagattgt agagagggca ggtgtctcca
130621 teccegtgee tetetattte etttecaaca etcageeetg tgtgageaga gagtaaggee
130681 cctctttgat gctgcacccc cctattataa gcacaatgct tgtaaccaaa cagatggttg
130741 gatgtttggg aaatgactga atgaaacaaa agaagaatgg tattctgtta aacaacagcc
130801 aacatecete tagageetgg acacaegtet ttatagagee tggatacaag tetttatatt
130861 ggatagcaac ctctttactt tcaaatgcaa tctcagagca acccagaaga ctgattcagg
130921 tagtatacca cagcicagic tettecatge teaacaatgg etecticeaa atggeattag
130981 cgttgtctac atatctgtac agtcagaget aatgtctatc tgttcgtctt taaatggccc
131041 agagetttte actttecata tactatttea cetaccette acetggatat tgtetataca
131101 ataaagagga ggaaaccaag gcttgggcga aactgacttg ccagatcaca tagagttata
131161 ggtatataaa tggaaaagct ctggtaaaga ttacagacta tgctgattct attagctggc
131221 tgtaggacag ctgaatggat ctctttcaag gaagggcatg atttgtaagc atcccacacg
131281 tragtgrtga gtaaggetee taagteetge tggettgetg cragcragta gacccettee
131341 agggtgaggg aactgggcag agctcagcac aacccagctc acctaactgt ggctgtcaga
131401 atcattggat cctgcccatc atacatccac tctgtggcca tatcccgcag cttcacaagc
131461 ttcatcaggg tcgggcctgt gtctgcaaaa agaccagctt gagaagacca ggaggtgtat
131521 atcaagtacc actcgttcat tcgttcgttc attcactagt cagaacttca aaccaataac
131581 cccagaagta aagacaaaaa ctccctctgc cctctggtgg cttgaacttc cctcttggac
```

Applicant: Farld, Abdol Hussain et al.
Attorney Docket No. P05562US00
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131641 ccaattccca caggttaggg tttccagaag ctttcaaaaa aaaatcactg atgaaacact
131701 agetetttag ettagegtat tatacaaaag getgatttee gatttateaa agggttaeca
131761 aataggaagt ataaaaaccg tatgatcctt acagggaatg gtcaaagagc agtgatcaag
131821 tccctgattt acataattct cagccattct tttggcagac ctctacagcc actaacattt
131881 ccatctacgg atgataaaaa gataactctt cccccttcct gggtgcctgt caggtataaa
131941 cttgaagccg ggcacagtta atcctcacaa ctccccaagg aagaggtatt accatgtttg
132001 ctccagatta agaaagaagg gtcaggcctg aattccaggc atcagaattc tactgagagg
132061 gaagagagac agacagacag ctttcactct ggcctagagt gcagagaaaa ctactacttt
132121 ctgtgggaag aaaagtcagt caaccacagg aatcacagaa ggatccatca caagaggacg
132181 ccactgtccc ttaagcagag acaggactga gaagaacaaa ggtactactg ggacagaata
132241 tggagaggtc agcttgagga agtgctctag caaatggcat caaggaggcc atttccccaa
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132421 aagaagaact gagagccagg ccggggccac tgctctgccc tgcttatgaa agccagggga
132481 ttaagtcaag acttcactcg tgattcaaaa agttatctga agacatggcg ggaagtacag
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132661 ctggcactcg gaagctgttg gcatggaaaa attaacaggc tgacttggcc agggaggaac
132721 ggttaacaca aatcaccagg aaaagatgag ccctgagcaa gcgggaggtt tgagggaagg
132781 gtcacggaaa aaagaagggg taagacacat ttgtcaggag tcccaggctg gtaccaagcc
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133921 ggcaaccctg tcgcaaataa ttcatgtcca ctacgcccct accacacaga atacaactac
133981 attgtgctgc tagaaatggg aaacctgatt cagatcagag aaggtagaaa aacatttagt
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FIGURE 6CCC

Applicant: Farid, Abdol Hussain et al.
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134041 tcaacacaat actgccttta tgaaaagggg cgtgcgccgg gtggtggtgt aaatcccaaa
 134101 cctgtaagtg gggatagatt tcaagacgaa ttttggaaga aaatgcagaa tcaaaatagc
 134161 atccagcttg actcagacca gtaataataa taacaacaac aaaaacccag ccagtccttc
 134221 tagagetacg cetgtttagg etgegeggtg etettaatgg ecagecagec tgtteteetg
 134281 ggaaatggtg ctgctgctac agcgtgactt gctttgcgtg ctgcaataaa aggaaaacaa
 134341 gcttgcacga cccaatagct tccttcagtt tgtgcctagc agcggggaga aaattaaaaa
 134401 aaaaaaaaaa aaaagcctga aaaggggtcc tgaggcaagg tgaatggagg acaggatcct
 134461 agagaacagc actgccctgc aggatttttg taaacagcag cctgtaatgg ccacggttag
 134521 agaaagaaaa acagacatgg agggaagcgt gtttacaact gcagcaaggg cgggaggcgg
 134581 agggaggat teceeggete tgegetegge cetetteact eeggtgggge eeeggggttg
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 134761 ttgcgggatg ccagcttgac tgacaagtcc tgcagccaat cgctggttgg gcccttagga
 134821 aaggtggaat ttcccaacaa actgattggc tgagagagag ggcgggccct ggtgattggc
 134881 gtgttattcc gagtagaaag tagtaaacaa ccttggtcca ggggcggtgg ggagggcgcc
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 135241 ccttcctcac cgtcgaataa aaatttaaaa ttgtctttta cgaaagttgc tttaaatcac
 135301 acactaaaat tgttgcacta tacaatacgc cattaatggc atgctggtgc accagacaca
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 135421 ggagaacata cacaaaaaga aacaacacat gacggcgttg ggaccacttg ataagaaata
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 136081 gttggttttt atgctttcta gactttacct ctgcctgccc aggagaggcc actgttctcc
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 136261 tgaacttcct gaggcctgga atggtattat cttcagattt gggtcttcag tggagatggg
  136321 cttctactaa agtgcccgct gcaaacttgc caaatatttc taggattctt ccctagcatc
 136381 ctgagtatgc ctacccctga aaatctttcc ttacacacgt gcaattaatg tgttttctct
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R)
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136441 gctgggtcaa caactctgtt ttctctactt ggcaaatttc cactctgcct aaaatcactt
136501 cctctcttgg catacctttc acaactccc accaactaca ctgtgtacca agagcagaga
136561 ggtetettte egttettaet tetetggeca gaattageta caaaactaga actageatgg
136621 tttgttggta aaggaagagc ggagaggcaa aatttcagaa gtggtatcac ctgtacatct
136681 gcataaccaa cccattgcca tctgtagggg tgaagcagcg ccatgctccg tttgtggctg
136741 tgatggtete eccetgettt etgtetgaag geetetgtet etgaggtgga ttettaetee
136801 cccctccatc tcaatcagag atctcctcct ggctcttatg ctatcaactc tgcacatccc
136861 cgattccttt ccaccatgac actgaactgt catctccttt tctatctgac cagtcactga
136921 cttgctcaca gtttagtctg tccgccatac agccatatcc agtgttctaa gaattgtggg
136981 gttccacagc tggggctcag tctctgctta cgccctcatg tgacatctgt catcaaccct
137041 ccatctttct aagcctttaa tggtggaaaa gagtaaacct gtagataaag agtggtacca
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137281 caaacatgat ggcctgagct tcaggatgct cattaccaca ctgcatcctc agtcaagaag
137341 cagagagaac tggtteccag gacagggget ggtgetgeet aagtettggt ettecegtet
137401 ttaagttaac acggcccttg gcaggcatgc ctgaagcttg acttttcggt gattacagac
137461 cctatcaaac accataaata ataacaccac acatatatta catataaaa aactataaca
137521 cagaacactc aattggcttt cagtaagtgg gctatttttt tttctttgtt gcttaatgta
137581 ttctagatct gtctatgttt acaaacggta tgtcttctct actaacaaat aggatttcaa
137641 attaaaaagc tgaacagcaa aggagacagt gaagagacaa tctgttagct tttcatctga
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137761 tcaaaccaat aaatgggcaa attaatcaaa tgggcagttt tcaaaagaag cacaaaagac
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138421 gccatcttat cttcacagac agcattctgc tctatagata accctgagtg gtccccaacc
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138601 ggcaggtttg agagataaca gtaaacctat ttattatccc tctactgata gaacttgtca
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138721 tgtctgtggg tgatcatcct gggagaatta ccatgtcaaa gggcacaggc actggtacat
```

Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity
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138901 ttcctataga tctgtcatca gaatctgtga ctgaacattt tagattttca agcatggggg
138961 gctgggagga gctctgaata ctcataaaaa ggactttctg atgtgttcta ctacaacatt
139021 tetggttett ttgtttgttt gttgttgttt acacetttat tatgaattea ettagagtet
139081 aacaaggatg gacccatgat ccaacatcag ttaagtaaag agcgtatttg ctccaatgat
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139201 tcatattctg tcccttcaca tactggcacc ttgctagcta tttaatcatt aattacttag
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139321 ttatcaccat tattattgtt gctattattg acgactgtag agtcagtttg tataattaaa
139381 agaaaccaca tagtatttga agtatattaa ctttataaac acaggaagaa ctgacatctt
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143581 gacacacgta cagatacaca aatacttagc aggacgtaaa cttctatttt ggcttcatga
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FIGURE 6HHH

Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 69 of 83

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FIGURE 6III

Applicant: Farid, Abdol Hussain et al.
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Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 70 of 83

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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R)
Polymorphic Alleles And Use Of The Same To Identify
DNA Markers For Reproductive Longevity
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Applicant: Farid, Abdol Hussain et al-Attorney Docket No. P05562US00 Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 72 of 83

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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-IR) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity Sheet 73 of 83

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 157441 tatgggcacg tgagaggaga ggtgaagaag cctcaatctt tctgagccca tttcatctcc
 157501 cctattattc tccacgattt tcaaaagcca ctctactaca aacctgagaa acacaggagg
 157561 tcagattcac aaatgaaatc tctttgtctc ccttctgtat gcacccagca tctgctccac
 157621 ctagctgacc gttacagggg cagggccagt gttaacatgt caagtgggca gagggctctc
 157681 tgcatctcct agtacatttc tggctacagg tgaactaaat gacaccaaag aactgctctc
 157741 aaacgtgcat cccaaggtag agctacaaag gttcaatata accccagtat tttgagagat
157801 cttaaaacgc agtcaaagtt aactgtcatg tccatagcaa ggaggaactc actccttagg
 157861 atacactggg ctggtcttcc agtgggcatt tcagagagcc ctgctttccc atatcagctc
 157921 catcetgtat ceteacegtt getteeaaca gggaacegte cacatetgae tgeecaatea
 157981 gatttcagca ggtggcatgg gggatccaga aagcatttct ctgcaccttt taactctgaa
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FIGURE 6MMM

Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R)
Polymorphic Alleles And Use Of The Same To Identify
DNA Markers For Reproductive Longevity
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```
158041 ctgcatgcag aacccgtggt aagaaatgct tctctgatcg ctcctagcac ttttagtggc
158101 catgttaaat cagctaagta agaatgactc ttccatctgt gttaagtggc cttttaaagc
158161 taattcacac tatctccaaa taccctgaat taccaagtct gagggctgcc tagattcctg
158221 cttgtaagca atagaatgac actgaggact ctcggctctc ttctctctc catatcctct
158281 ctggttcact gagaaccaaa catcatctaa acagaacaaa aacatcagaa ctgaaggaat
158341 cgccaaagta tatacacgtg actcccgtct gaatgcattc agatccgacc agtgactcca
158401 agetttetge atgttteeca tgagetattt geeetgaate aacteeteae cacaactete
158461 gaaatgacat ggacctctgc tgttgcactt attattcctt ctctgatggg gagtggttat
158521 gactgaaaat tttcgcgcaa ccagcctcta aacagaaatg cagtttattc tgagtactca
158581 aattcaaatc tcagtaacac agcaattaca ttttaaaata aattctctcc atgttcttgc
158641 ttctaagaag agggtttagt gggatatgaa gagtctaatc ctaggcaacc atatggccag
158701 gccaattata aaccccatat atgttcacac atcaagcagg gcaaactgct ctgggaacaa
158761 cccttaatat tttattgcct gcccaggtgc ctgagcactt ctgagataca cctgtcacag
158821 gaaaatacag cactteette catetttace ceatagttge cagacaattg ggcagageta
158881 aagtgggtaa tgttgcagtc aacacttccc taaaacattt atacatgcca ctgataaaca
158941 atagaaattt aaaggagata caaatgtact ccccaaaccc taaatcttcc tcccacggag
159001 gcctaaaaac ttaaaaatct tcatctacct ggcacagaag aaggaataca aattccaagg
159061 ggagcctaga tttatggcct aaatatgtct tcctctagaa tatttaatac acttacgctt
159121 tagtcattcc atacattttg aaaatactct agcacgataa attacagtag ctgcatcttt
159181 catggttgtt cacataaata tcatttggct ttcaggaaac caattacgga gcggtaaatc
159241 atttttacat tgcttttcaa aatctttaat ttaacttgca acacggacag ggcacacaca
159301 ggctcccagg aagaacgcag ctcacacatt gtatcattcc ctcttgtccg agtttagcaa
159361 caacccctga gtaagccaca acaatatttt catgttctaa aatatttggg ttatatctat
159421 atatttcaca caggitattag gctcagcaag tatttcctaa cctcctgtag tatttagttt
159481 tcaagacttg gggtaggatt gggaccacat tgatttcagg gtgctgactg tacttctgtc
159541 accatcaget agtttaagga caatgactgt cetggeagac accgcaaaca cagageetet
159601 atggtgttct tgagaaatca agaccatcat ctcattggaa aggcggtggg aaacaaggtg
159661 cagggccatt gaggccacac aatcaacaac aggatttgct ccagcctcag agacacagag
159721 tggactggca tactatatca atttggaatc ccctcggcta tgacaagttt gtgaaatgaa
159781 agaacaaaac aatacagtac aagatcaagt tttagtagag agaatgcaga caaccacaac
159841 cgagtactta gggagaaaaa aaaaaaaccc agctcttagc tggggagtga cagtcaaggg
159901 gggaaatcac gcgaaggaat ttctttcagg tgcaagaagg catgagagtg ggagacaagg
159961 attgagcatc ttggcagaag gatcagaaac ccagtccaag gtctacactt ccgtggagca
160021 gccgcagaca agcttaggta tccggtgggg ataacctaca gttaagacat cacattacag
160081 acactaggga gccattgtga attacagcca acaggtgaga atcataactt agcggggcgg
160141 acacaacaac caagccgtga tctagagcag gaagcggggg atagacagaa agacaggttg
160201 taatggagga gaccaagcag agggctgcat taaggcagag gcttctgtgc tgaccaagtg
160261 actaacacaa agggcccttg cagaagacca gagatgtcag gggggagaat tccttctgag
160321 ggggcctagt tacactagcc acttgatatt caccgggaag gaggcagaga agctaggagg
160381 gtgggcacaa aggcagtggc tctcttctac ctgaggagag ctggcagctc tatcaatacg
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FIGURE 6NNN

Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity
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160441 gactactcct aaatgggctg cagtcagccc agaaaaaaaa aaaatcactc aatactctca
160501 ggttacctac cctggctccc aaagctggtt atgcaagaaa tattttgttg ttgttttgct
160561 tttaaaagga gacaactgat gcttgtttta gtattgtttg aactacatca ctcacaaatg
160621 agagactgat aaagagattg acaagtctga ctctagaagc aaacacttcc agtctgctct
160681 catctgccct ccccacttc ctccattctc ctcggcctct accccacat atgcttgaaa
160741 atagtaggta actgagtttg ttttgtacgg tggttgtctt gttatttcaa gcagggtggt
160801 ccagagagaa aaaaacattt tctggaaaga ttttaactac tggatcataa acaagcgtgt
160861 ggtggccacg cagcaatctt tgtaataacg aatccaagac gtcctggttg tgaagcccag
160921 tgaatgtaat gttagetgaa aageacegee tgacacacet ggaaatgtge caegggeeag
160981 ctttgtagac tgcgacttca aagggaaact ggagtctact tccgcttttc agtgagttag
161041 aaatgatttt ggctgtattg gtagggcaga accagaaaat aagttttgaa tttgtcataa
161101 taatttacaa aaaccaacca acttgttaag atgcatecte caagcacaac taagetgget
161161 atagaggeta ttetgeatte tgeatetggt eccetggaet geteaggett agggtgaggg
161221 aagaaacatg acttaagcca ttacccttat tetgtgggca tttgaagtgg etgecaggca
161281 cgtgacacca actgttccaa ttggttcaca atgacatcac tttattattc aactctgttt
161341 gcatttcagt ctcctccttt ttcgggggac ttgaggccaa tcctggccaa gtcactttag
161401 aagagaaaga agccagtctt aggtagtcat gaagtgccaa gtgtcttttg tgtctcctag
161461 cctagtttgt aagccgggaa ctggtgacat aaagggctct gtcaacagcg cccgggtgtg
161521 ctgcttccct cgcagcggga gagggagatt gggaacccag tagaaggtgg cggttcggcc
161581 ttgggtaggt cccaactgac caagttcaat aaaatccctg agagtttgtt ttcaattggg
161641 taagaaaact ataaaatgga aaccgcatgc gagcttacgc tggaacctca tttaaagcac
161701 ttgtacttcc aaaaaaaaaa aaaaaaatgc agaaaacatt tctaatttag tcaagcgcac
161761 tcttctttcc tcgagcaatt agtcttaatt catgtcacct aactactatt tctgcattaa
161821 gcaacgtgac ctctagaacc tacagtaacc ccctttctga tggtgaaagt caattattct
161881 gatttttttc atcttcacac aataagttac cagagaataa actaacttca aaaaagcatt
161941 acagtgtcat gtgtgttcgc ctgttgatat gtgcacacat tcatggcacc tgtgggtctg
162001 tatccagete ttetaetgtg cetgtetete aattttgeag eteaacetag agateaceaa
162061 gtggccagcg agetecatgg atcagecegt ttetgecete etetgtaeta agattacacg
162121 caatcatgct gagcctctaa gaacatggcc gctcatggtc tgaactcgga tcctccacct
162181 agcatggcaa acatgttagg aacggggcca tttcctcaac acaaaggaaa aaggcttcct
162241 catgggagag aatgactgga tgggcgagta gacagatgaa aatggacttt tcacaccaac
162301 tggttagaga ccctgttgct tccaggcaca aagcacacca gtttctagta tcagtgggtt
162361 gaaatgcatg tttaagctct ctcatcctta cttggctctt gacaggaaac ttttatggtt
162421 gaactttatt ctgtagaaat gcagtggcat taaccaagca cacaaagcca actcaagcct
162481 aaagtgtgcc catgactcca accctacact cattggttct tgccagaggc aacacgaaac
162541 agtgtataga ctcagaactc tgcccaggcc tgtgaggcag ggccagggct acacaccttg
162601 caaacacaca gcaggetetg eccaaggage gtteactagg taactateag ggtggeteaa
162661 gggctgtacg acagacagac cctgcacagg cagggaaagt ttcctggaga cctaccatct
162721 attttcttgt aatctcaact gatgtgttct gccaagtcaa ctattattta tcatttataa
162781 agcageteet attggeagea tactetgeat atgttteage aetteataaa ceeteeaegg
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FIGURE 6000

Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity
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162841 tataatcgct tgtggaggag actgggttgc ctggagctgg aatacagacc cacacgcatg
162901 tgagcccgtg taaatgcaca catgaccgtt ccctaccgct ctttcacgag tcatgtcttg
162961 gctctgtagg ccgagtatct ataactaaag ctctctacag gtgtcaaatc aagcaatctt
163021 ggtcctctca gatgtcagct aatggcttag agccccccc ccctaacagt tctatttgca
163081 ttgctaacca aggctatcca ccttagtatc atcatcagtg caagagaagc caaactgcac
163141 acgaagacct ggagagcact tcagagcaca catccgacct cttcctcatc cgagtaccta
163201 gcaagccagt tttccttcct ctcctgtctc cagaagaagt tccaactcca cctcggagca
163261 aagcagcagc tgcagtgact agcagtgact atgcctttgt gactgcgtat cactcatcca
163321 attcacagac tacttagcat cgacttgtat acactttgcc caagaataaa tatttatatt
163381 cccgaaacac agtcgatcga gagcagctga tacagggctg gagatgacag aggcagcaaa
163441 cacaaacaag taagacetee aggagtgtga gggacaggge tagagacaaa gtaaacagag
163501 gccacaagat cctaggcgct aggtgggggc tactggtacg taagggctgc tggaaagaag
163561 agtcctccgg ggactggctc ccacaaaggc tgcccaatct caagtggtca gctctgggcg
163621 cttgtctaga tggcaatgct gcgtggactc agtgggttat gtatgcgtgt gtacatgcac
163681 atgctaggat aataattaaa aataagaggt catgagtttg ggaggttgga gtttgctcat
163741 gggagcaact ggaagtgggg ggcagagaaa gggctgggag tgacacaggt gtagttcttg
163801 tgtatggtct aaagattaaa ttagagttag ggaaaaaaaa accccacagg aattgggaag
163861 agtgattccc aagccactga cacagcacag cactgcttgc ctgtggaggt gctcctgtgc
163921 catcctccct aatctttatg ttgcacatct taagagtggt ctgtttgtgt atcctctcat
163981 gcatgtatct cctcagcgtt ctcatccacg aatgcttaca ggtgacgtta gcatcagcag
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164161 ttgccactct tcaaactgaa ggtctgtgat acttaaaagg gggaaaaaac ccattgaaat
164221 gcagaaggca gagaccatca ctcaaagata ctcatcgctt ggaggtcagt ttatatttgg
164281 aacagtgact tgggattggc caaccatcac tcatgccatt aaagtaatat ttcagtttca
164341 actgttttac atgcatttag catgtagacg tattcaattt atagttgtaa acggtctaca
164401 agtgtaagaa gccgataccc aagcttcgaa tttgtaggtg tttaagtcta agtcccaatt
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164581 tecteatege caggaggaag ttggtttata cecateatag tteetttaac acaagcaatg
164641 cagaatggcc ttcacaagtg tctcctctaa aatgactgtt taaattgtgt tgtttcaaaa
164701 agaggacatc cacgagccgc atgtttccat tatcccttgg aactggagct acaggcagtc
164761 aagggettee ttacttgggt getgggacae aactetggee etcatggtag aacagcaage
164821 ccagcaacct ttctagttcc aggattaggt cttaactttc gtggtctctt gatggcacac
164881 agaaattgtc tagaagtcag aggctatgta cgggtggatg atgtacaccc agtcgccctc
164941 caagcaagcc aaatatgaaa gcaatttttt ttttcaaaaa taaaatagca ccacactgct
165001 ctcccaccag gcgtgtagag attttcccca acaaaatatg acacactca tatctatgat
165061 tgcttctaga tgaagaagtc tttaaaagtt ttaacttctt aagggccaaa tatcaatgga
165121 tatattccat ataaataaaa attcatcaga tttgtcttca cctattttgg ttgctgtaac
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R)
Polymorphic Alleles And Use Of The Same To Identify
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165241 ccaaccaacc aaccaggtag tetetgtett gttccagtec catttteetg gtgcacaaaa
 165301 ggactgattt tacttgttag aaggttatcg ggcaattctc tgtgtctctc ttacaagggt
 165361 actaagcatt ctcaggaggg ttcaatcttt atggcccaat cactccctt cgccccaaaa
 165421 caatgtcttg atgaccttcc aacacagtca tggcagatta tggtctgtag gctttactgg
 165481 tgccaagggc tcctgagaat acaacaccaa agtgatctca tcccaagaag gatgcttcag
 165541 acagacagte ceacatgett teetetacte etggeteate ecetgtatte catacateta
 165601 tatcccccgt ccctcttagc tctcctctga gtggcaaaga ggcatctccg acttaacgag
 165661 atgcaaggca gagttctagc tccatctgcc atccccacca cacacctccc aaacaaacct
 165721 actecettee teetgecage tegggaacgg geaactteae ttetegttte atetaageea
 165781 aatgcctgaa tcatccttta tccctctttg ccccacctgc aagatttcac aagtccctaa
 165841 atattcctga atctatctac tcttagttac tgtaactgat gggtgcagtt ctcaacaagg
 165901 aactettetg teeeggeact acetgeatee agceatgtet agatacattt tteaattgtg
 165961 tctgcgtaga caaggggtag atgtaccacc agtatctagc aggaagatgc atagacaagg
 166021 ggtagatgta ccaccagtat ctagcaggaa gaggccaggg atattgatag acattctata
 166081 atgcacacag cagetectat cateaaagae tgtecagete aaaatgteaa tggcattgag
 166141 aaactgaget ctattecaag ceagtagaat etetecaaaa aaacettete aaaacaaett
 166201 cctaactgtt ttccctgatt tttcactctt aattgctctg ctcgatgaaa acaactaaag
 166261 ttatcagcag aaatacagag tgtgacccac tagagtgctt tgtcttgtgt catggcctgt
 166321 aggecettge etactecaet ggeceeegge eagetectae atteteatee caccaaatte
 166381 ctacgtcaga gccctaagcc ctcaggtgac tctgctagag acagactcta cggtagtaac
 166441 tgaaggttaa gtgagcccgt aaggaggaac tgagccttgt aatggttggt gccttcatga
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 166561 tctgctgtga gaagatggcc atccacacag aaaaaggacc tcactggaat ttgaccttgc
 166621 tggaactctg atcttggcct catagttcta ggactttgag aaaagaaacg tcagcagctt
 166681 aagccaccca ggctgtggta ctttgttgtg aaagcttgga ctaatacatt caaccttgct
 166741 accttgecaa gttccagttc actggccatc agtccactta caccacagcc cttgccctcc
 166801 ctcttctgaa ctgccattag ctgaccccta ataaccccta gtcaaaatgc caccctcccc
 166861 aggagggact gtgttactcc atcettectg gattttgagg acagececat ggttggcatt
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 166981 gtcttgcccc ttgtcgaatc aggataagat aatctgtgtg gcttatatct cttaaactca
 167041 aaacaaggaa aggtaggggg tetgtgaaca tgetttgeee catgacceag tteeetetge
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 167161 gggcaggagg gaatcattac atgtaacatc tagaatgagt acttccttat attctttcac
 167221 tttagggagc agcaaaggtt caaggttacg gtaaagtaca ggtatggggg catcaccctg
 167281 atatgatgaa gcaccaagga actccccaat gttaactatc tagccacaga gcaggtaagg
 167341 gtatgctcca gttagtttta cgaggtcaac tctgaacaga ggaaacaaaa gcaagcagta
 167401 aaggactatg aggcacgtgg gtttgtgctg tgtacagccg agcagtcaca gatgcctcac
 167461 taaggaggtg gcatggggac taagagtgac tggagttagc catgggaaat tcaggcaaaa
 167521 agcaaggaag gtctctctga actggtgagg aacagcagtc gtcagtcagt attactgtcc
 167581 gttatttctc tacactggcc ccaggcaggt gcttttgaat gactggctgt ttgcaaaggg
```

FIGURE 6QQQ

Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity
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167641 aagtccagat acagatggcc gaatgagggg ccacgagcct gtgcaggaca tacagataat
167701 ggaacaggaa cettgcaatg etgagaceta etacgtgacg ggtgettaac agettttgag
167761 gttaacggtg ggaatttctg agagaatccc aaagagagct agctggttgc ttgaggccaa
167821 caagggcaag gettataagt caggggcace ceagaaagga ggcacacece agaaaggagg
167881 cagcttctaa gaaagaggaa aagagagaaa gcaggcaagc ctgagaaatc aggcttgggt
167941 ggcctgaaaa ggcagaccat gatgtgtgtg gtgggaaagc agggcaggac tcctcctct
168001 tctggaggaa tacatgtgct ggcactggga catccagctt caaccccagg agcccaggcc
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168121 cgatggaggc cctgtcccat cctccaagat gtctaaaggg ggtgggactg gctaagtccc
168181 teettetett ceteacece teeacateca acetecteca aaaacateaa atattttett
168241 ccataaataa ttgcaacagg atgccattgc tttcctcaaa gaaacaatgg cctgattgtg
168301 ccetgteega caegggeeca geeceageae acacaaaggg tggettteag aageeetget
168361 gtagagaatg ggggtacacc teetttette tettetgtae caagteeact geeeteeace
168421 aggaaaggac atcaggaagg ggctgctacc acaggaataa ttcccatcag aggacagcta
168481 teatteacet geetttagat tgeeetttgg ceaectetag aaagttette gttetggtga
168541 acagaagagc acctattggc tctaaaatcc aacacacac cacatacagt tttaggctgg
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168781 tctcaaaaaa aaaatacata agtaaataaa aattataaat agataataaa atagtcccat
168841 tttacgtgta catcttcagt ttcacaattt gagataaagt gaatacagat attttcattc
168901 agtatctgtg gtagtgccaa cactggatgc tcacacaggc tcatgcatgt gaacatctgg
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169201 ataagccctt tattacttaa gttgtttttg taacactaat caagtgtgtt tcttggttaa
169261 aaaggacagg ctggactaaa ggattcaatt tcagcactta aggggctgga gagatgagag
169321 agcctgctgc tggggctgtg ggctgggata agttgaggcc cgagtttagt tctcagcacc
169381 catgtttggt tcccaacatt ccatgttcac cccaggcctc caagggcctc cacacacata
169441 gatataccca ccctgagata ttcgcatgca tataaataaa cataaaataa atatttaaag
169501 aacccatcac tttgcagcca tggcttgaca aacaggctct ttggtgtctt cctcacatag
169561 gtgcaagagg aactcaggca ggggtgttga gtttttggat tttgcactta tttatttatt
169621 tattggtggt actetetgtt tattggggga gttgtggtgg gttgccattt gtaacattte
169681 cattaaagat gttaaaaggc tgccttccca ttttcaaaga ctcagactga tcactctcc
169741 agccagaggc ttctgtcagc tactaaaggg ttcctttccc ttctatttta tggcctttct
169801 cccatccact tctgctttgt tcttttaatt aaaatgaaat gtaggccctc catggctgag
169861 atgctaataa ggcccctgaa agtcatctgt gagcacatcc actcttgttt aaagagccat
169921 gtaaacagac cacagacggc tacagagagc aacgctgcac agggcaagga aaagggaatc
169981 tggctgcacc gtacagtgct acgttgaagt gatttatcac ttatgccagg aggaggaact
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Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R) Polymorphic Alleles And Use Of The Same To Identify DNA Markers For Reproductive Longevity
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170041 gaaacacaaa cgcaatttca agtttaagca tggcgtctcc acaggatctt ccatagcaag
170101 cactetgcac tegggggete cetgteagga aacteetgga caaeggttgg tttgeegtet
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FIGURE 6SSS

Applicant: Farid, Abdol Hussain et al.
Attorney Docket No. P05562US00
Title: Insulin-Like Growth Factor-1 Receptor (IGF-1R)
Polymorphic Alleles And Use Of The Same To Identify
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Applicant: Farid, Abdol Hussain et al.
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exon 1
61 ctcgctgtgg gggctcctgt ttttctccgc cgcgctctcg ctctggccga cgagtggaga
21 a atetgeggg cegggcattg acateegcaa egactaceag cagetgaage geetggagaa exon 2 Primer 1
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241 cagetacege titeceaage teaeggteat cacegagtae etgetgetgt teegtgtgge
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081 aggatgcacc atcttcaagg gcaacctgct cattaacatc cgacgcggc acaacattgc
exon 5
141 ctcggagcta gagaacttca tggggctcat cgaggtggtg acgggctacg tgaagatccg
.201 ccactcccat gccttggtct ccttgtcctt cctgaaaaat cttcgccaga tcctagggga
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Primer 9
1681 ggtggacgtg gacctccctc ccaataagga cgtcgagcct ggcattctac tgcatgggct

FIGURE 7A

SNP1762 SNP1772

Primer 7

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1801 tgaccacate cgtggggcca agagtgagat cttgtacatt cgcaccaatg cttca gttcc

Primer 8

1861 ttccattccc ctggatgtcc tctcagcatc aaactcctct tctcagctga tagtgaagtg

- 1921 gaaccegeec teeetgeeca atggeaacet gagetactae ategtgeggt ggeagegeea
- 1981 gcctcaggac agctatetet accggcataa ttactgctcc aaa

Primer 10

ggtgagg gggacatggg acacctgtgg ctctgactcc cgagccctat gctacgcatt cagcatcagg ctgctgctgt Intron 9

gtgcagcctt ggccatggtc acaggtgcta accgYggtgt cYtcccgtac cggttgggta acttggctct SNP9i112 SNP9i119

cettgggetg etetttteea agea |gacaaaa teeccateag exon 10

2041 gaagtatget gatggcacca ttgatgtega ggaggetaca gagaacceca agactgaagt

2101 gtgtggcggg gagaagggac catgctgtgc ttgccccaaa acagaagctg agaagcaggc

2161 agagaaggag gaggctgagt accgcaaggt cttcgagaac ttcctgcaca atgccatctt
Primer 11

2221 tgtgcccaga| cctgaacgga agcggaggga ggtcatgcag gtcgccaaYa ccaccatgtc exon 11 Primer 12 SNP2269

2281 <u>cagccggagc agaaacac</u>ca cagtggtgga cacctacaat gtcacggacc cagaggagct Primer 13

2341 tgagacagag tacccttttt ttgagagcag agtggataac aaggagagaa ctgtcatttc

2401 caacctccgg cettttactt tgtaccgaat tgacatccac agetgtaacc atgaggetga

2461 gaagetggge tgeagtgeet ceaactttgt etttgeaaga accatgeetg calGGTAT

Intron 11

exon 9

GTATGTGAGGTCAGGTCAGGAGAAATGAACGCAGGTACCGTCTGCAGTCC TGTATGCATGGTTGAAGAGGGTATTCTTAGGTTTAA TCCAGCATTTTTTGTCTTTTTTGGGGGGACTTGAGTCTAACTTCATTGCTTGT

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exon 12

- 2521 agatgacatt cetgggecag tgacetggga gecaeggeet gagaacteca tettttaaa
- 2581 gtggccagaa cctgagaatc ctaatggatt gattctaatg tatgaaataa aatacggatc
- 2641 acaagtcgag gatcagcggg aatgtgtgtc cagacaggag tacaggaagt atggaggggc exon 13
- 2701 caagetgaac aggeteaace eggggaacta caeggeeegg atteaggeea cetetetete

Applicant: Farid, Abdol Hussain et al.
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exon 15
2941 gctgtatgct tccgtgaacc cggagtactt cagcgcagct gat gtgtacg tgcccgacga
exon 16
3001 gtgggaggtc gcccgggaga agatcaccat gagccgggag ctgggacagg gctccttcgg
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SNP3085 SNP3088
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3121 cattaagaca gtgaatgagg ccgccagcat gcgYgagagg attgagtttc tcaacgaggc
SNP3154 <u>Primer 15</u>
3181 ctccgtgatg aaggagttca actgtcacca cgtg GTGAGGGAAGGCCCCAAG
Primer 16 Intron 16
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Primer 18 SNP3385 Primer 20 Primer 19
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Primer 22 SNP3832
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Ser Lys Ala Glu Asp Tyr Arg Ser Tyr Arg Phe Pro Lys Leu Thr Val 65 70 75 . 80

Ile Thr Glu Tyr Leu Leu Leu Phe Arg Val Ala Gly Leu Glu Ser Leu 85 90 95

Gly Asp Leu Phe Pro Asn Leu Thr Val Ile Arg Gly Trp Lys Leu Phe 100 105 110

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Lys Gly Val Cys Val Pro Ala Cys Pro Pro Gly Thr Tyr Arg Phe Glu 260 265 270

Gly Trp Arg Cys Val Asp Arg Asp Phe Cys Ala Asn Ile Pro Asn Ala 275 280 285

Glu Ser Ser Asp Ser Asp Gly Phe Val Ile His Asp Asp Glu Cys Met 290 295 300

Gln Glu Cys Pro Ser Gly Phe Ile Arg Asn Ser Thr Gln Ser Met Tyr 305 310 315

Cys Ile Pro Cys Glu Gly Pro Cys Pro Lys Val Cys Gly Asp Glu Glu 325 330 335

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Phe	Ala 450	Phe	Asn	Pro	Lys	Leu 455	Cys	Val	Ser	Glu	Ile 460	Tyr	Arg	Met	Gl u
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Arg	Asn	Asn	Gly	Glu 485	Arg	Ala	Ser	Cys	Glu 490	Ser	Asp	Val	Leu	Arg 495	Phe
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Tyr	Arg	Pro 515	Pro	Asp	Tyr	Arg	Asp 520		Ile	Ser	Phe	Thr 525	Val	Tyr	Tyr
Lys	Glu 530	Ala	Pro	Phe	Ъуз	Asn 535	Val	Thr	Glu	Tyr	Asp 540	Gly	Gln	Asp	Ala
Cys 545	Gly	Ser	Asn	Ser	Trp 550	Asn	Met	Val	Asp	Val 555	Asp	Leu	Pro	Pro	Asn 560
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Asp	His	Ile 595	Arg	Gly	Ala	Lys	Ser 600	Glu	Ile	Leu	Tyr	Ile 605	Arg	Thr	Asn
Ala	Ser 610	Val	Pro	Ser	Ile	Pro 615	Leu	qaA	Val	Leu	Ser 620	Ala	Ser	Asn	Ser
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- Lys Tyr Ala Asp Gly Thr Ile Asp Val Glu Glu Val Thr Glu Asn Pro 675 680 685
- Lys Thr Glu Val Cys Gly Gly Asp Lys Gly Pro Cys Cys Ala Cys Pro 690 695 700
- Lys Thr Glu Ala Glu Lys Gln Ala Glu Lys Glu Glu Ala Glu Tyr Arg 705 710 715 720
- Lys Val Phe Glu Asn Phe Leu His Asn Ser Ile Phe Val Pro Arg Pro 725 730 735
- Glu Arg Arg Arg Asp Val Met Gln Val Ala Asn Thr Thr Met Ser 740 745 750
- Ser Arg Ser Arg Asn Thr Thr Val Ala Asp Thr Tyr Asn Ile Thr Asp 755 760 765
- Pro Glu Glu Phe Glu Thr Glu Tyr Pro Phe Phe Glu Ser Arg Val Asp 770 775 780
- Asn Lys Glu Arg Thr Val Ile Ser Asn Leu Arg Pro Phe Thr Leu Tyr 785 790 795 800
- Arg Ile Asp Ile His Ser Cys Asn His Glu Ala Glu Lys Leu Gly Cys 805 810 815
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- Val Ser Arg Gln Glu Tyr Arg Lys Tyr Gly Gly Ala Lys Leu Asn Arg 885 890 895
- Leu Asn Pro Gly Asn Tyr Thr Ala Arg Ile Gln Ala Thr Ser Leu Ser 900 905 910
- Gly Asn Gly Ser Trp Thr Asp Pro Val Phe Phe Tyr Val Pro Ala Lys 915 920 925
- Thr Thr Tyr Glu Asn Phe Met His Leu Ile Ile Ala Leu Pro Val Ala 930 935 940
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- Val Asn Pro Glu Tyr Phe Ser Ala Ala Asp Val Tyr Val Pro Asp Glu 980 985 990
- Trp Glu Val Ala Arg Glu Lys Ile Thr Met Asn Arg Glu Leu Gly Gln
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1085	1090	1095	

Val Glu Gln Asn Asn Leu Val Leu Ile Pro Pro Ser Leu Ser Lys 1105 ·

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- Asn Ala Asn Lys Phe Val His Arg Asp Leu Ala Ala Arg Asn Cys
- Met Val Ala Glu Asp Phe Thr Val Lys Ile Gly Asp Phe Gly Met
- Thr Arg Asp Ile Tyr Glu Thr Asp Tyr Tyr Arg Lys Gly Gly Lys
- Gly Leu Leu Pro Val Arg Trp Met Ser Pro Glu Ser Leu Lys Asp
- Gly Val Phe Thr Thr His Ser Asp Val Trp Ser Phe Gly Val Val
- Leu Trp Glu Ile Ala Thr Leu Ala Glu Gln Pro Tyr Gln Gly Leu
- Ser Asn Glu Gln Val Leu Arg Phe Val Met Glu Gly Gly Leu Leu
- Asp Lys Pro Asp Asn Cys Pro Asp Met Leu Phe Glu Leu Met Arg
- Met Cys Trp Gln Tyr Asn Pro Lys Met Arg Pro Ser Phe Leu Glu
- Ile Ile Gly Ser Ile Lys Asp Glu Met Glu Pro Ser Phe Gln Glu
- Val Ser Phe Tyr Tyr Ser Glu Glu Asn Lys Pro Pro Glu Pro Glu
- Glu Leu Glu Met Glu Leu Glu Met Glu Pro Glu Asn Met Glu Ser

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Glu Arg His Ser Gly His Lys Ala Glu Asn Gly Pro Gly Pro Gly 1325 1330 1335

Val Leu Val Leu Arg Ala Ser Phe Asp Glu Arg Gln Pro Tyr Ala 1340 1345 1350

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Glu Ser Val Pro Leu Asp Pro Ser Ala Ser Ser Ala Ser Leu Pro Leu 65 70 75 80

Pro Glu Arg His Ser Gly His Lys Ala Glu Asn Gly Pro Gly Pro Gly 85 90 95

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